



Facioscapulohumeral Dystrophy

Key Opinion Leader Breakfast Forum

8:30 a.m. – 10:30 a.m.

November 7, 2019

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This presentation also contains estimates and other statistical data made by independent parties and by us relating to market size and other data about our industry. This data involves a number of assumptions and limitations, and you are cautioned not to give undue weight to such data and estimates. In addition, projections, assumptions and estimates of our future performance and the future performance of the markets in which we operate are necessarily subject to a high degree of uncertainty and risk. Neither Fulcrum nor its affiliates, advisors or representatives makes any representation as to the accuracy or completeness of that data or undertakes to update such data after the date of this presentation.

Agenda

8:30am Welcome – **Robert J. Gould, Ph.D.**, *President and Chief Executive Officer, Fulcrum Therapeutics*

8:40am Introduction to FSHD – **Kathryn Wagner, M.D., Ph.D.**, *Professor of Neurology and Neuroscience at Johns Hopkins School of Medicine and Director of the Center for Genetic Muscle Disorders Kennedy Krieger Institute*

9:00am Biologic rationale and genetics – **Peter Jones, Ph.D.**, *Mick Hitchcock, Ph.D. Endowed Chair in Medical Biochemistry and Associate Professor of Pharmacology at University of Nevada, Reno School of Medicine*

9:15am The role of DUX4 - **Peter Jones, Ph.D.**

9:30am Overview of p38i – **Fran Sverdrup, Ph.D.**, *Associate Professor of Biochemistry and Molecular Biology at Saint Louis University, School of Medicine*

9:50am Imaging and biopsy as an approach & clinical trial design implications – **Kathryn Wagner, M.D., Ph.D.**

10:10am Q&A ***Please hold questions until the Q&A*

Fulcrum at a glance

Proprietary Product Engine



Fulcrum's product engine is designed to systematically address the root cause of many genetically defined diseases

Our Progress to Date

Developed proprietary drug discovery platform

Established patient-driven discovery & development process

Ongoing **Phase 2 studies** in Facioscapulohumeral Muscular Dystrophy (FSHD)

Initiated FTX-6058 **IND-enabling studies** for select hemoglobinopathies (Sickle Cell Disease and β -thalassemia)

Demonstrated systematic target and therapeutic discovery potential

Integrated FSHD development strategy

Natural history & preparatory studies

Ongoing

Assessing disease progression and validating clinical endpoints

Phase 1

Enrollment complete – analysis ongoing

Demonstrated losmapimod target engagement, muscle penetration, and safety in FSHD patients

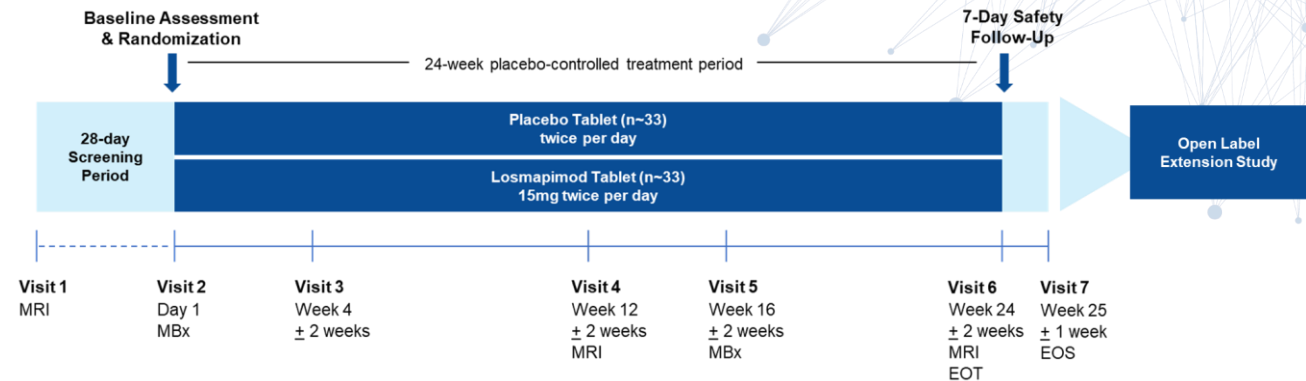
Phase 2 Open Label Study (52 weeks) with interim analyses

Ongoing

Assessing molecular endpoint (DUX4 muscle biopsy), MRI disease measurement, clinical assessment of mobility, and PROs

Phase 2b (ReDUX4) 24 weeks dosing

Open Label Extension



Day 1 & Week 16: Muscle Biopsy (MBx)

DUX4-driven gene expression in skeletal muscle needle biopsy

Visit 1, Week 12, Week 24: MRI

lean skeletal muscle volume; skeletal muscle fat fraction

Day 1, Weeks 4, 12, 16, 24: Clinical assessments

PK; safety; Reachable Work Space; FSHD-Timed Up & GO, Muscle function measures, dynamometry and Patient Reported Outcomes

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Introduction to FSHD

Kathryn Wagner, MD, PhD

Center for Genetic Muscle Disorders

Kennedy Krieger Institute

Johns Hopkins School of Medicine

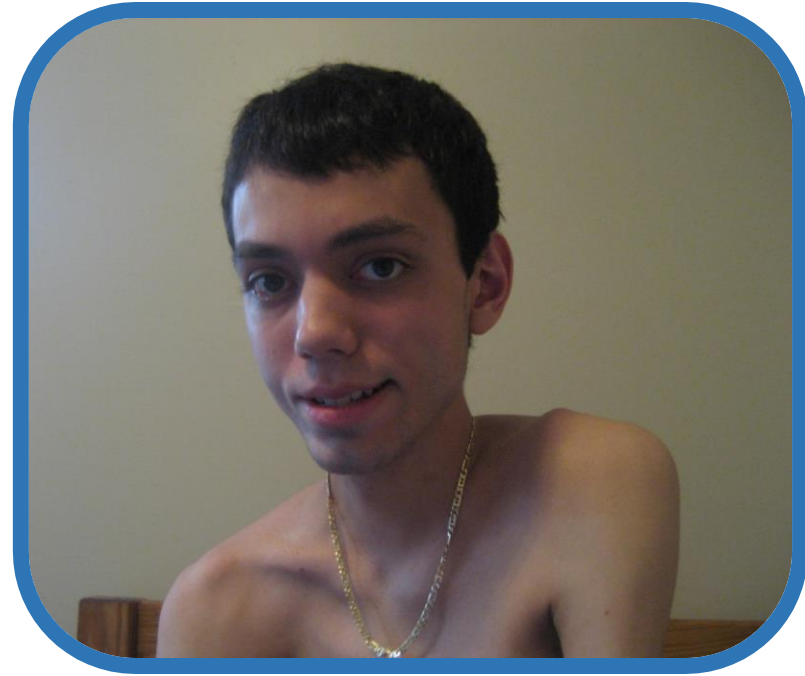
Facioscapulohumeral muscular dystrophy (FSHD)

- Prevalence of ~1:8,000 - 1:20,000
- Autosomal dominant disorder of families
- Extremely disabling but not fatal



Where it gets its name

facio= face



**scapulo= muscles of scapular
fixation**

**humeral= muscles overlying
humerus**



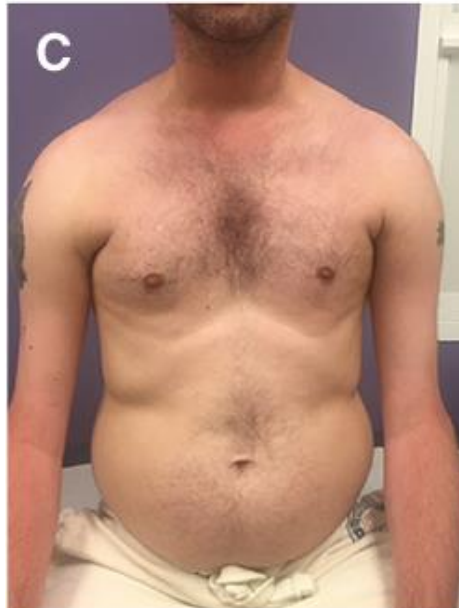
Clinical Presentation:

- Symptoms first noticed in muscles of face and scapular region
- Patients present with wide open eyes and have a history of sleeping with their eyes partially open
- They have an inability to pucker and may never learn to whistle
- They frequently have a transverse or asymmetric smile



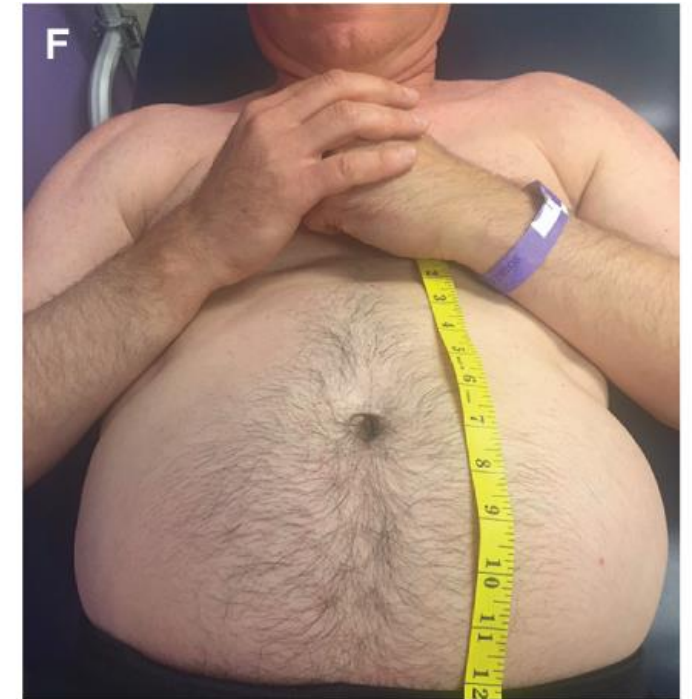
Clinical Presentation continued

- Muscles of scapular fixation (rhomboids and serratus anterior) are weak
 - Medial border of scapula “wings”
 - Rostral border rises up: Poly-hill sign on arm abduction
 - Inability to slowly abduct or extend arms to 180 degrees
- Pectoralis weakness
 - Horizontal clavicles
 - Deep axillary creases



Clinical Presentation continued

- Biceps and Triceps are disproportionately involved compared to deltoid and forearm flexors
- Weakness of tibialis anterior results in footdrop
- Paraspinal and abdominal weakness leads to lordosis and protruberant abdomen.
- Umbilicus moves rostrally when the individual attempts to sit up: Beevor sign
- Eventually forearm flexors and extensors, knee flexors and extensors may become weak
- Weakness and wasting are frequently asymmetric



Clinical Presentation

- Two forms: FSHD1 and FSHD2 present similarly
- Onset and severity vary widely
- Most classic onset is teenage or early adult years
- Range from infantile onset to nonmanifesting carriers
- Correlation to size of allele

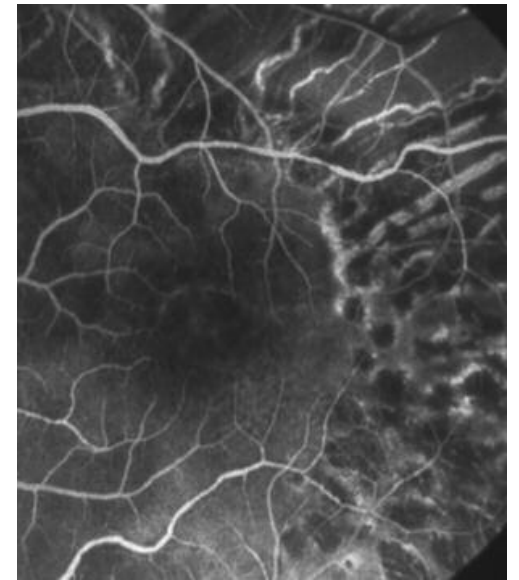


How does FSHD affect day to day life?

- Inability to communicate via facial expression
- Inability to do activities requiring upper arms including brushing hair, putting dishes on a shelf, shampooing
- Difficulty getting out of bed
- Tripping and falling
- 30% lose ambulation
- Chronic pain and fatigue

Associated Symptoms

- 30% of FSHD who are nonambulatory have respiratory involvement
- Cardiomyopathy not associated with FSHD
 - Conduction defects or arrhythmias might be more prevalent
- Retinal vasculopathy
 - 50% have mild retinal abnormalities such as telangiectasias or microaneurysms
 - 0.8% have vasculopathy with neovascularization, retinal detachment, neovascular glaucoma
 - Severe retinal vasculopathy associated with early-onset FSHD and large 4q35 deletions
- Hearing loss
 - High frequency hearing loss more common in early-onset FSHD
- Musculoskeletal pain
 - 88.6% of patients reported current pain



Diagnostic Evaluation

- Clinical presentation fairly distinct from other myopathies
- CK will be normal to mildly elevated
- EMG shows nonspecific myopathic features (small, polyphasic motor units) and occasional irritability (fibrillations and positive sharp waves)
- Muscle biopsy rarely indicated: Nonspecific myopathic features
- Genetic testing is commercially available, sensitive and specific

Current Management

- No accepted pharmacological treatments for progressive muscle weakness: Challenge managing the disease
- Dilated eye examination to r/o reversible retinal vascular disease
- All patients with early-onset FSHD screened for hearing loss
- Pulmonary function testing at baseline and annually for those with severe weakness, kyphoscoliosis, wheelchair dependence
 - Referral to sleep specialist when FVC<60% or excessive daytime somnolence, frequent nocturnal arousals or morning headaches
- Pain management
 - PT
 - NSAIDs
 - Antidepressants

Current Management continued

- Surgical scapular fixation
 - When can't abduct to 90 degrees but good deltoid preservation
- Bracing
 - Ankle foot orthoses
- Exercise
 - Cycling 30 min/day, 3X/wk
- Annual DEXA
 - treatment of low bone density



Conclusions

- FSHD is an autosomal dominant disorder that preferentially affects face, muscles of scapular fixation and arms
- Eventually, most skeletal muscles are affected
- FSHD is associated with rare extramuscular manifestations including retinal vasculopathy and hearing loss
- There are no treatments for the muscle weakness associated with FSHD

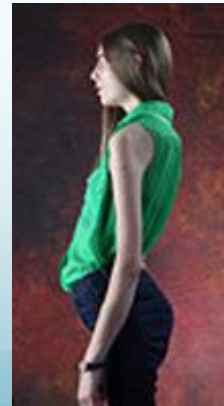


University of Nevada, Reno
School of Medicine
Department of Pharmacology



FSHD Genetics and Epigenetics

Peter L. Jones, Ph.D. and Takako I. Jones, Ph.D.
Co-Principal Investigators



Epigenetics

“Treasure your exceptions.”

William Bateson “The Methods and Scope of Genetics” 1908



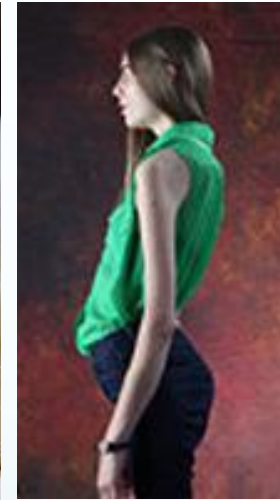
PEV



In utero diet

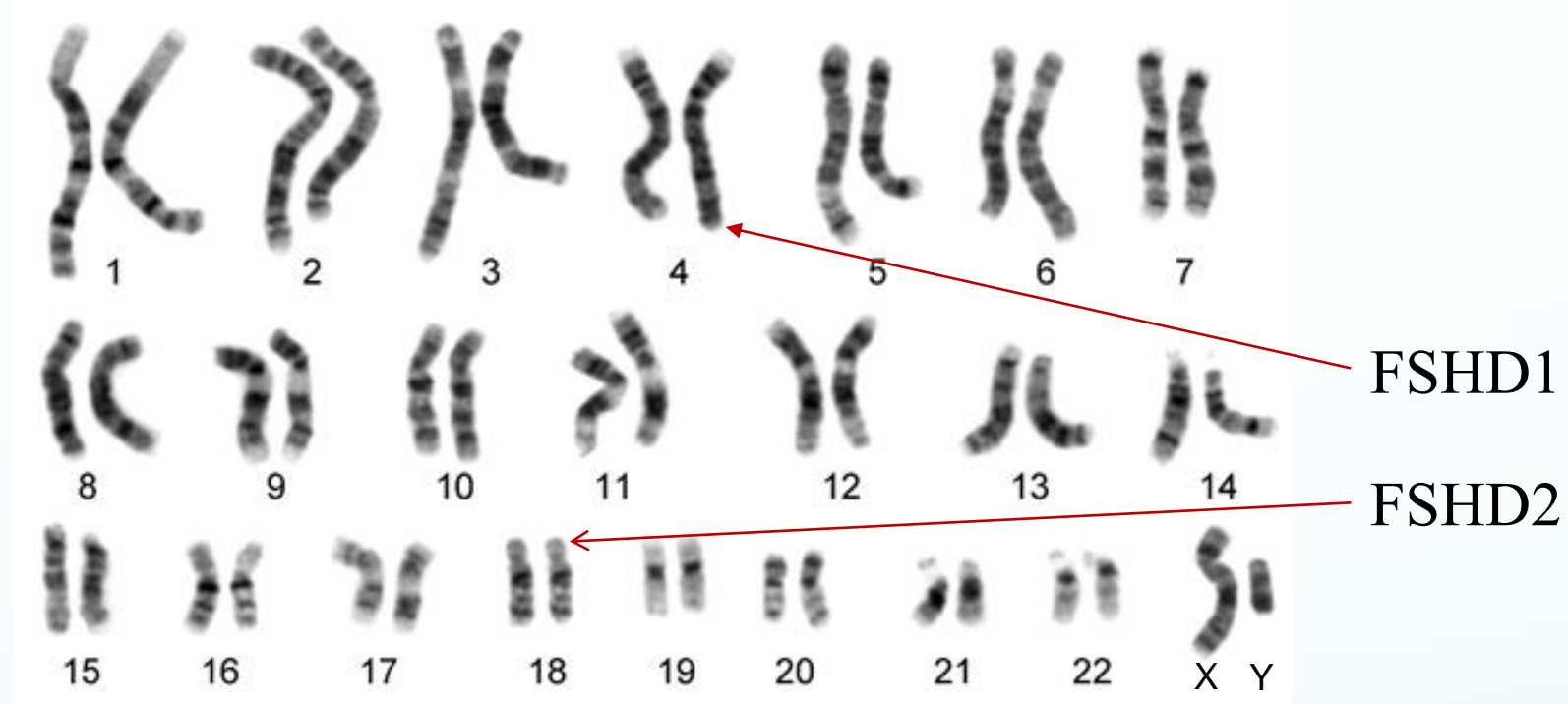


X-inactivation



FSHD

FSHD is caused by genetic changes that lead to epigenetic changes at Chr 4q35



Human haploid genome has ~3,100,000,000 base pairs of DNA (GATCs)

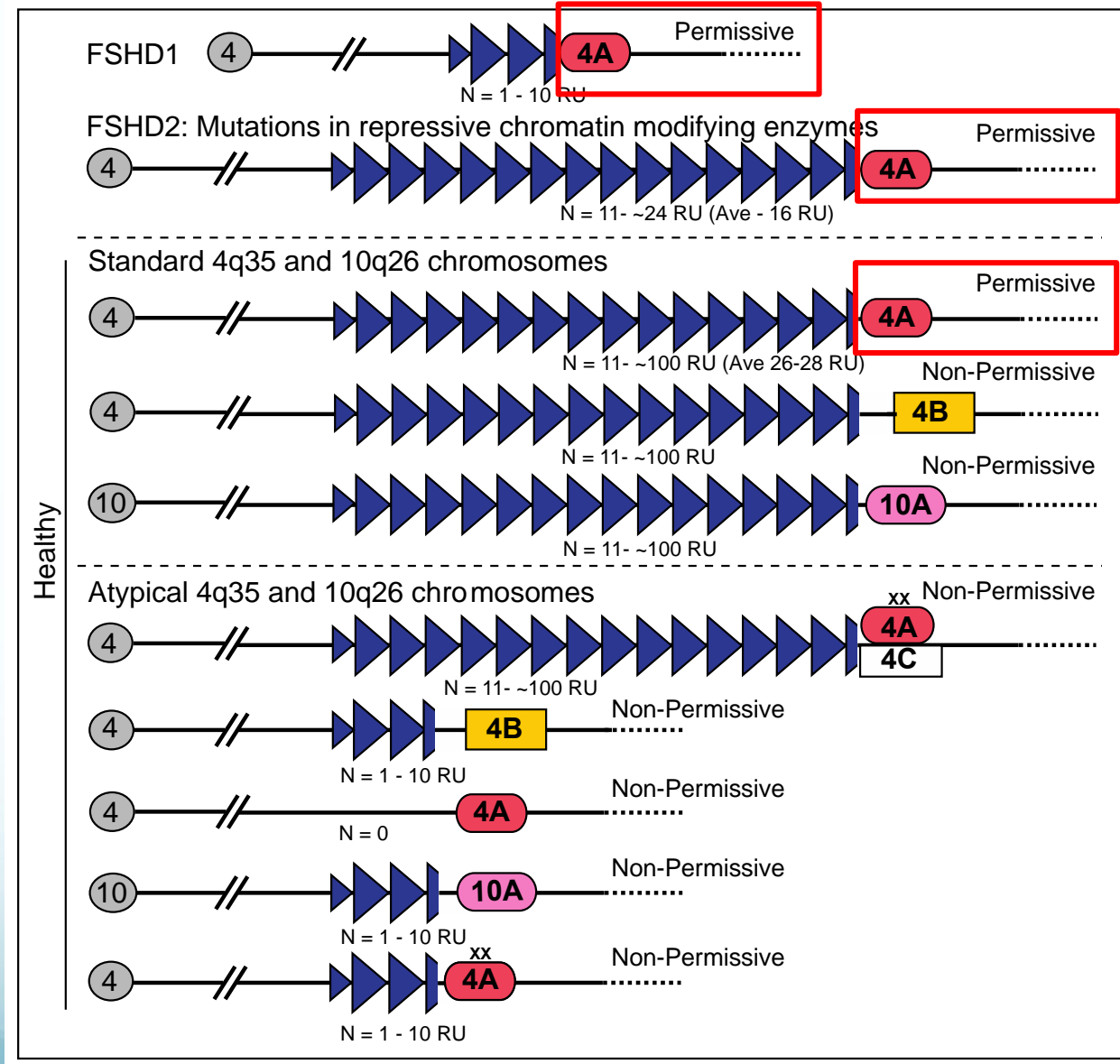
FSHD1 is caused by small deletions on Chr 4q

→ lead to epigenetic changes at Chr 4q

FSHD2 is usually caused by mutations on Chr 18p

→ lead to epigenetic changes at Chr 4q

FSHD genetics are complex

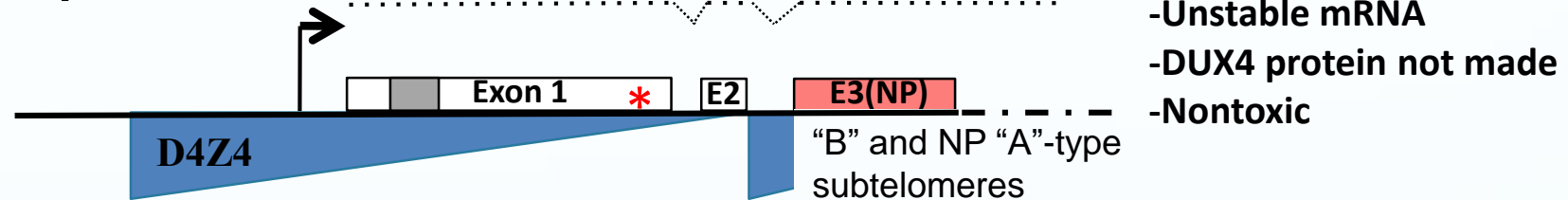


A Unifying Genetic Model for Facioscapulohumeral Muscular Dystrophy

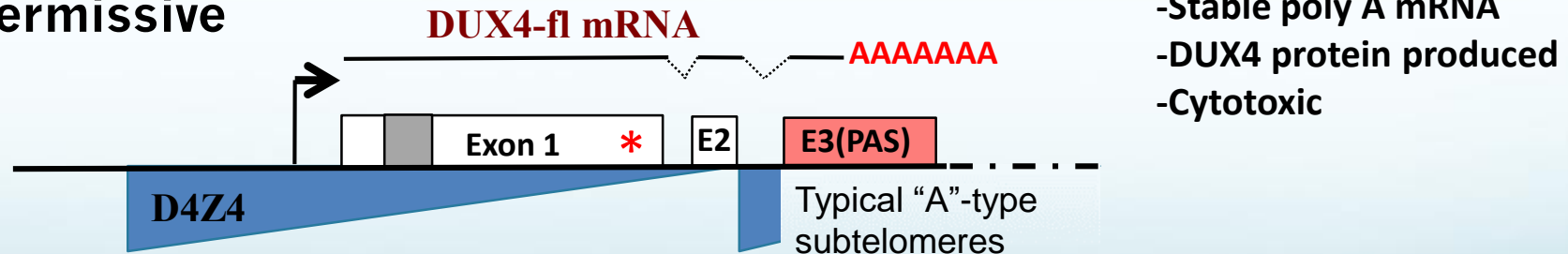
Richard J. L. F. Lemmers,¹ Patrick J. van der Vliet,¹ Rinse Klooster,¹ Sabrina Sacconi,² Pilar Camaño,^{3,4} Johannes G. Dauwerse,¹ Lauren Snider,⁵ Kirsten R. Straasheijm,¹ Gert Jan van Ommen,¹ George W. Padberg,⁶ Daniel G. Miller,⁷ Stephen J. Tapscott,⁵ Rabi Tawil,⁸ Rune R. Frants,¹ Silvere M. van der Maarel^{1*}

DUX4 genetics link all forms of FSHD

Non-permissive

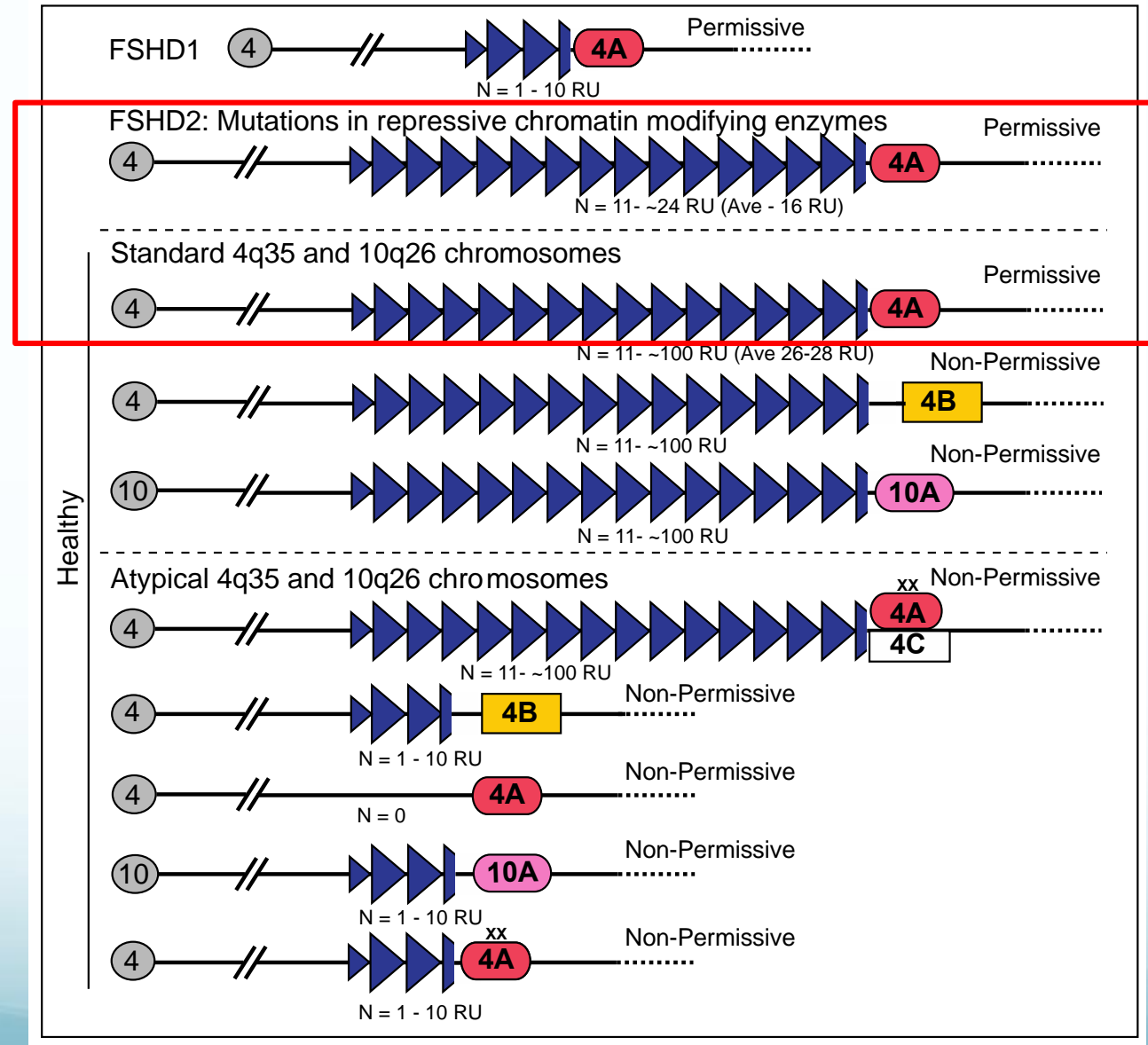


Permissive

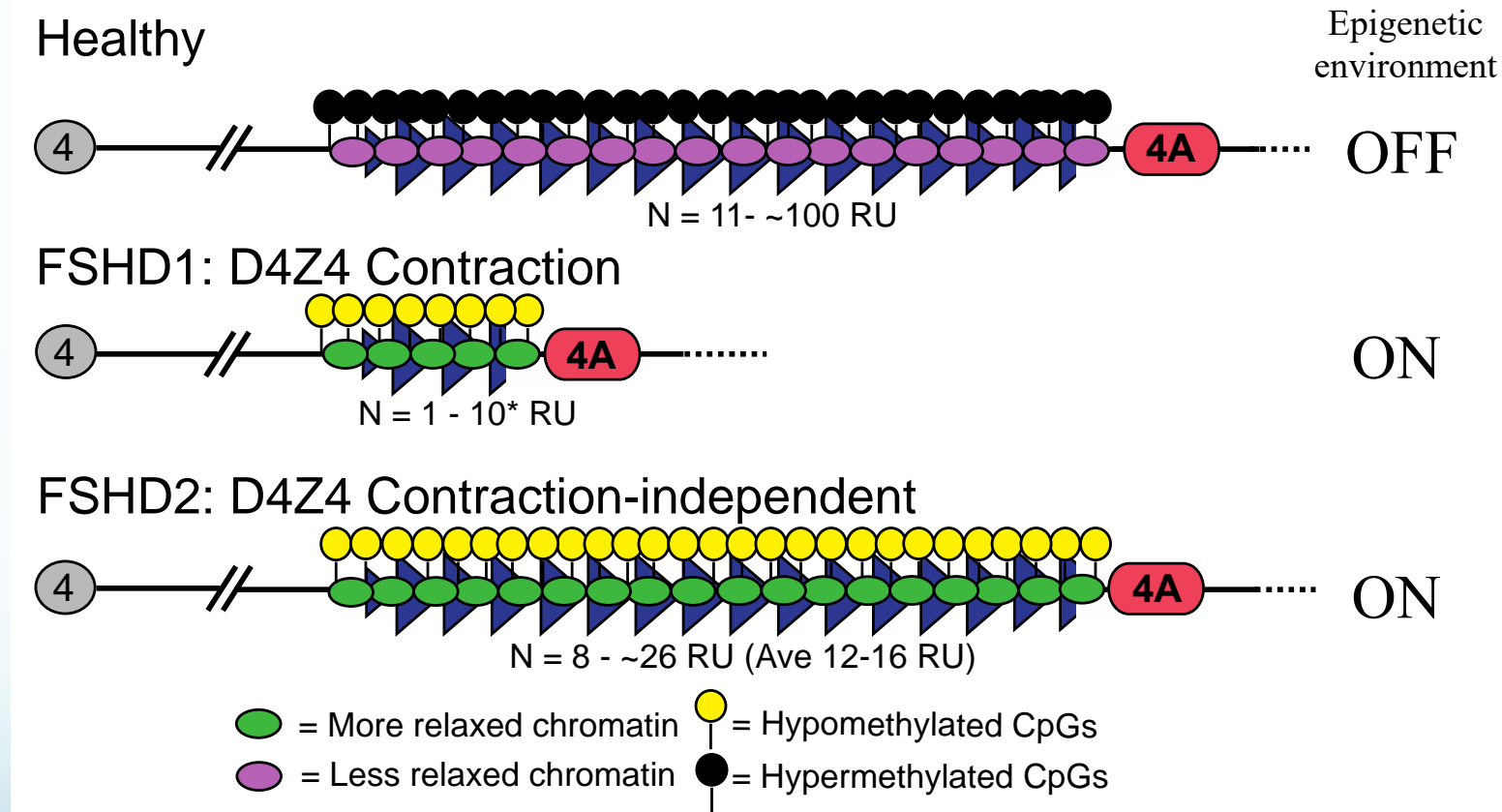


* = translation stop PAS = DUX4 mRNA polyadenylation site NP = non-permissive

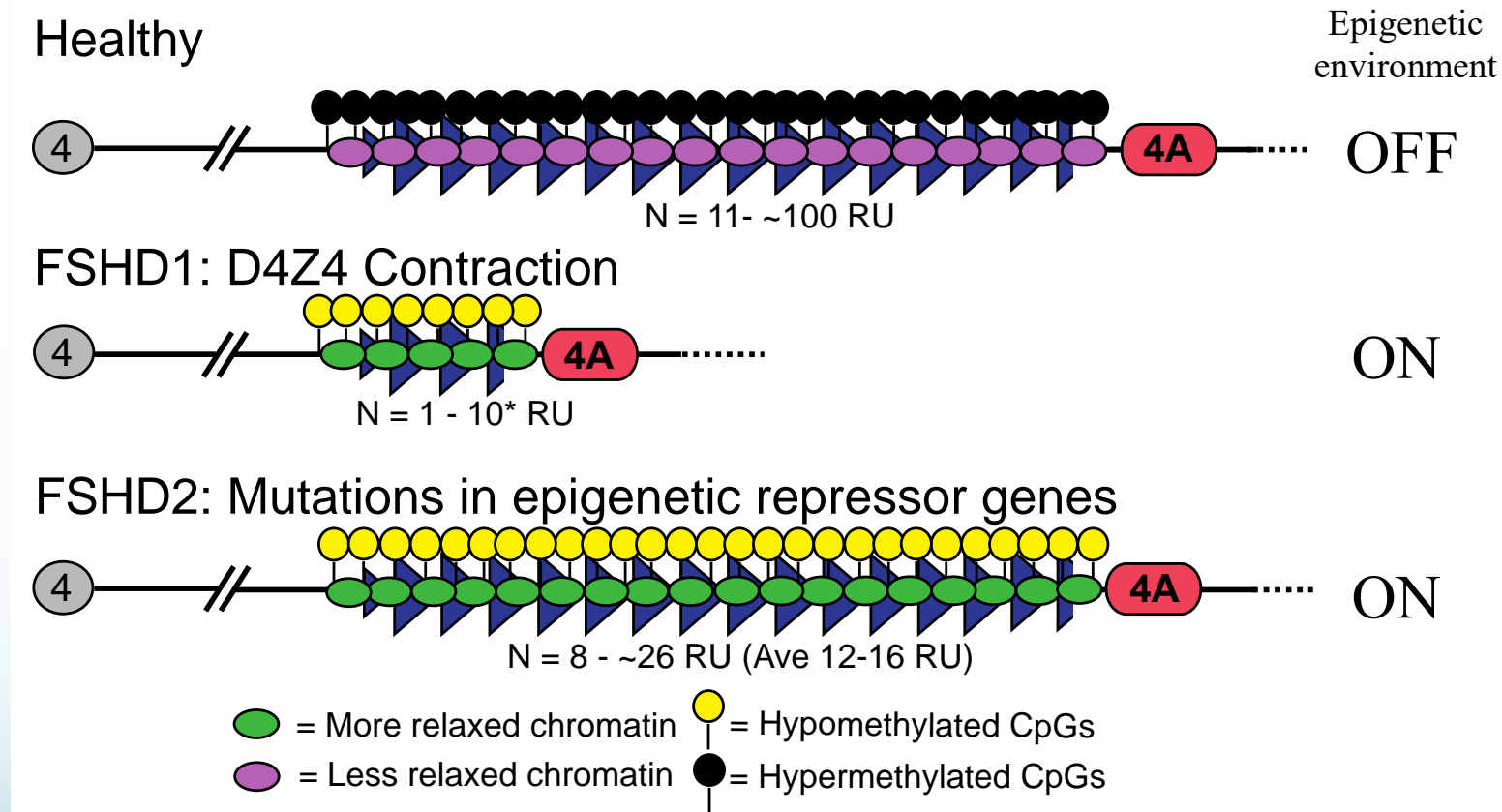
FSHD genetics are complex



Epigenetic dysregulation links all forms of FSHD

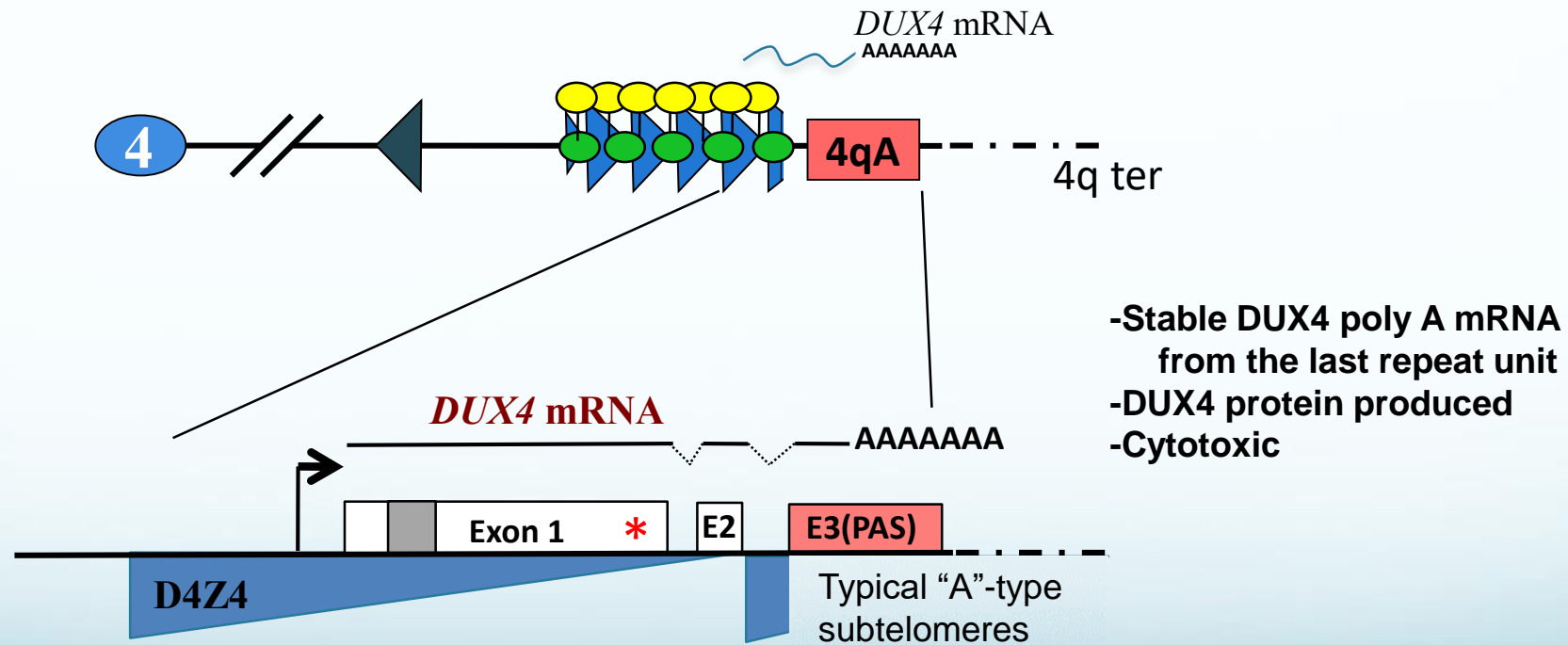


FSHD is an epigenetic disease with a genetic component



**What are the consequences
of dysregulated epigenetics
in FSHD?**

Aberrant epigenetics combined with DUX4-permissive genetics leads to expression of the *DUX4* gene resulting in FSHD

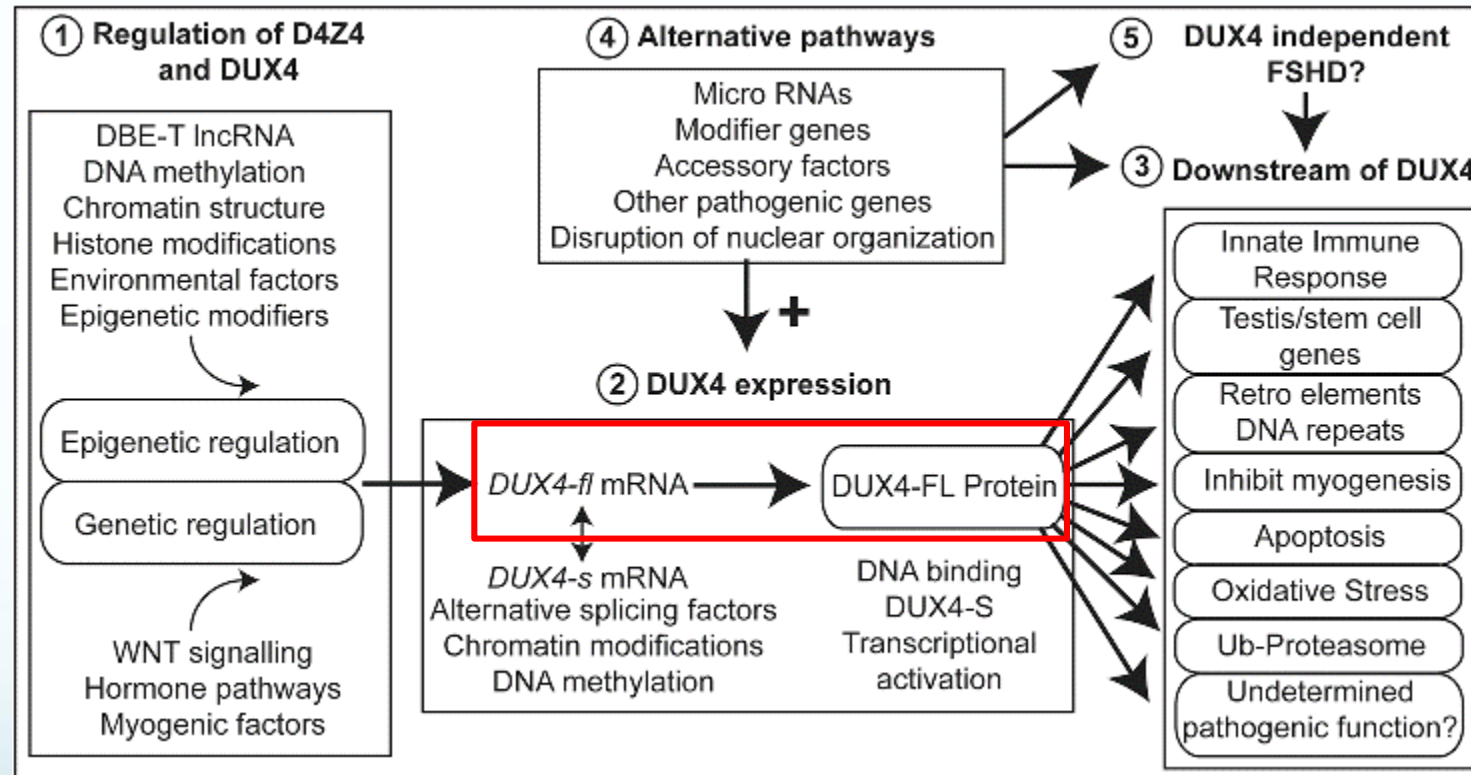


* = translation stop PAS = DUX4 mRNA polyadenylation site

***DUX4* encodes an early
developmentally active
transcription factor that is
silent in healthy somatic cells**

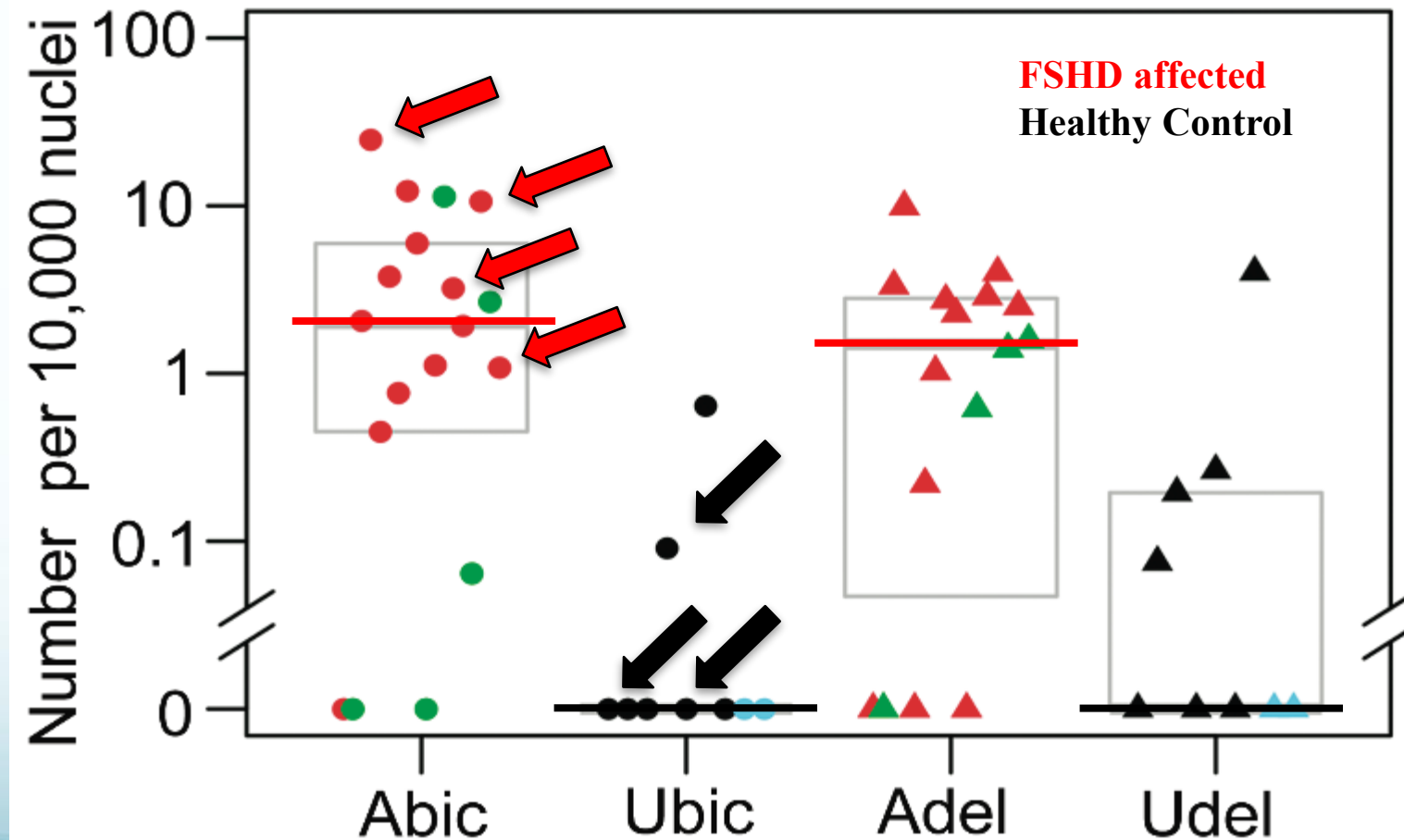
***DUX4* expression is
aberrantly increased in FSHD
skeletal muscle**

Pathogenic mechanisms of FSHD are dependent upon DUX4



Quantitative model of DUX4-FL expression

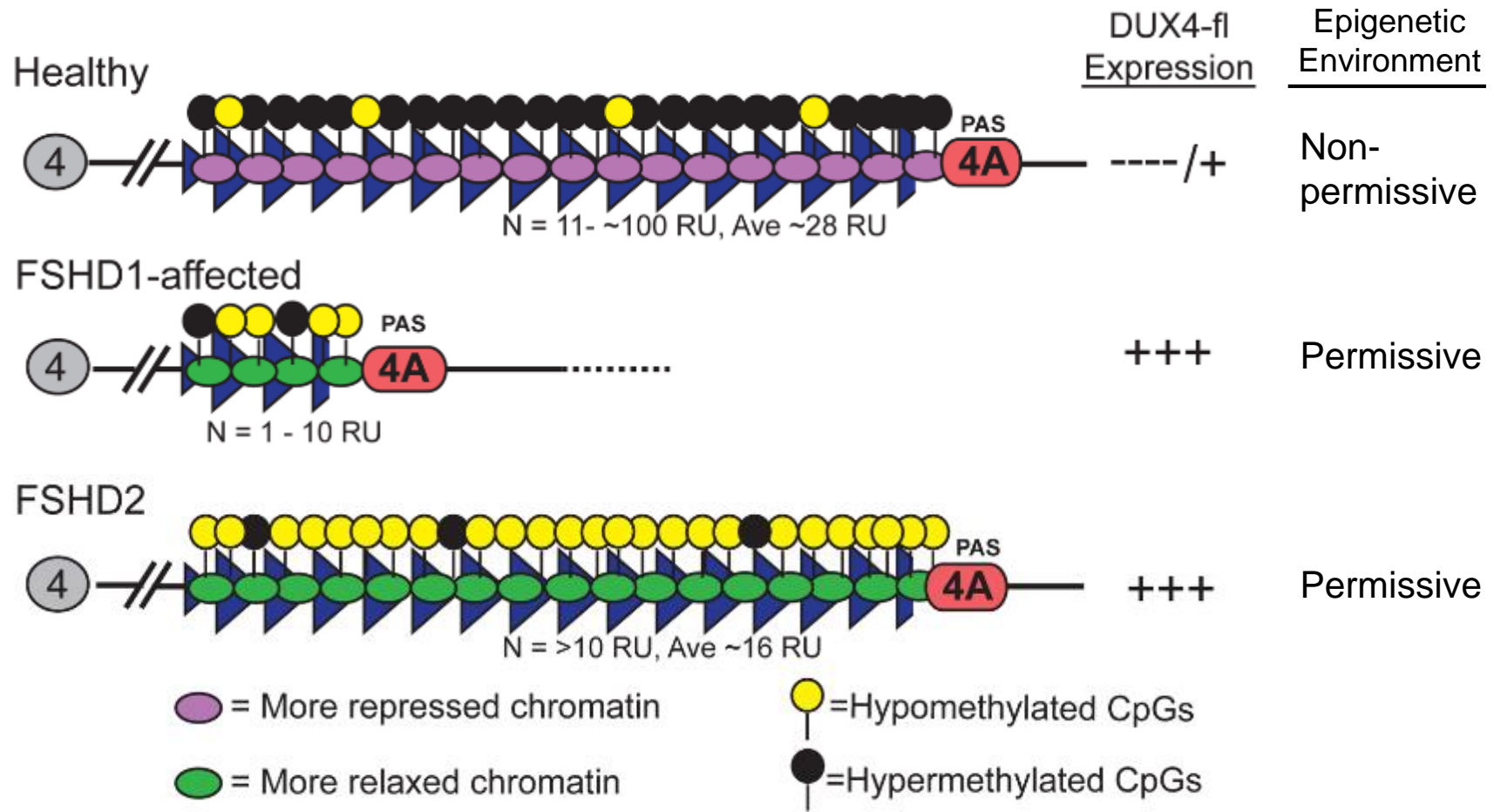
DUX4-FL expression in <1% of nuclei of myogenic cells from FSHD subjects



Aberrantly increased expression of the *DUX4* gene is the key pathogenic mechanism in FSHD

- FSHD is a pathogenic gain-of-function disease
- Most muscle diseases are loss of function
→ need to “fix” or “replace” something
- FSHD → need to remove an unwanted protein

FSHD is an epigenetic disease with a genetic component

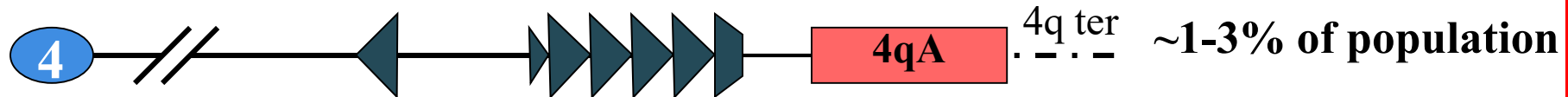


FSHD1 genetics fail to account for the large number of asymptomatic individuals

Healthy



Asymptomatic: Genetically FSHD1, clinically unaffected



Scionti *et al.* (2012) *J Med Genet* 49:171

Ricci *et al.* (2013) *Brain* 136:3408

FSHD1



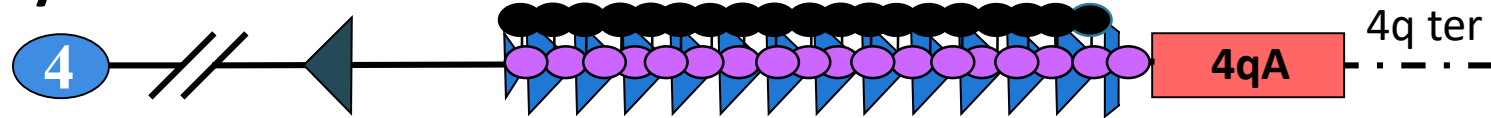
The deletion itself is not pathogenic

The 4qA sub-telomere is permissive, not pathogenic

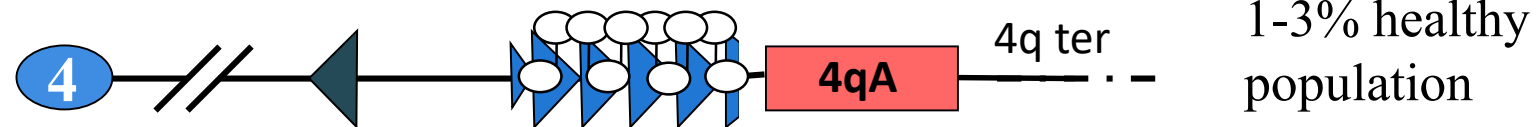
Existence of modifiers of disease severity

What is the epigenetic state of asymptomatic FSHD subjects?

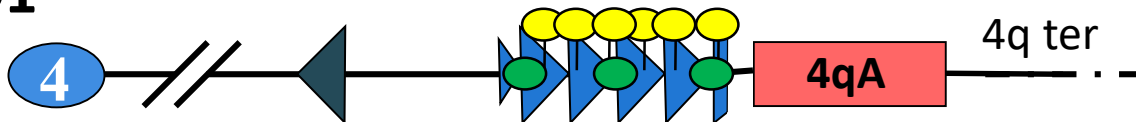
Healthy



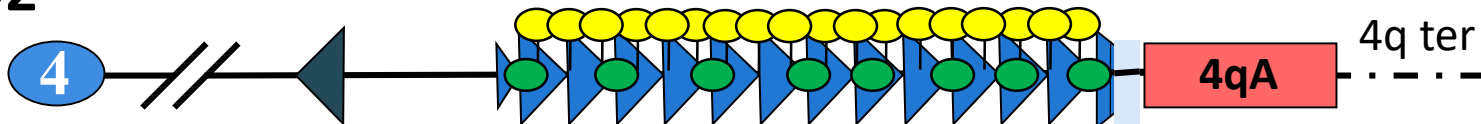
FSHD1 asymptomatic



FSHD1



FSHD2



● = Hypermethylated CpGs

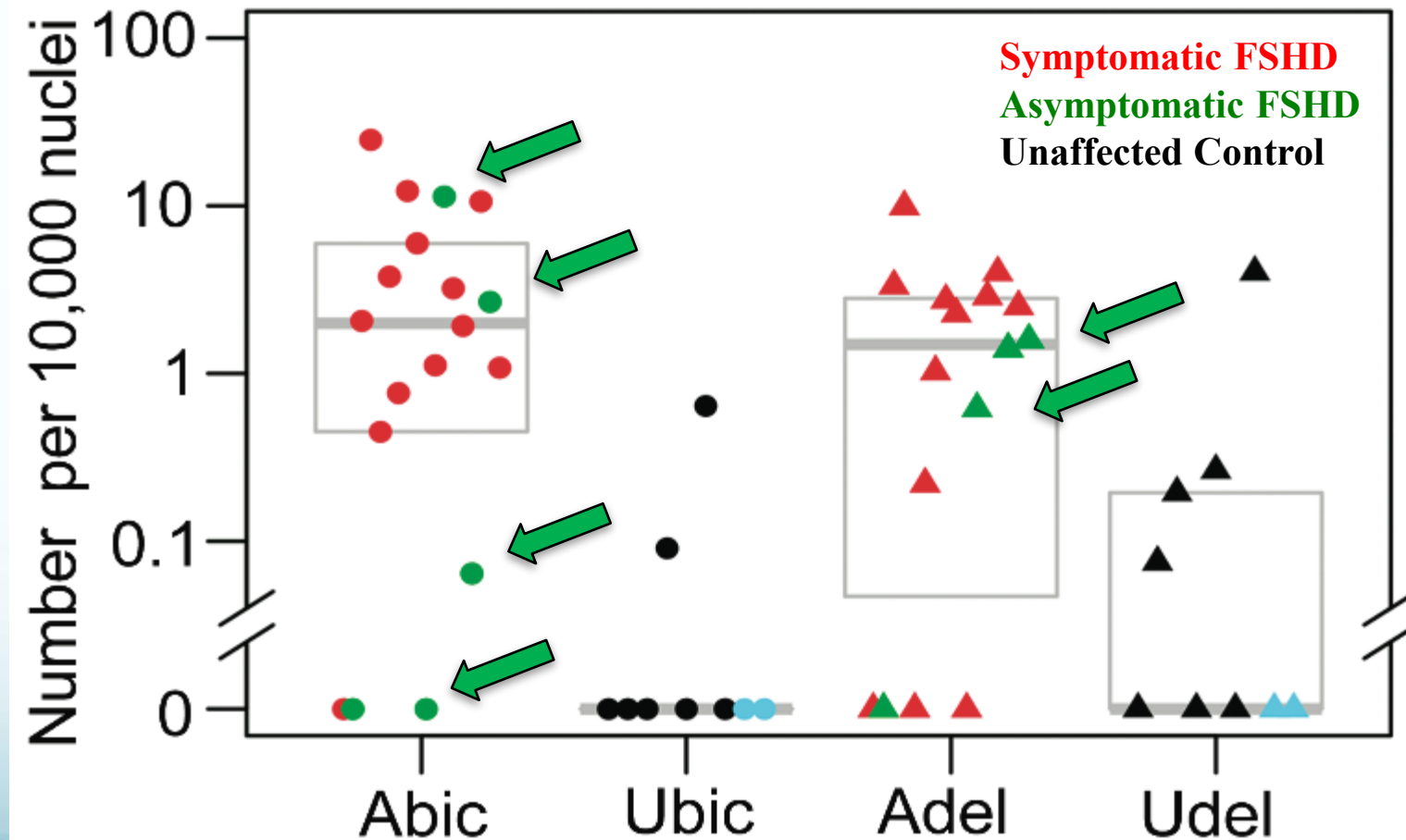
● = More heterochromatic

● = Hypomethylated CpGs

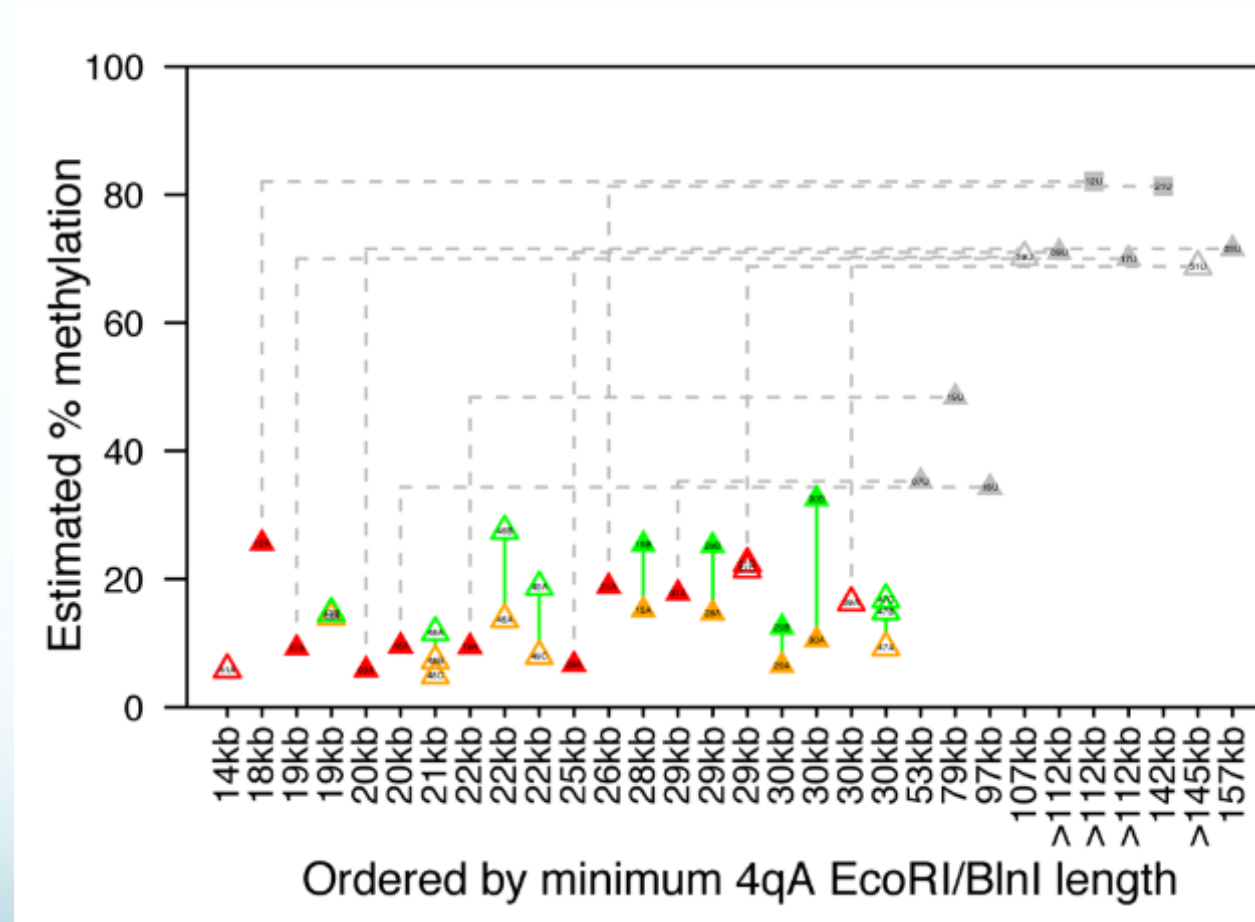
● = More euchromatic

Quantitative model of DUX4-FL expression

DUX4-FL expression in <1% of nuclei of myogenic cells from FSHD subjects



Asymptomatic FSHD1 subjects have an intermediate level of DNA methylation

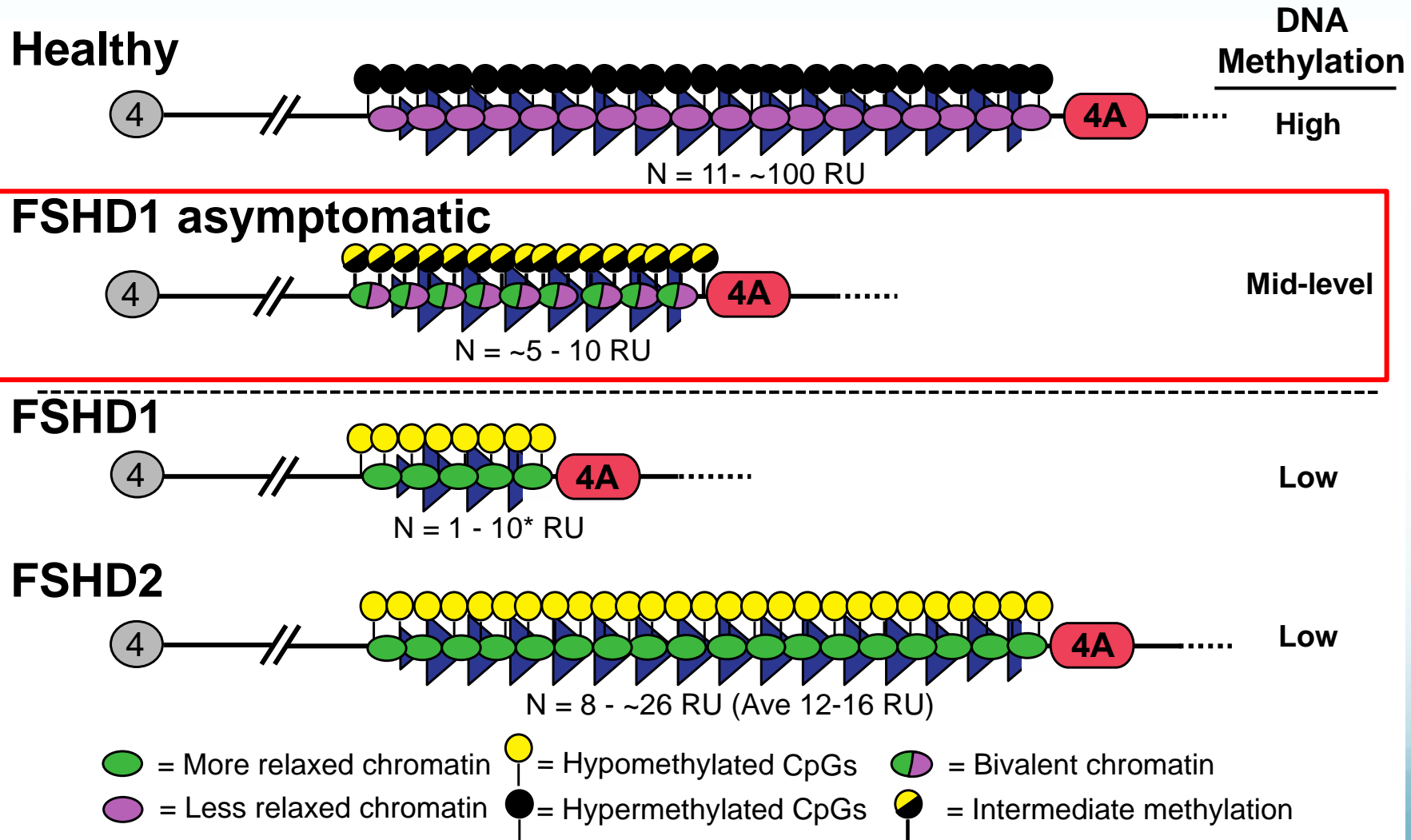


Asymptomatic FSHD1 subjects have an intermediate level of DNA methylation that is significantly higher than FSHD1-affected and significantly lower than healthy controls

| Cohort | Manifesting | Nonmanifesting | <i>EcoRI/BlnI</i> | D4Z4 RU* |
|--------|-------------|----------------|-------------------|----------|
| 15 | 15.2% | 25.4% | 28kb | 8 |
| 28 | 14.6% | 25.2% | 29kb | 8 |
| 29 | 6.5% | 12.5% | 30kb | 8.5 |
| 30 | 10.6% | 32.6% | 30kb | 8.5 |
| 43 | 14.2% | 15.5% | 19kb | 5 |
| 46 | 13.7% | 27.6% | 22kb | 6 |
| 47 | 9.3% | 14.9% & 16.9% | 30kb | 8.5 |
| 48 | 7.3% & 4.9% | 11.7% | 21kb | 6 |
| 49 | 8.0% | 18.8% | 22kb | 6 |

*Calculated as $D4Z4\ RU = (EcoRI/BlnI\ fragment\ kb - 2kb) / 3.3$

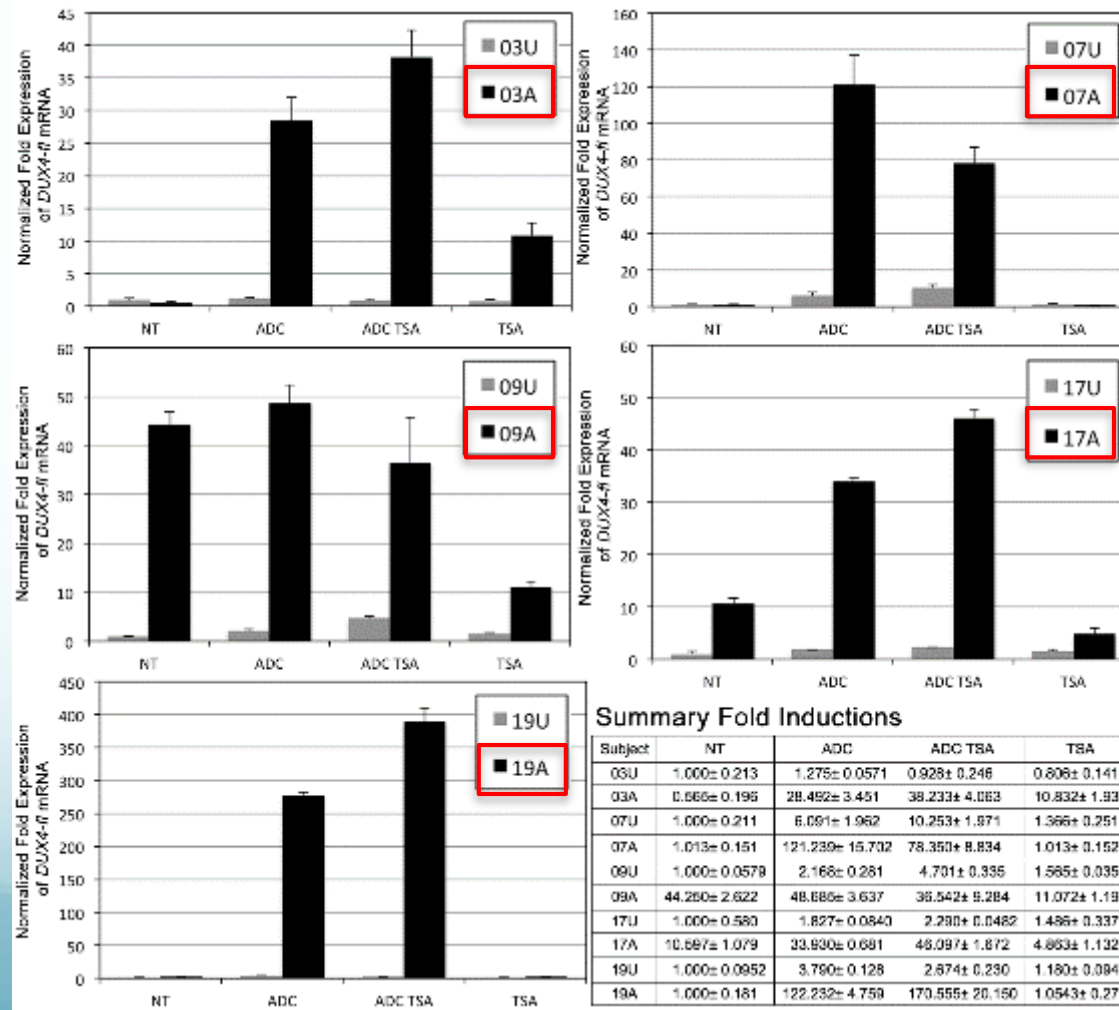
Epigenetic status correlates with FSHD disease presentation



**What are the consequences of
epigenetic dysregulation?**

Do these small differences matter?

Epigenetic repression at the 4q35 D4Z4 array is very stable in healthy controls and epigenetically poised for expression in FSHD1

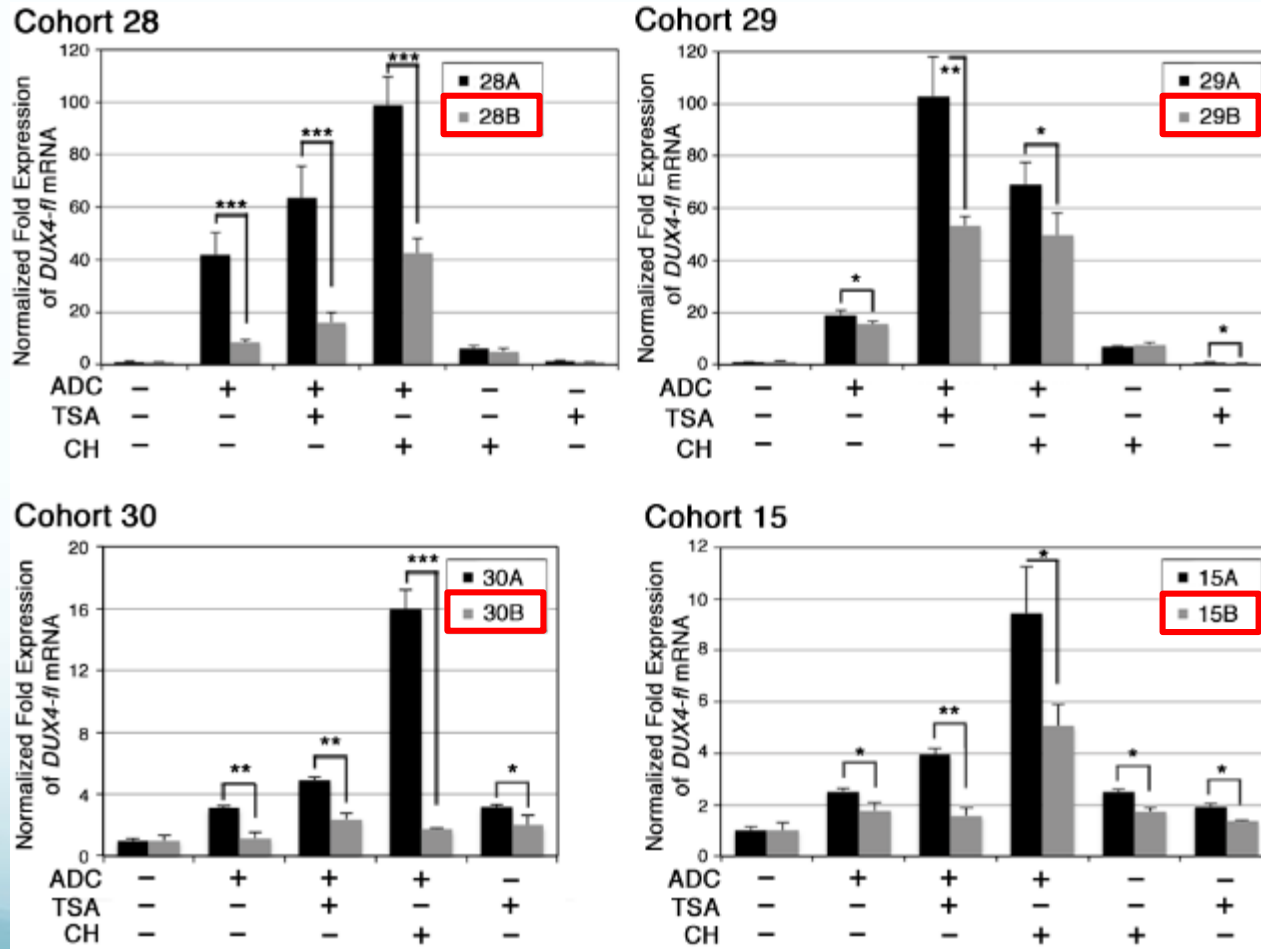


NT = not treated

ADC = Decitabine
inhibits DNA methylation
→ more euchromatic

TSA = Trichostatin A
HDAC inhibitor
→ favors histone acetylation
→ more euchromatic
→ can affect other proteins

Epigenetic repression at the 4q35 D4Z4 array is more stable in asymptomatic FSHD1 subjects compared with affected FSHD1 patients



NT = not treated

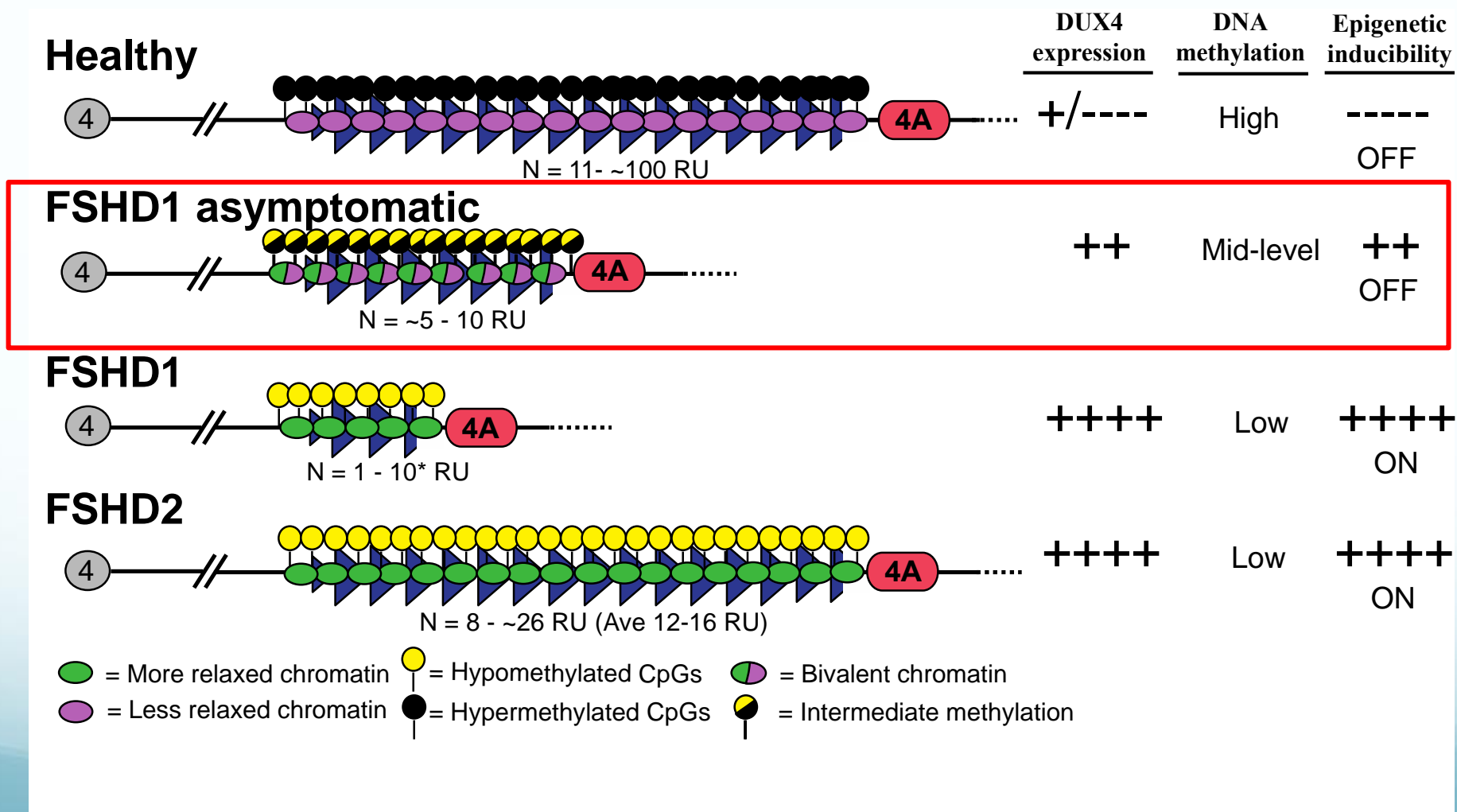
ADC = Decitabine
 → inhibits DNA methylation
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 HDAC inhibitor
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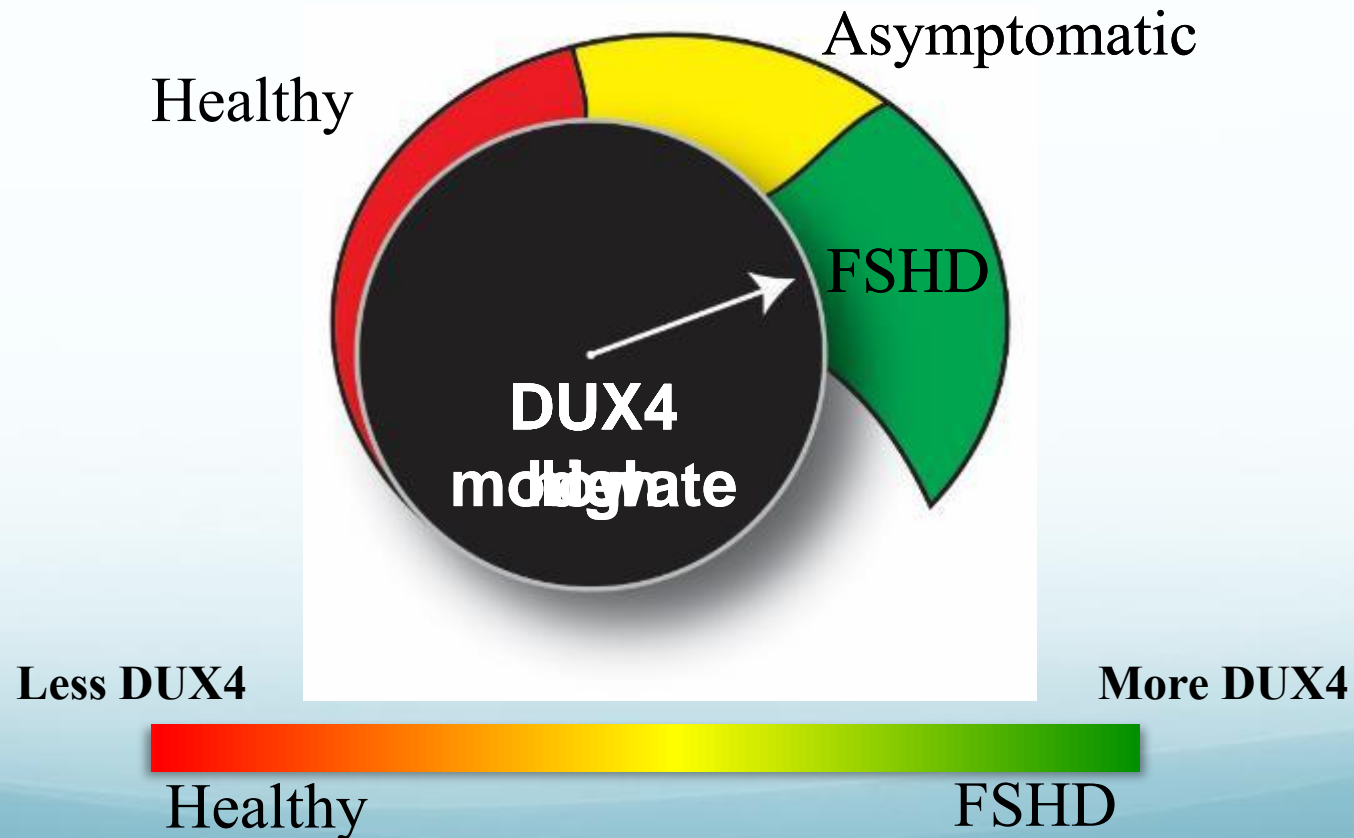
CH = Chaetocin
 → inhibits H3K9me3
 → more euchromatic

B = Non-manifesting

Asymptomatic FSHD1 subjects have an intermediate status

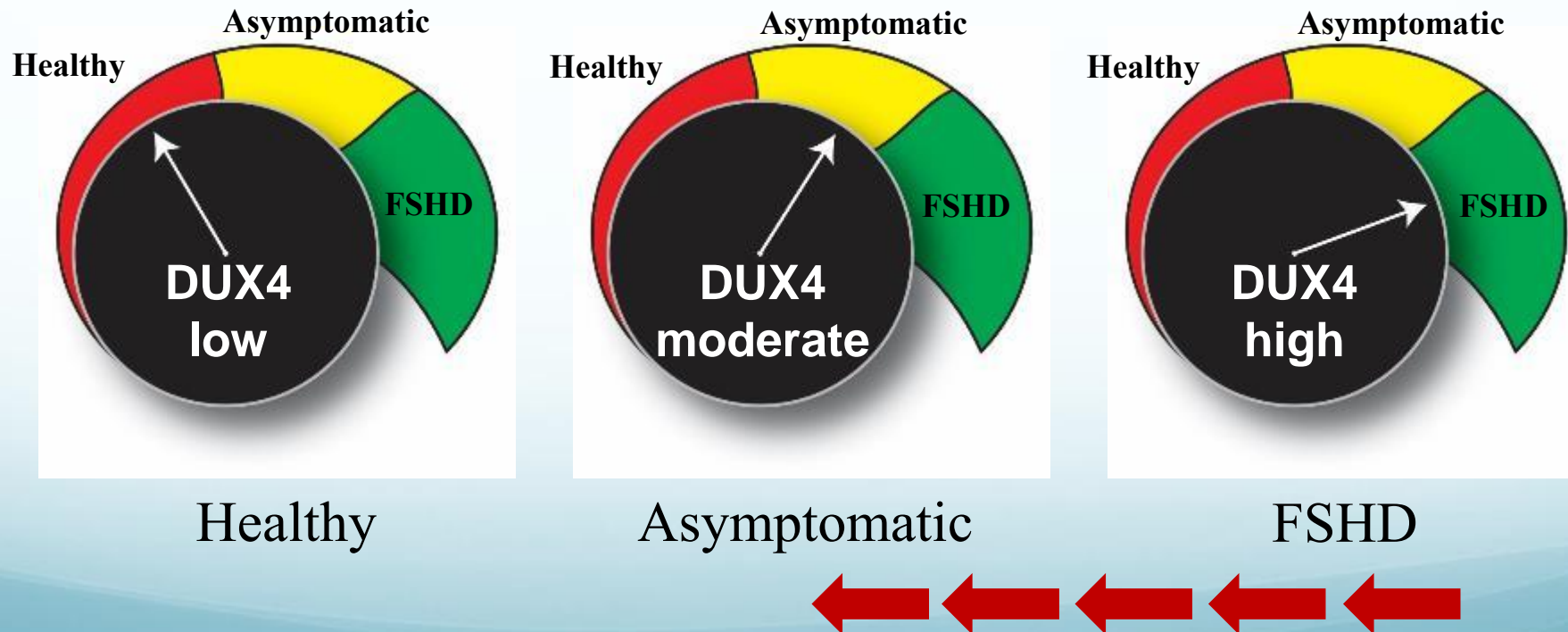


Small changes in epigenetic state and/or DUX4 expression levels have large clinical consequences



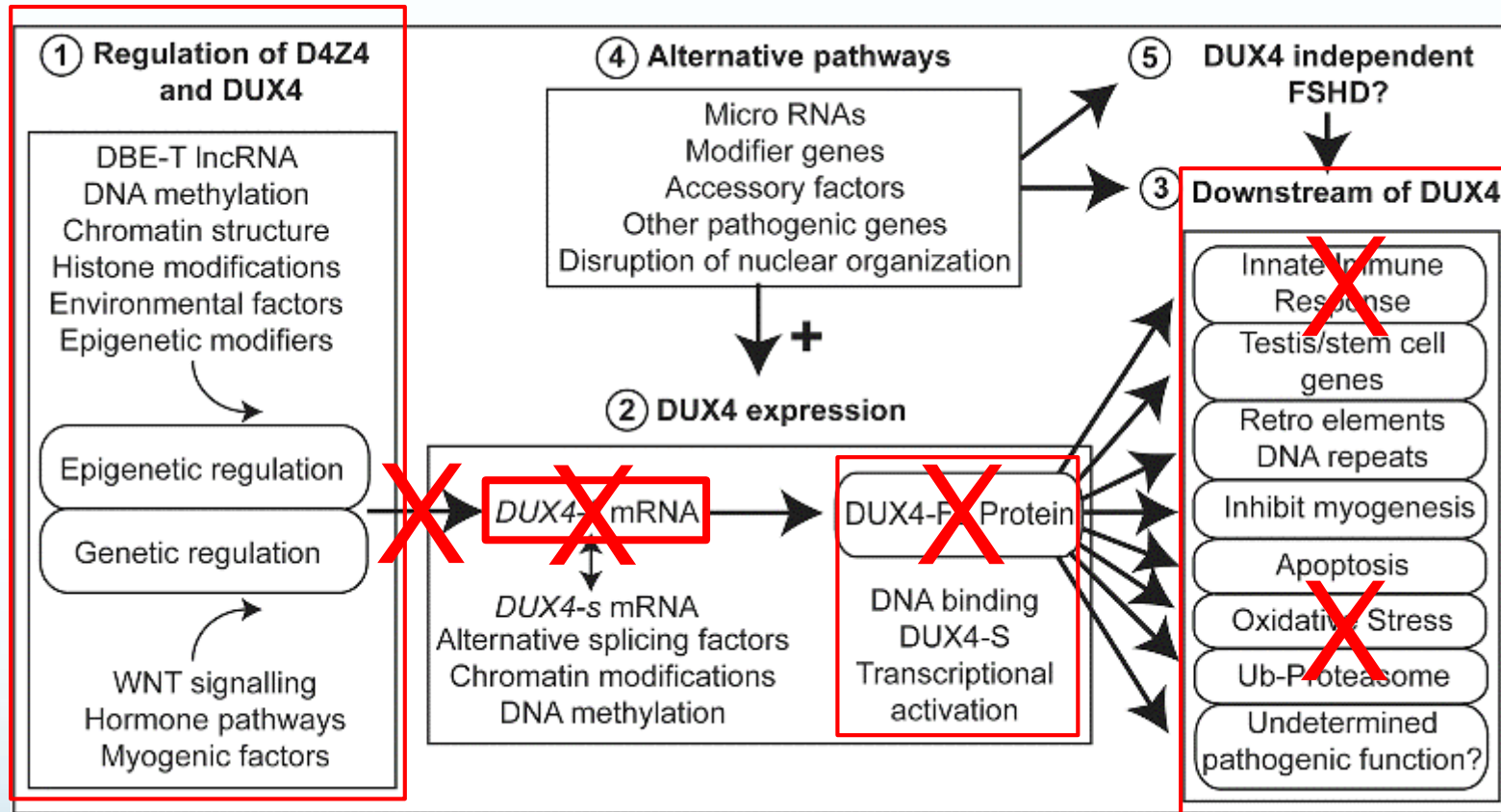
**What does the epigenetic and
DUX4 expression data tell us
about FSHD therapies?**

Small changes in epigenetic state and/or DUX4 expression levels have large clinical consequences



FSHD in 2019

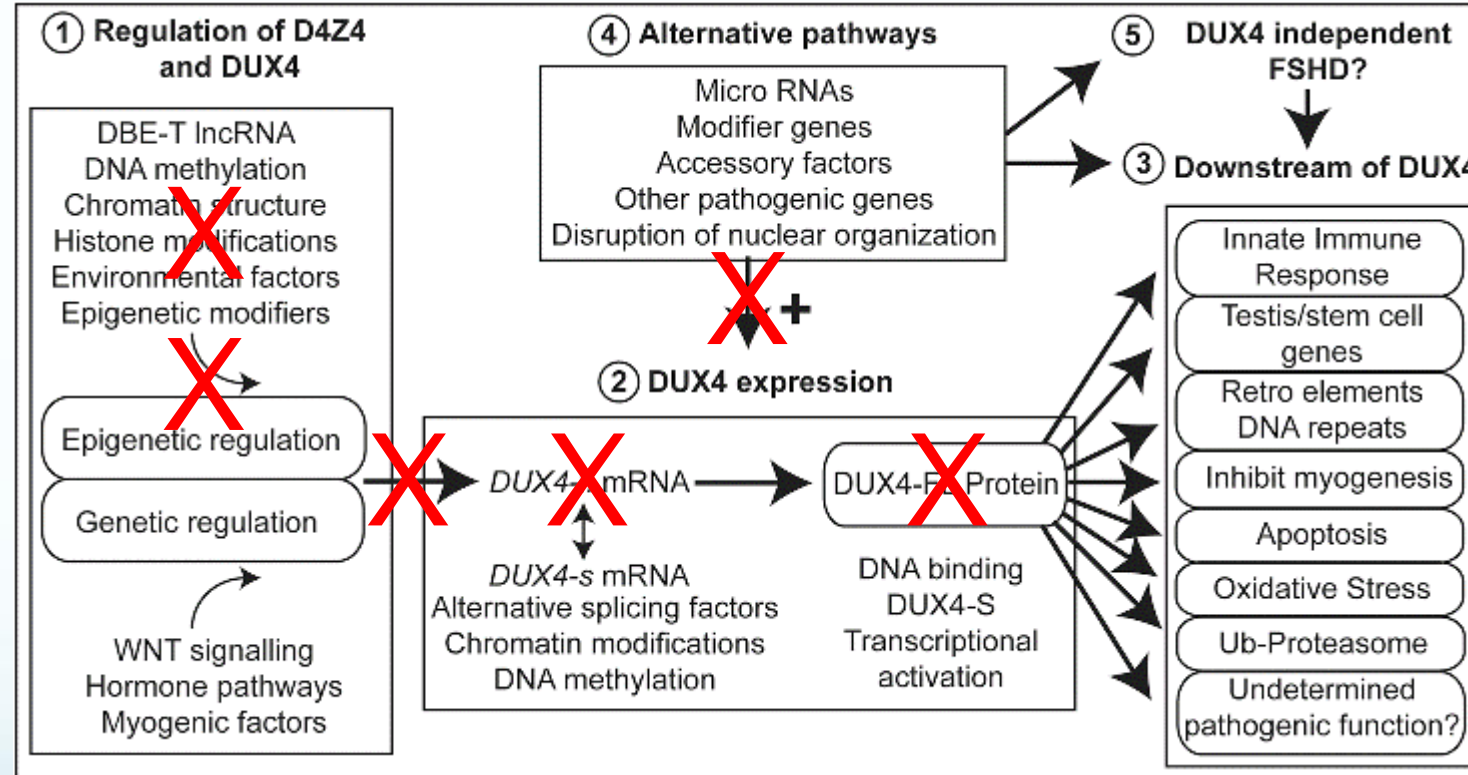
Many viable therapeutic approaches



Anti-DUX4 tools (DUX4 inhibitors, siRNA, DUX4 protein RNAi)
 Drugs blocking DUX4 protein function (Icagen)
 (e.g. MyoStatin, MyoStatin, MyoStatin, MyoStatin, MyoStatin)

Reduction or elimination of DUX4 expression targets the key pathogenic mechanism

Any level of DUX4 reduction may have therapeutic benefit!



Small molecules targeting DUX4 regulation or function

***Morpholinos/PMOs/shRNAs & *miRNAs**

***CRISPR-inhibition**

***delivery concerns**

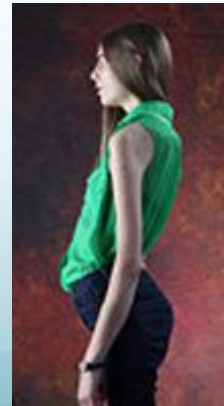


University of Nevada, Reno
School of Medicine
Department of Pharmacology



The Role of DUX4 in Development and Disease

Peter L. Jones, Ph.D. and Takako I. Jones, Ph.D.
Co-Principal Investigators



We now have a clear therapeutic target: DUX4

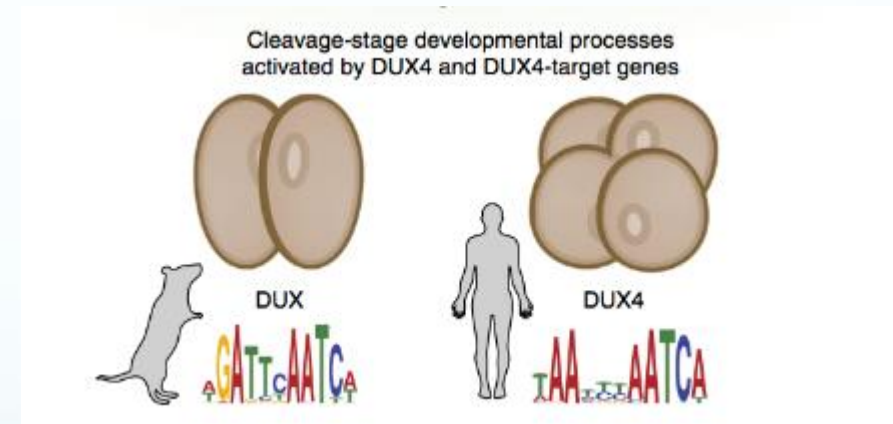
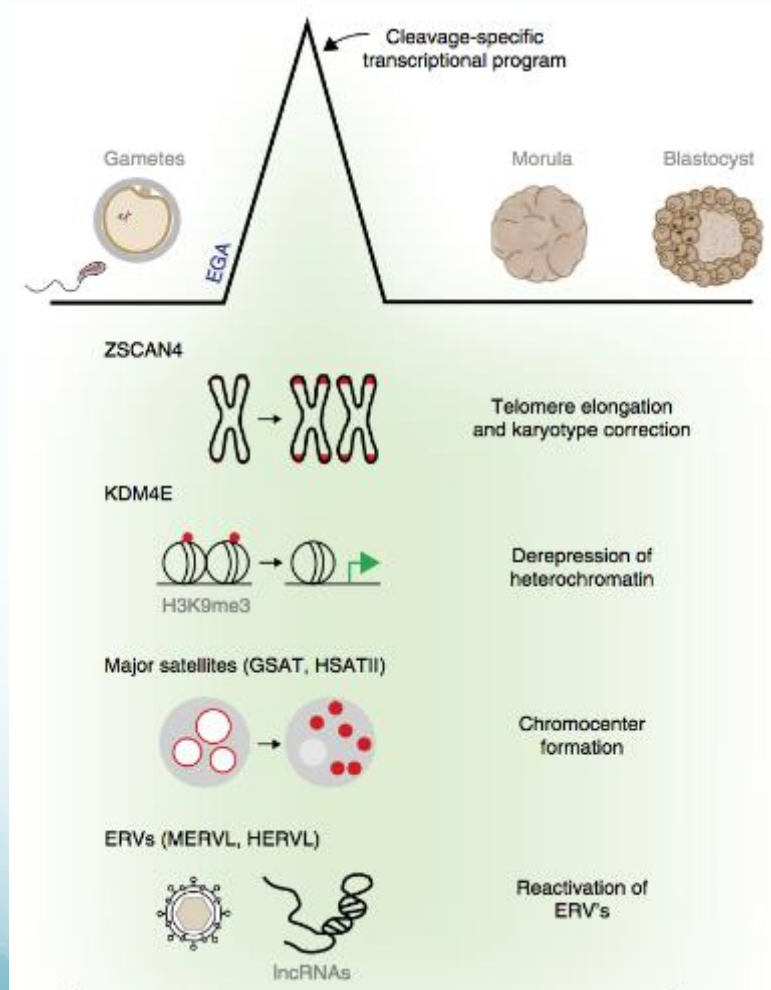


Produced by Bill Milling and Susan Egert
Directed by Arie Ohayon
Featuring Steven Blier and Kelli O'hara

DUX4 expression is a FSHD therapeutic target

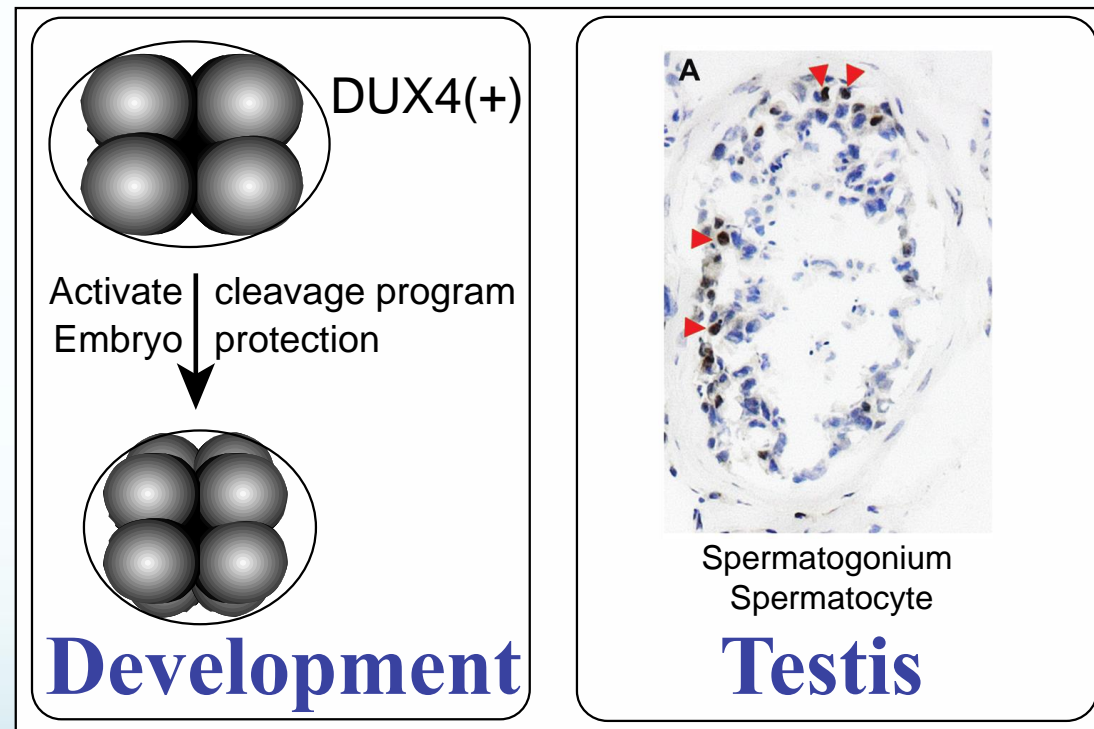
- **Why do we have DUX4 and what does it do?**
- **How does it cause FSHD?**
- **What will reducing expression do outside of FSHD?**

DUX4 encodes an important developmental transcription factor



**Conserved function in DUX
gene family in mammals**

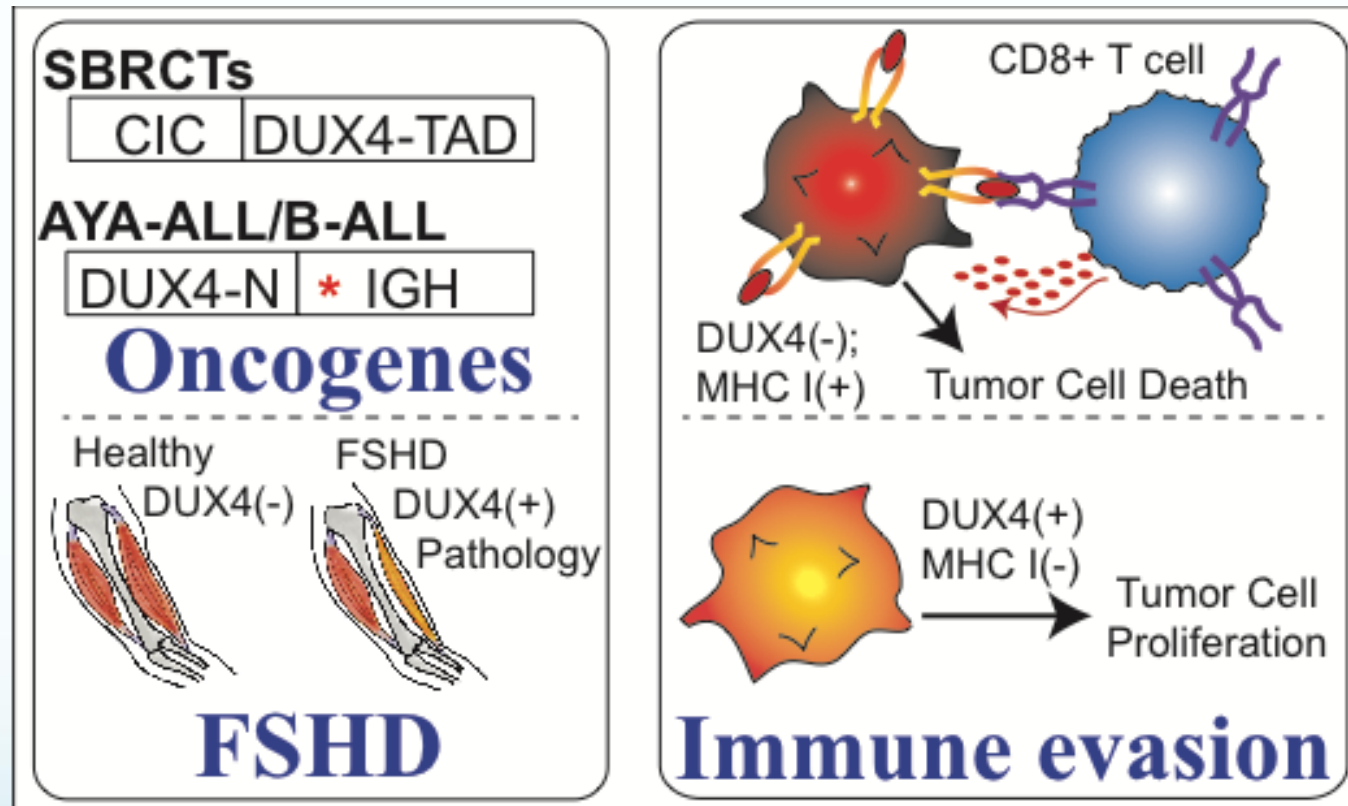
Healthy DUX4 expression is extremely limited



Hendrickson *et al.* 2017 Nat Genetics

Snider *et al.* 2010 PLoS Genetics

Aberrant DUX4 expression is pathogenic



Kawamura-Saito *et al.* 2006 Hum Mol Genetics

Wei *et al.* 2018 Cancer Discov

Yasuda *et al.* 2016 Nat Genetics

Lemmers *et al.* 2010 Science

Snider *et al.* 2010 PLoS Genetics

Chew *et al.* 2010 Dev Cell

What happens when DUX4 is misexpressed?

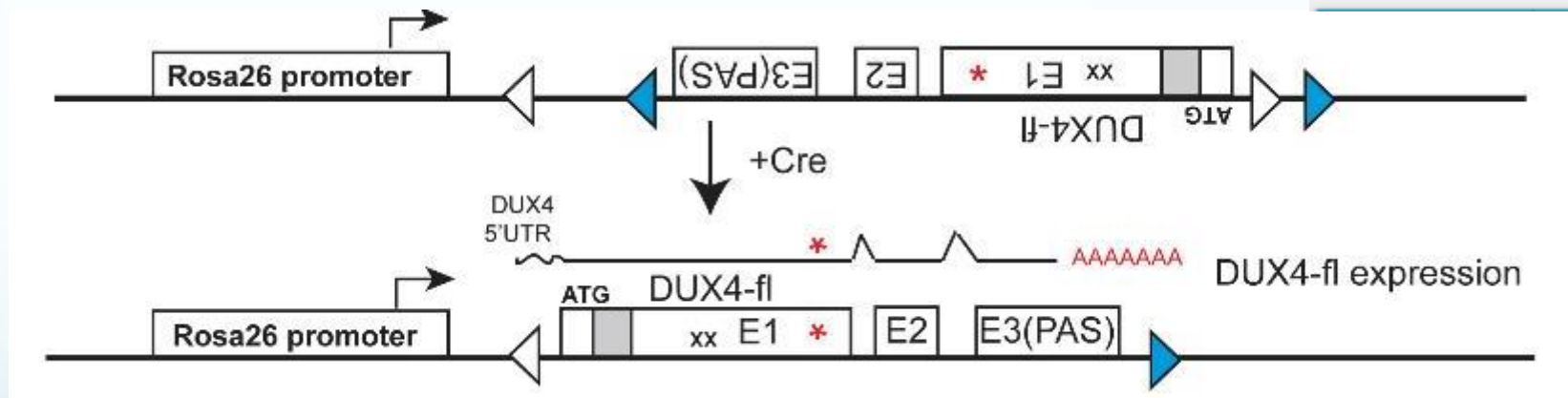


Generation of an FSHD-like mouse based on low level DUX4 expression



Takako Jones, PhD

FLExDUX4



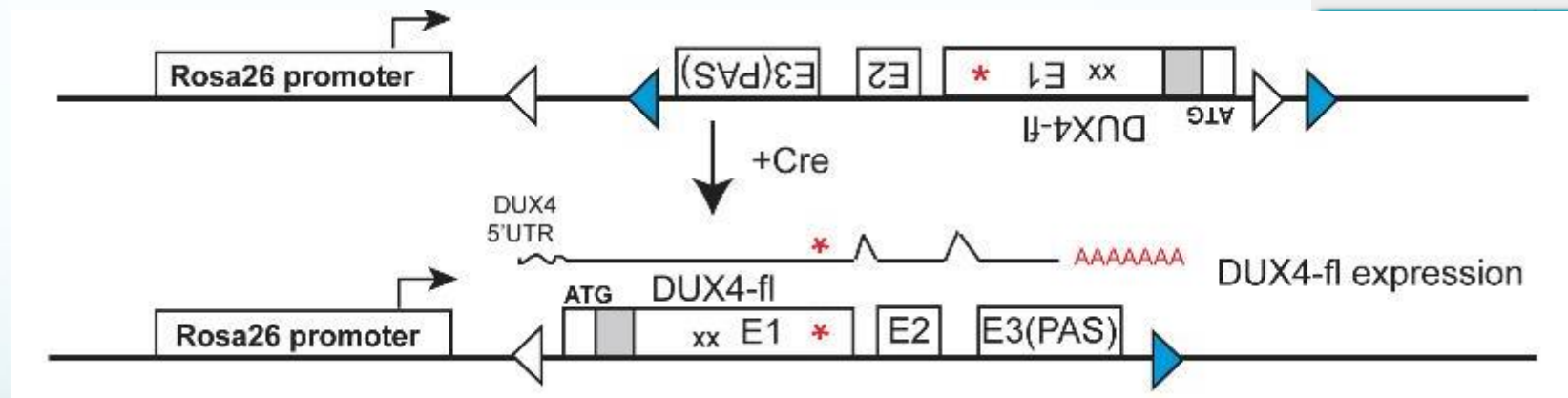
The *Rosa26* promoter ensures robust DUX4-fl expression in all cells that underwent cre-mediated inversion

The *FLExDUX4* mouse model allows for control of the timing and level of human DUX4 expression in muscles of an adult mouse



Takako Jones, PhD

FLExDUX4



No DUX4

More DUX4

Healthy

FSHD-like pathology

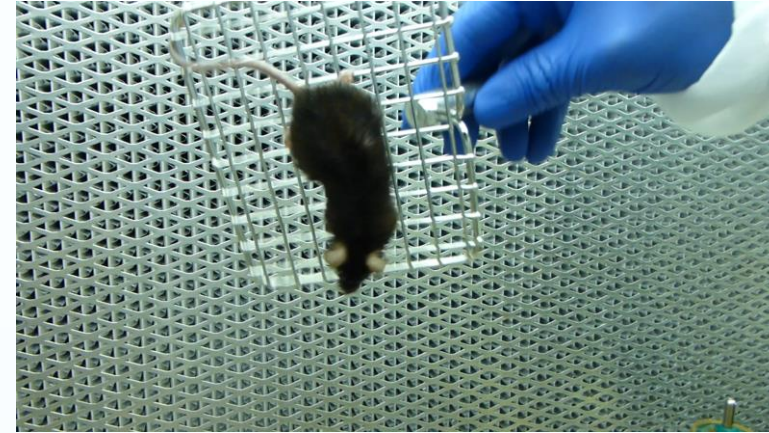
Induced expression of DUX4 in adult mice leads to an FSHD-like myopathy

>1 min suspended



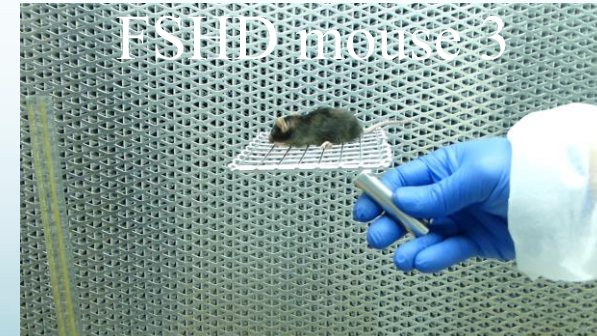
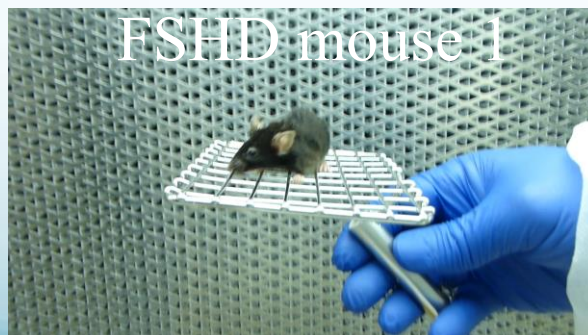
Healthy control

>1 min suspended



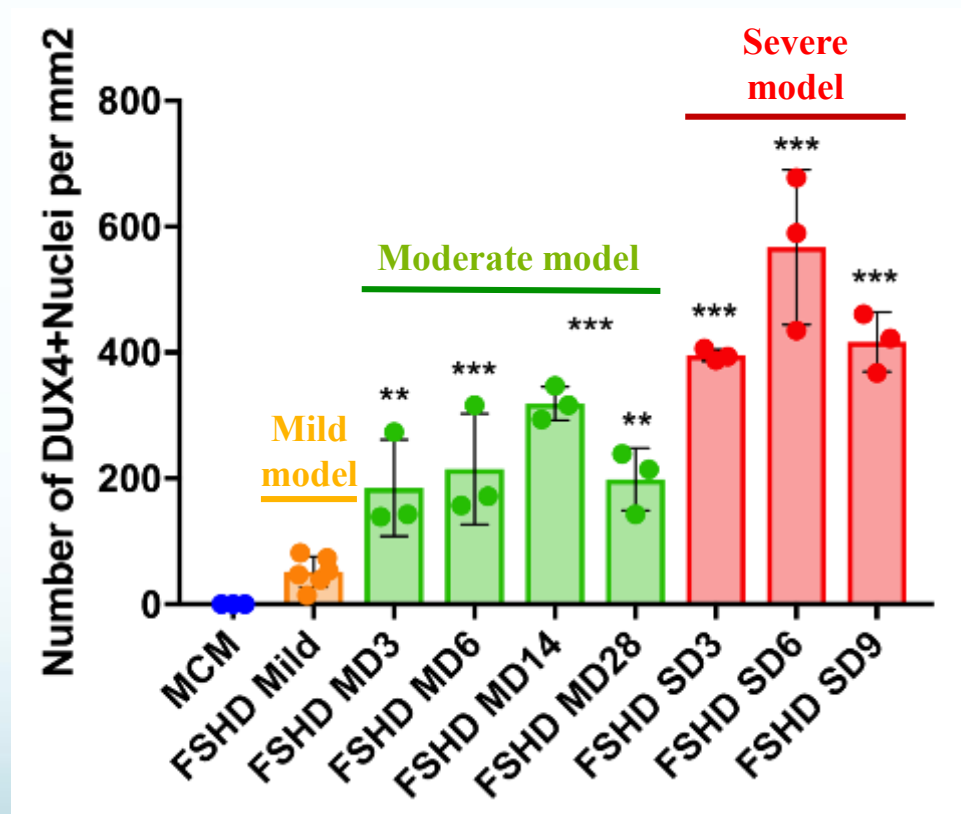
FLEXDUX4 mouse before disease onset

<2 second suspended



FLEXDUX4 mice after induced DUX4 expression

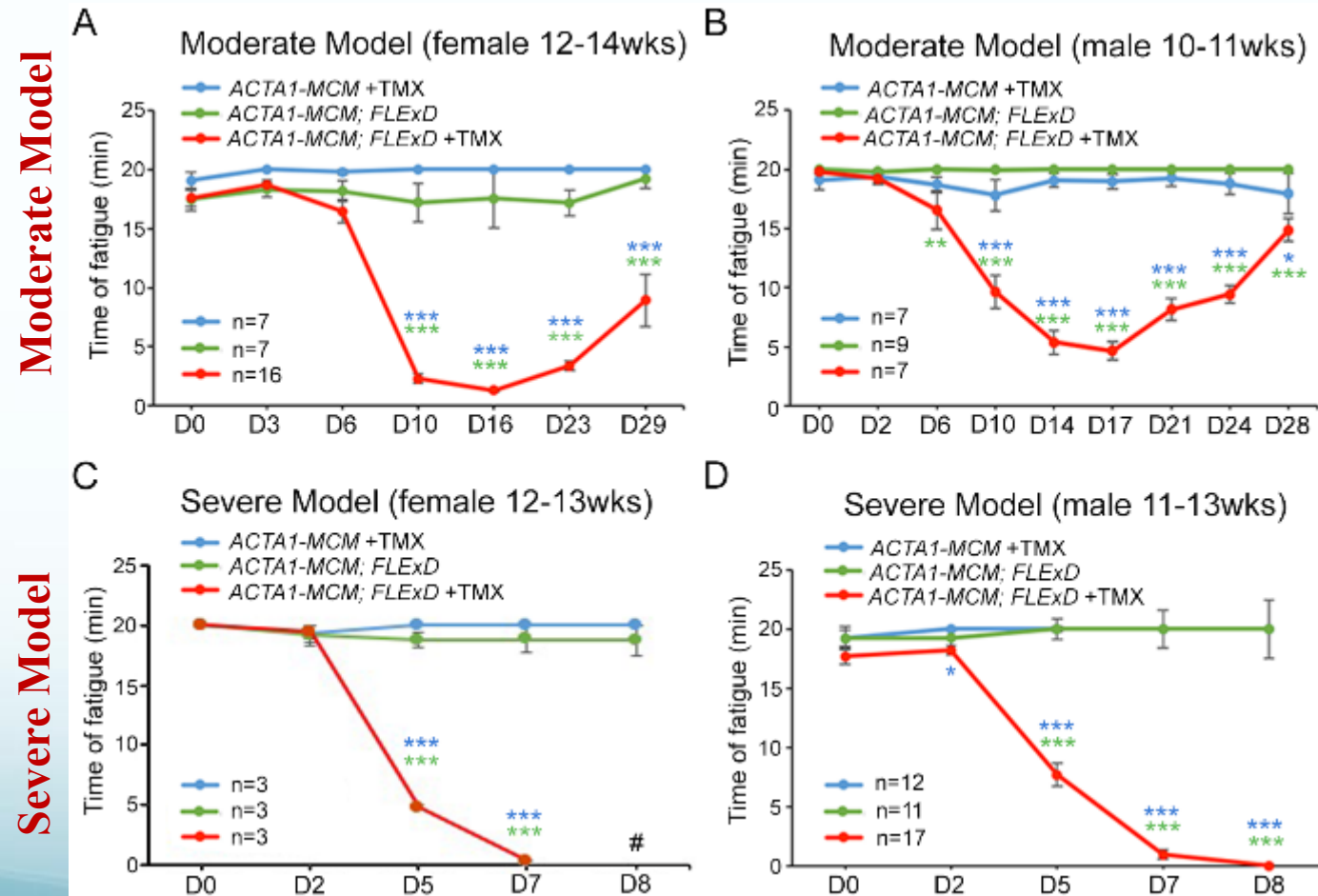
Small increases in DUX4-FL protein levels lead to increased severity of FSHD-like disease



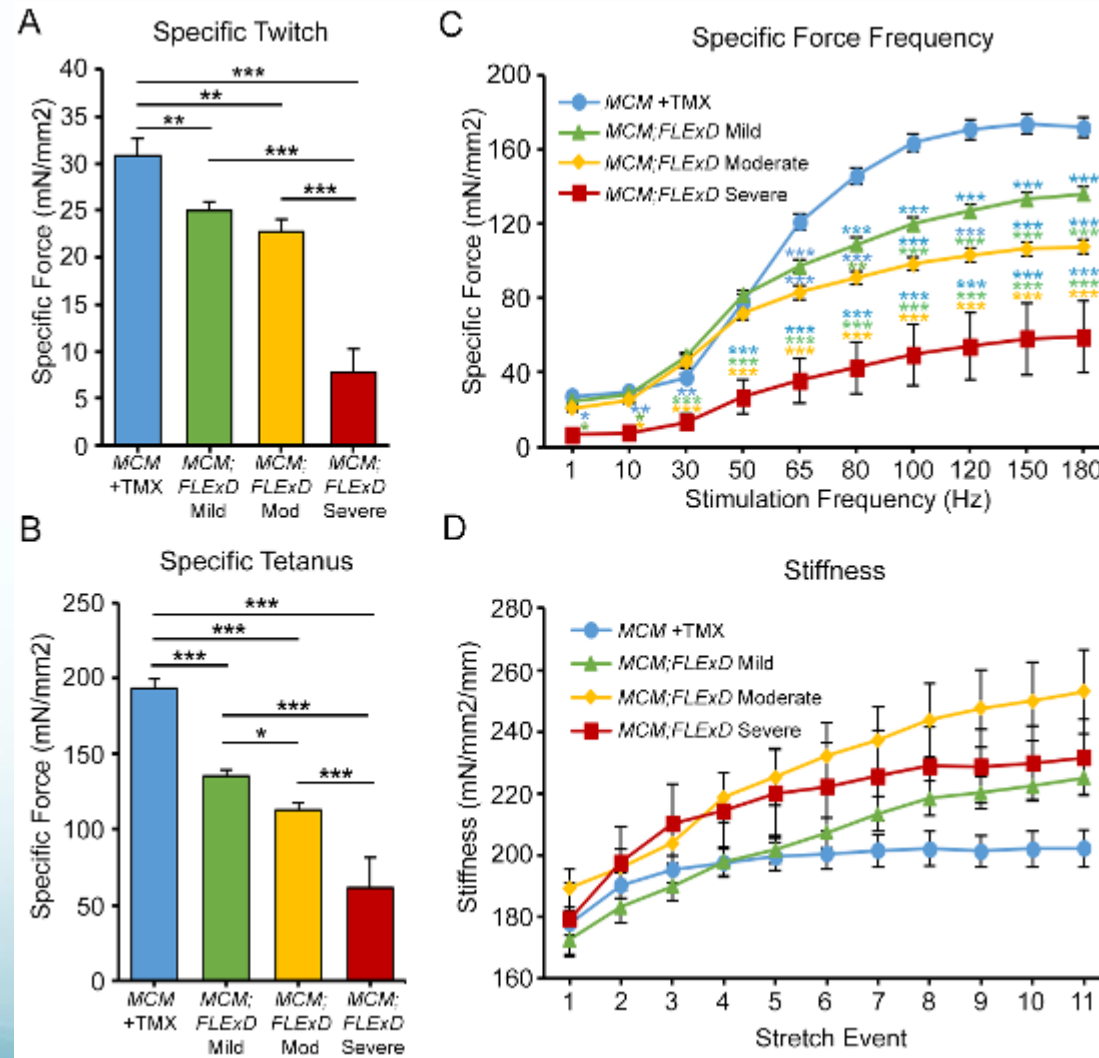
As DUX4 expression increases, treadmill running fitness declines

Female mice

Male mice

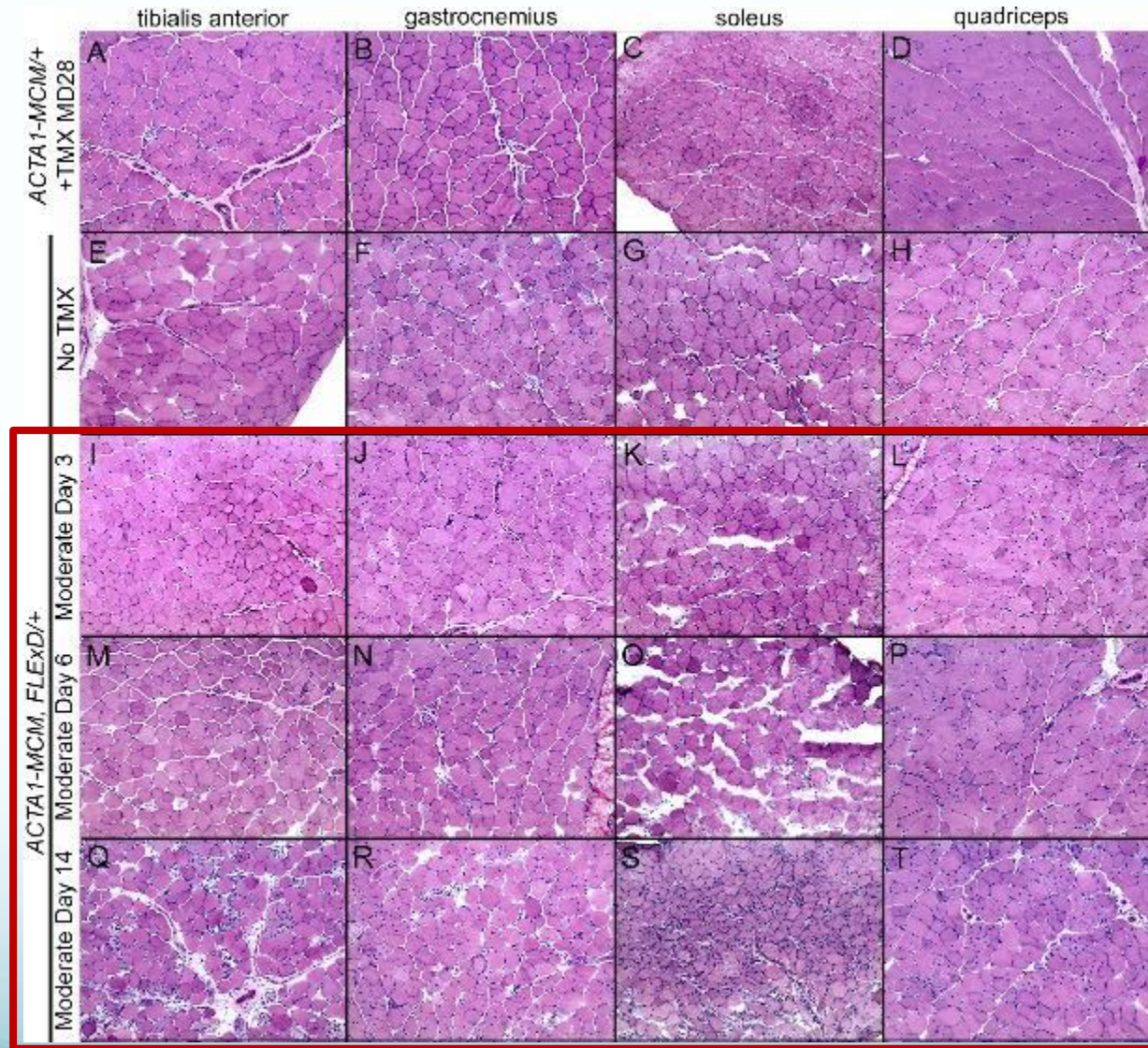


As DUX4 expression increases, skeletal muscles get weaker



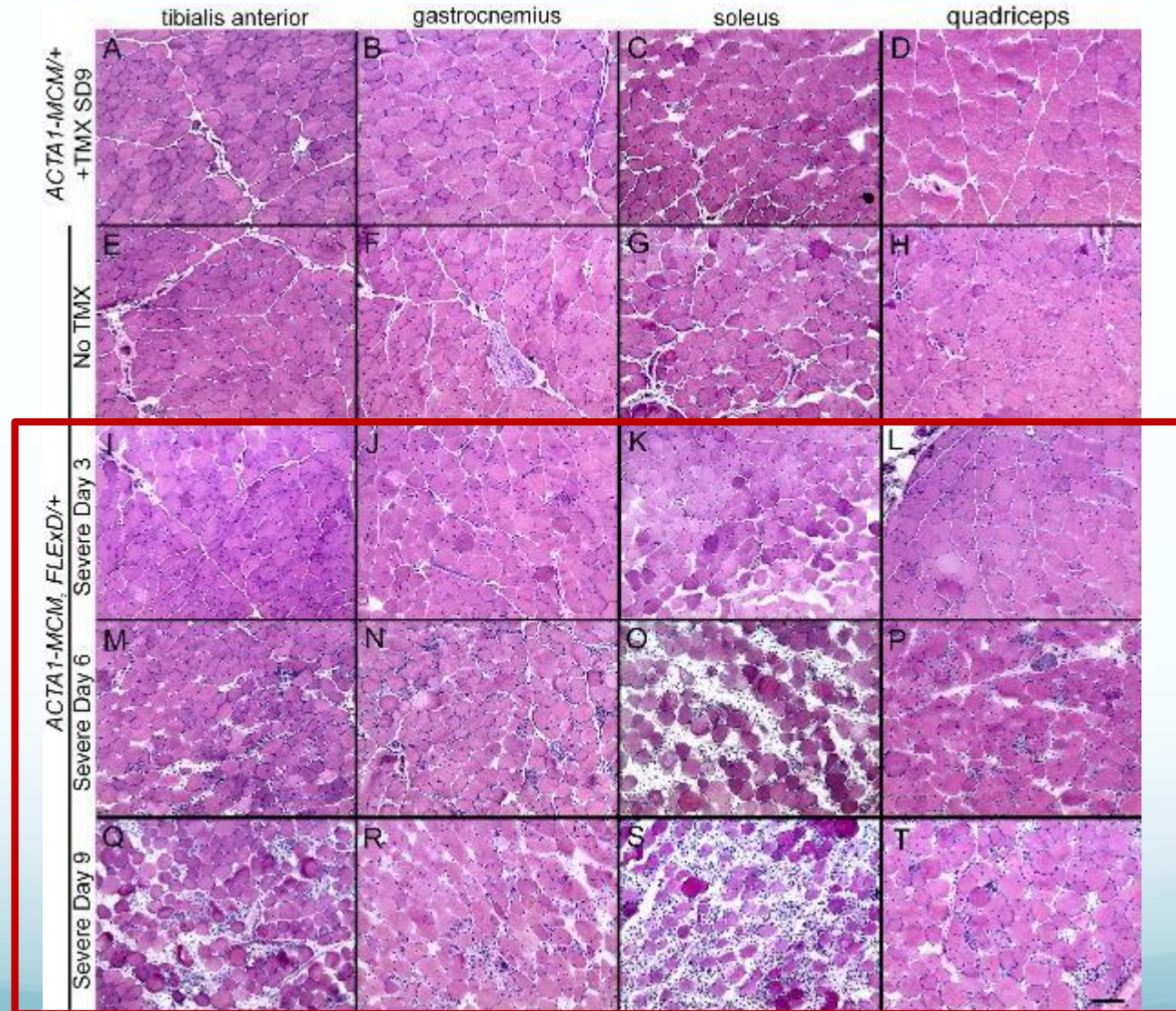
Increased DUX4 expression leads to increased muscle histopathology

Moderate Model



Increased DUX4 expression leads to increased muscle histopathology

**Severe
Model**



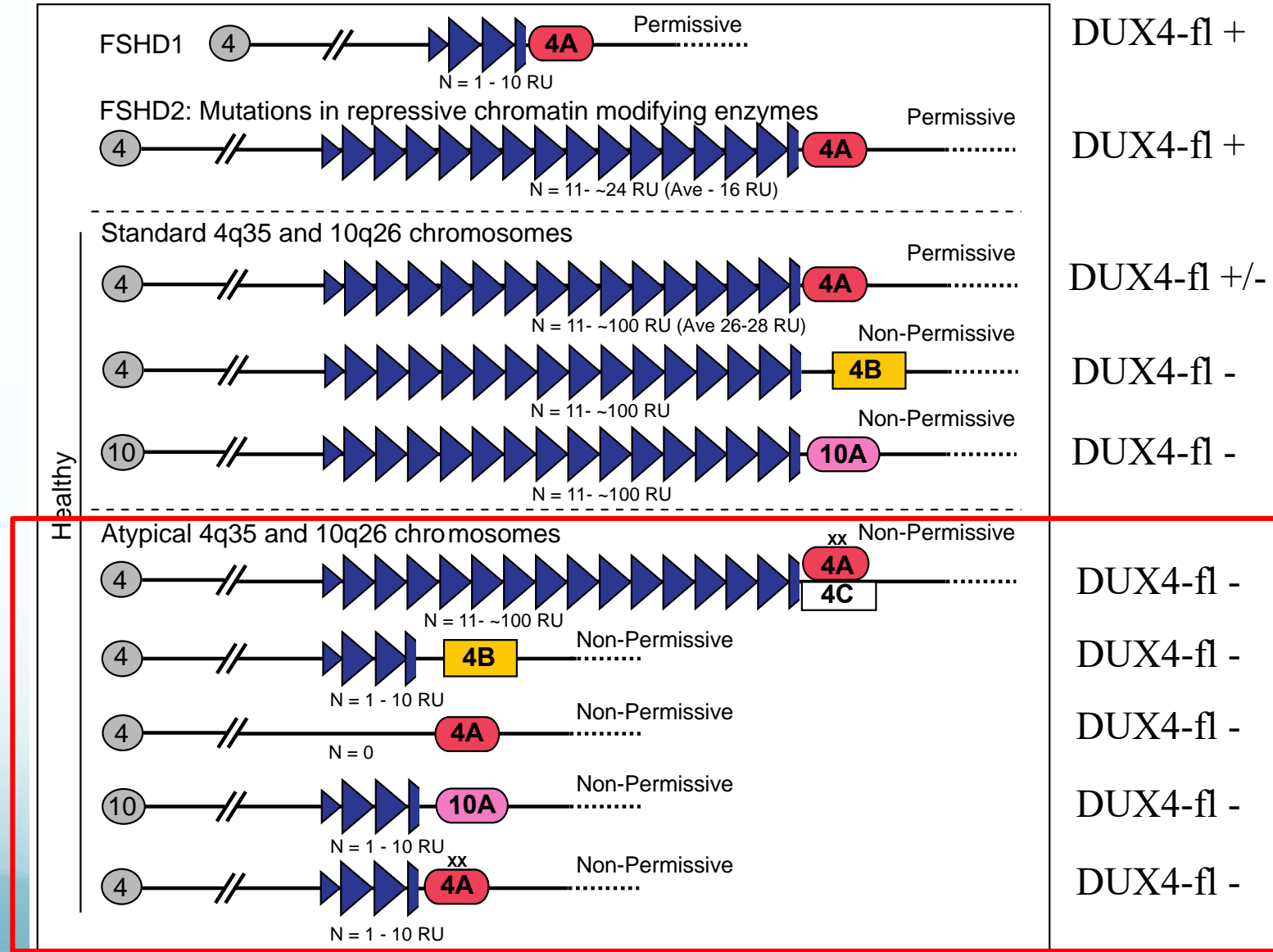
Dose dependent increases in DUX4 expression in skeletal muscle lead to:

- **Decreased muscle function**
- **Decreased muscle strength**
- **Increased muscle histopathology**

Do we need DUX4 expression as adults?

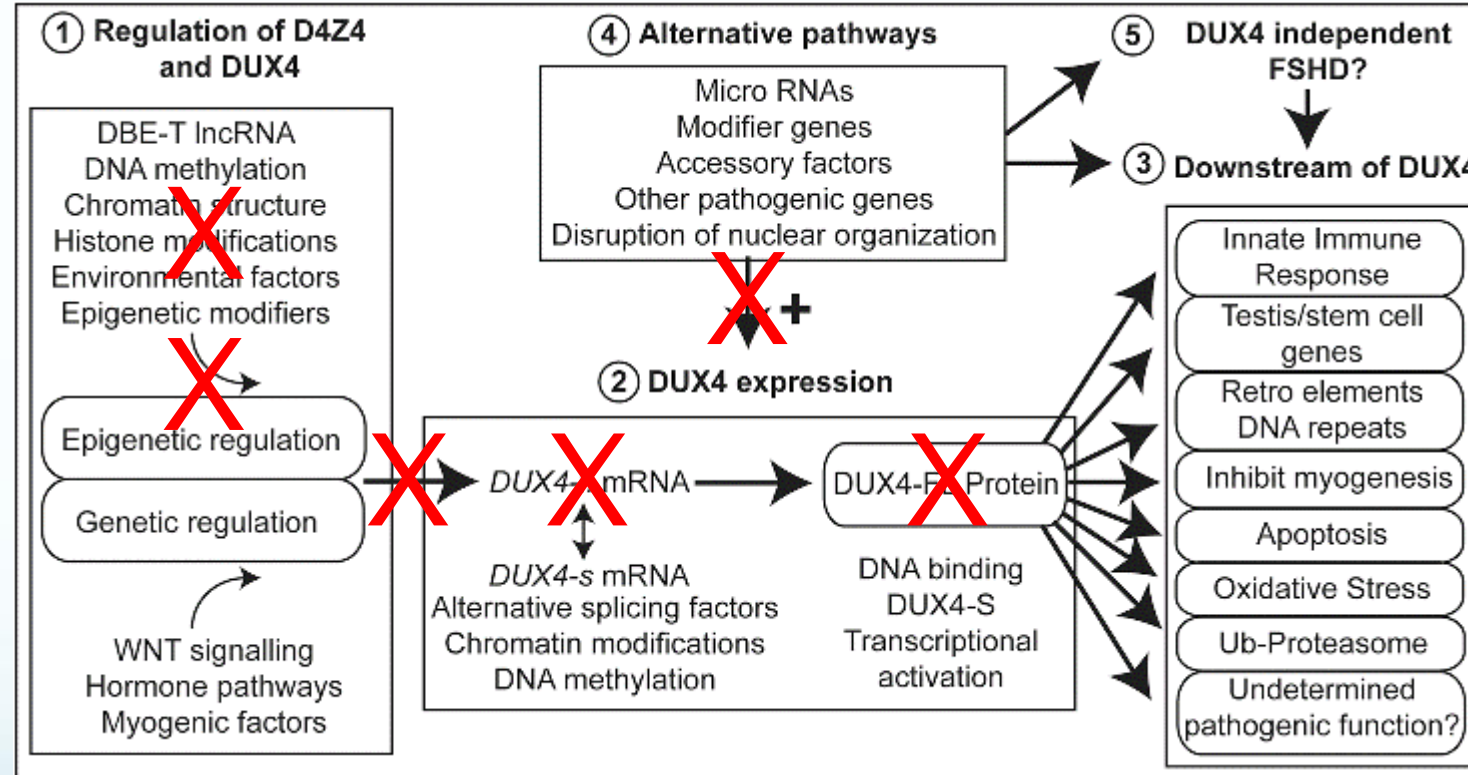
***25% of the population is “non-permissive” for the
somatic DUX4 mRNA polyadenylation signal**

Half of the 4q chromosomes are non-permissive for DUX4



Reduction or elimination of DUX4 expression targets the key pathogenic mechanism

Any level of DUX4 reduction may have therapeutic benefit!



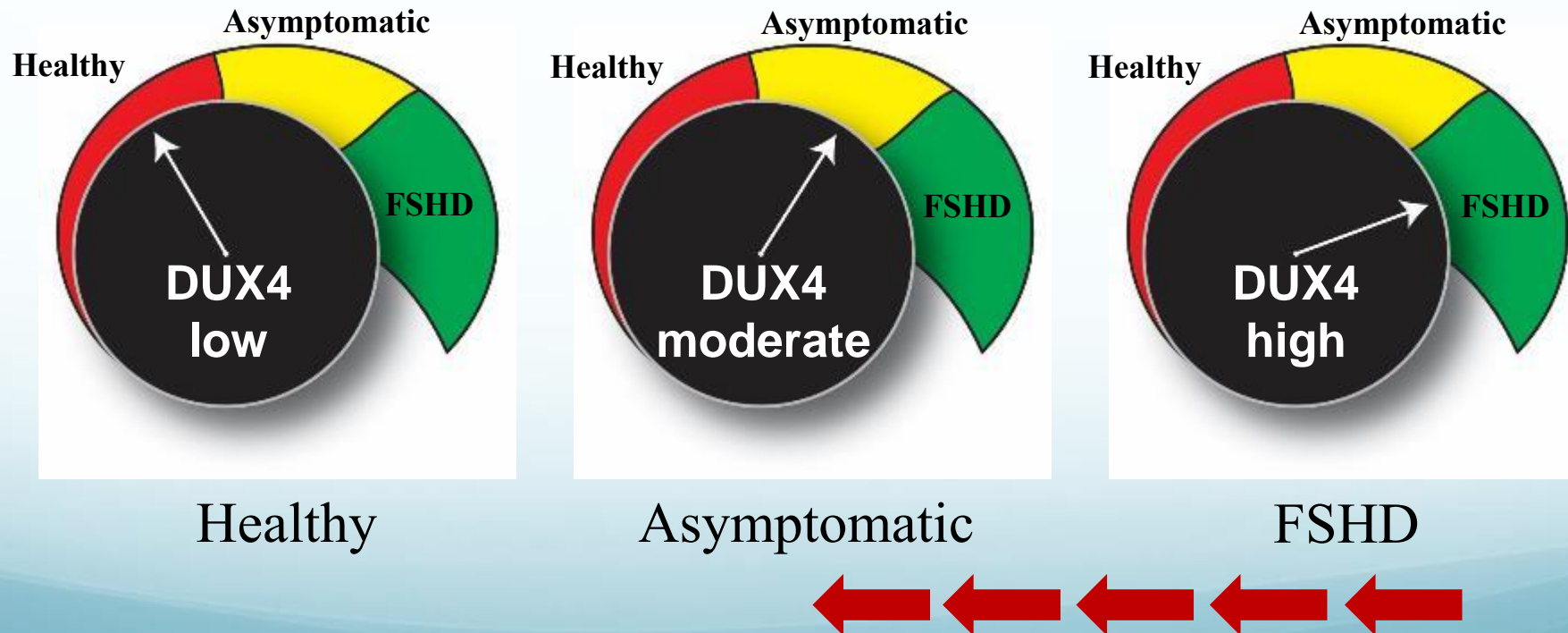
Small molecules targeting DUX4 regulation or function

***Morpholinos/PMOs/shRNAs & *miRNAs**

***CRISPR-inhibition**

***delivery concerns**

**Small changes in epigenetic state
and/or DUX4 expression levels
have large clinical consequences**





Questions?

Contact: peterjones@med.unr.edu
<https://med.unr.edu/jones-lab>

p38 inhibitors for FSHD: turning off DUX4

Fran Sverdrup, PhD

Department of Biochemistry and Molecular Biology

Saint Louis University

November 7, 2019

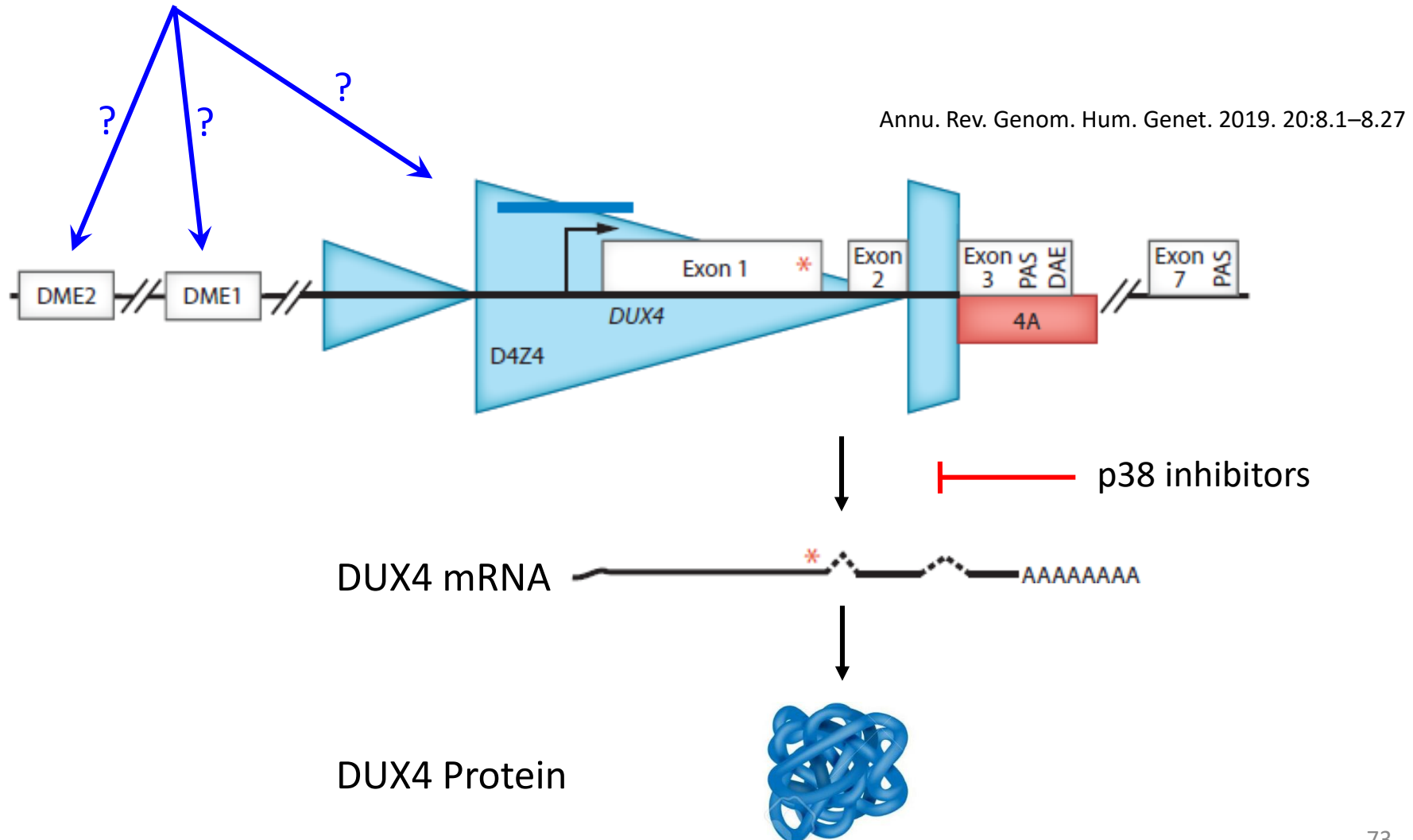
p38 inhibitors for FSHD: Turning off DUX4

- Therapeutic strategy: targeting DUX4 expression
 - Identification of p38 inhibitors
- p38: muscle biology
- Choice of losmapimod
- Example of losmapimod turning off DUX4 in xenograft mouse model
- Role of p38 in promoting DUX4 expression (work in progress)

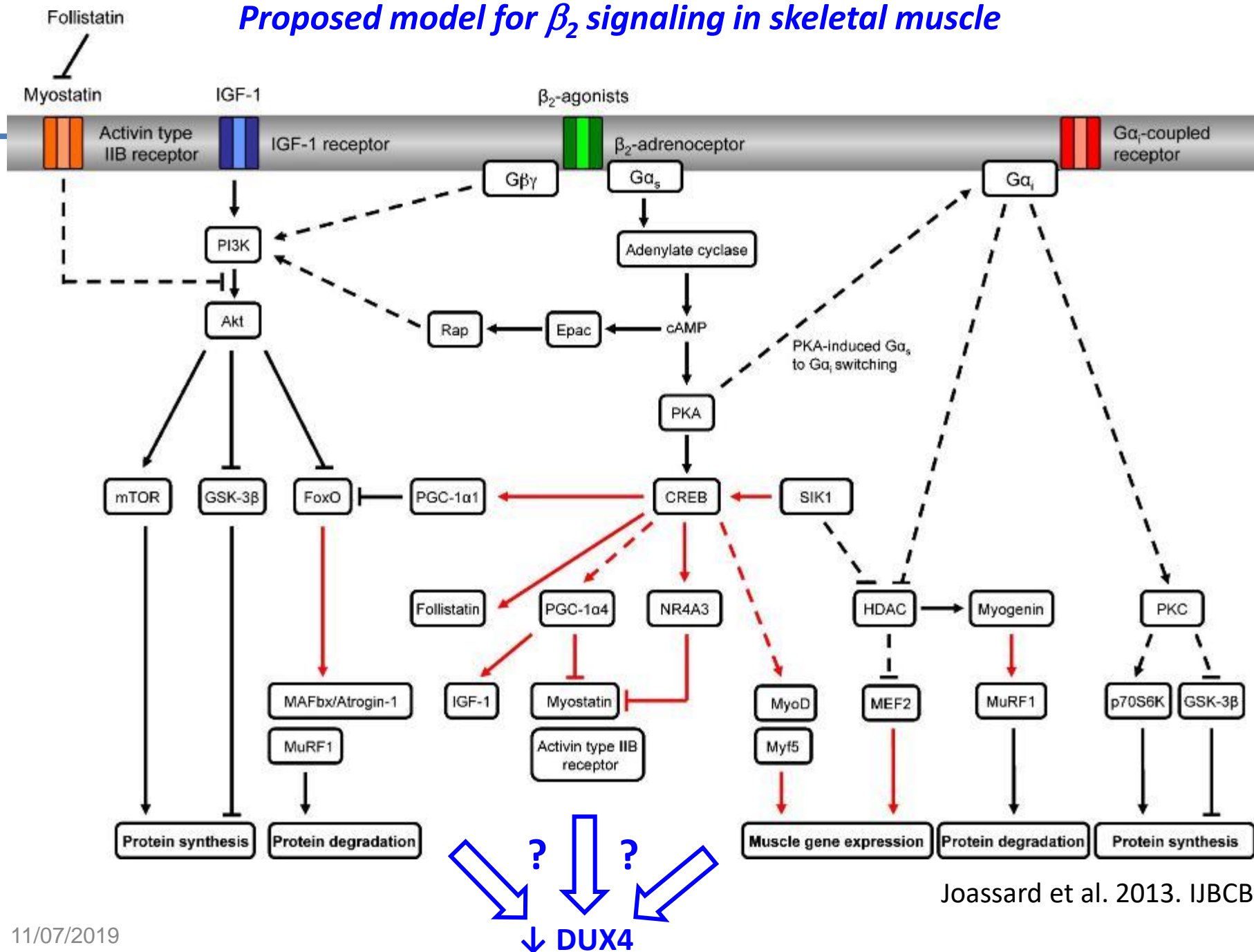
Therapeutic strategy

Suppress transcription of DUX4 mRNA

Factors that promote transcription are not understood

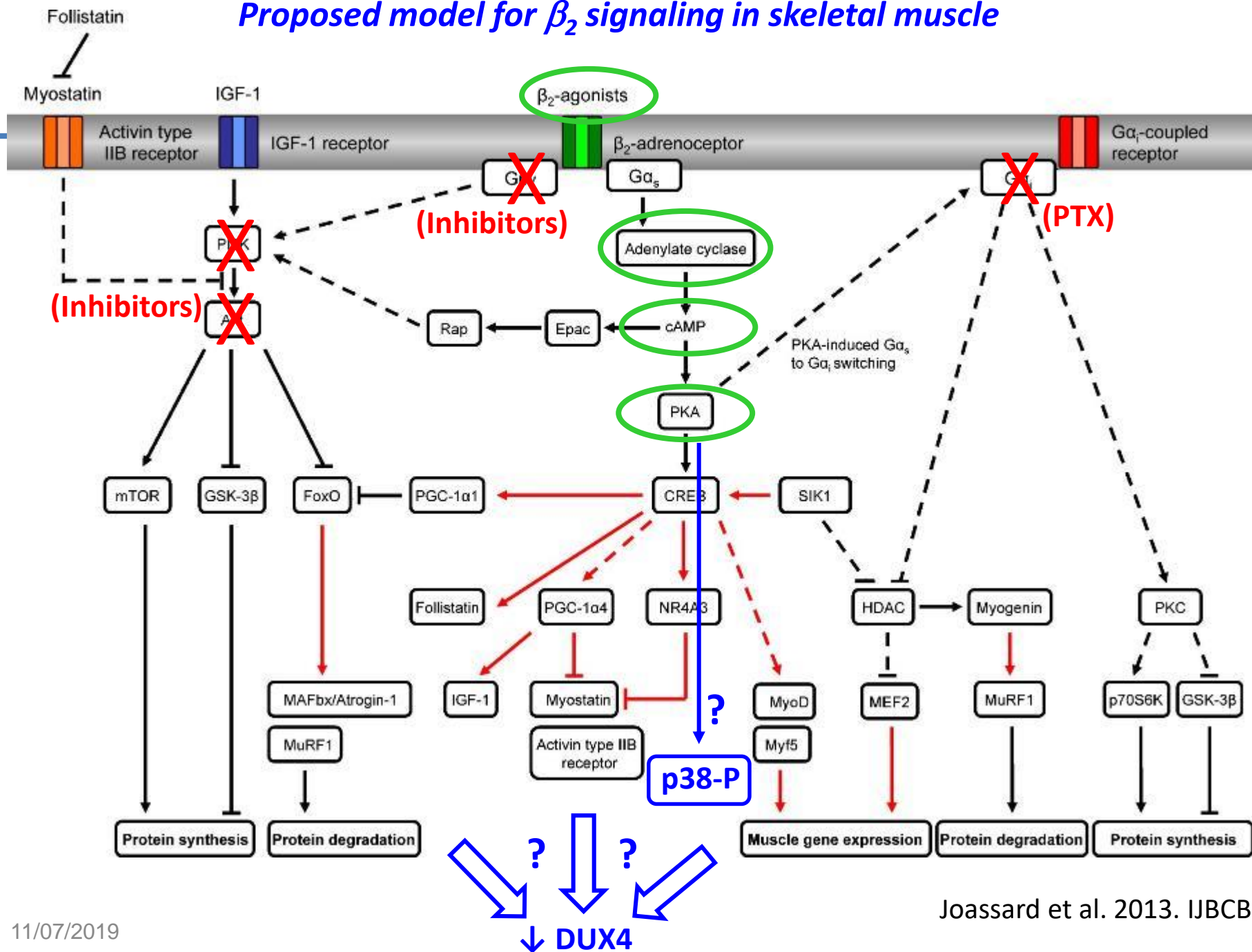


Proposed model for β_2 signaling in skeletal muscle



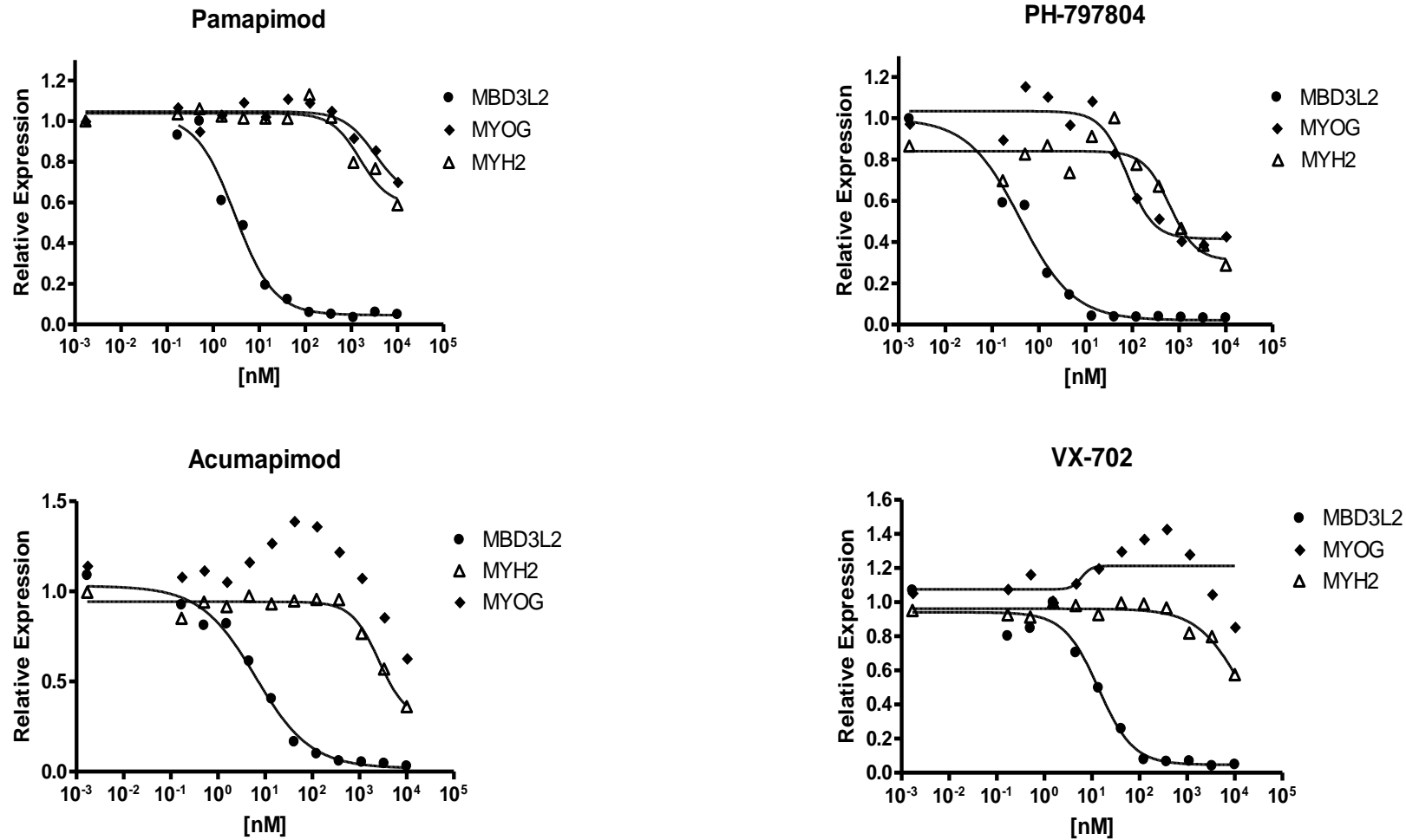
Joassard et al. 2013. IJBCB

Proposed model for β_2 signaling in skeletal muscle



FSHD drug targets

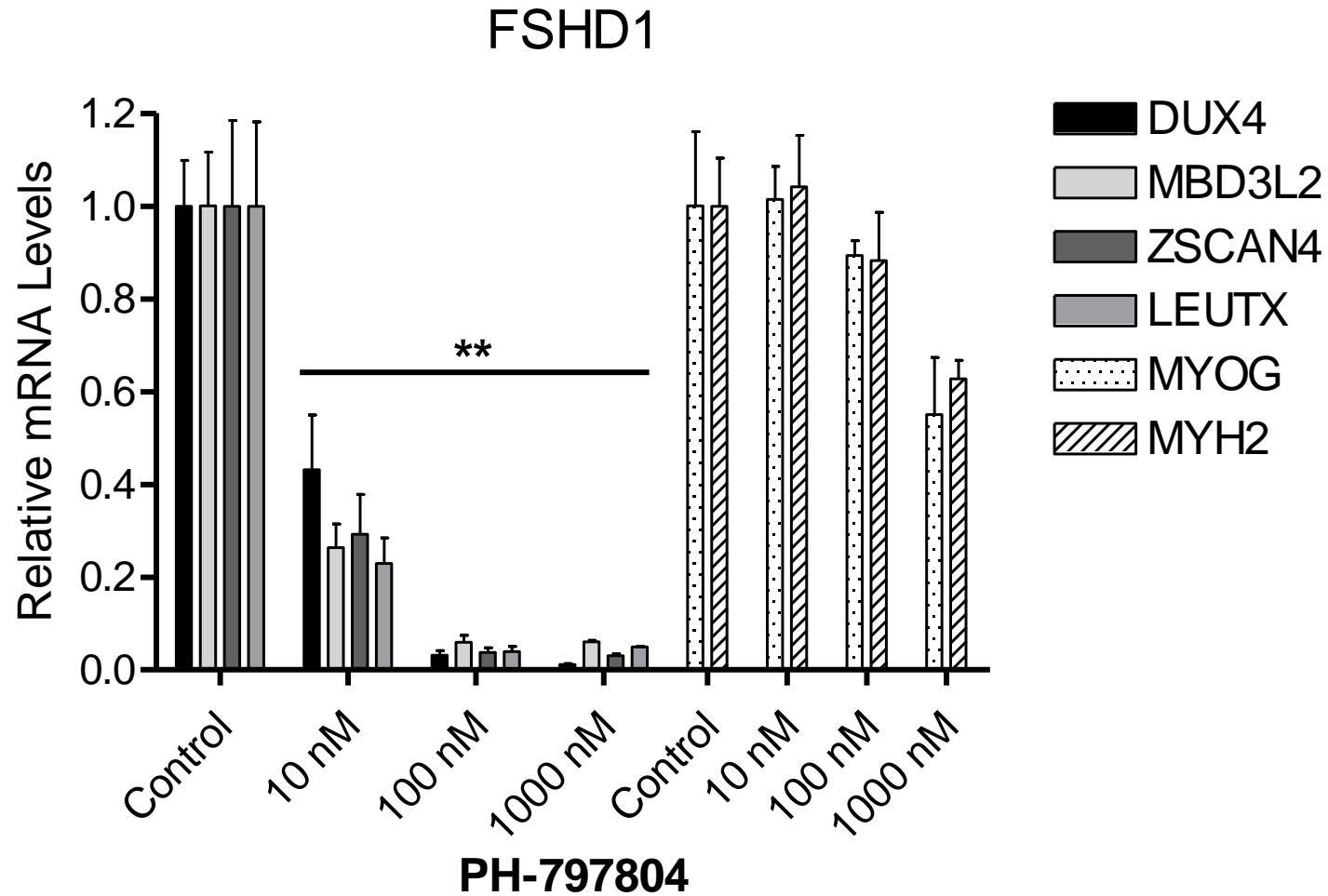
Clinically advanced p38 inhibitors suppress DUX4 expression



➤ p38 α/β inhibitors suppress DUX4 at levels that do not inhibit myogenesis

FSHD drug targets

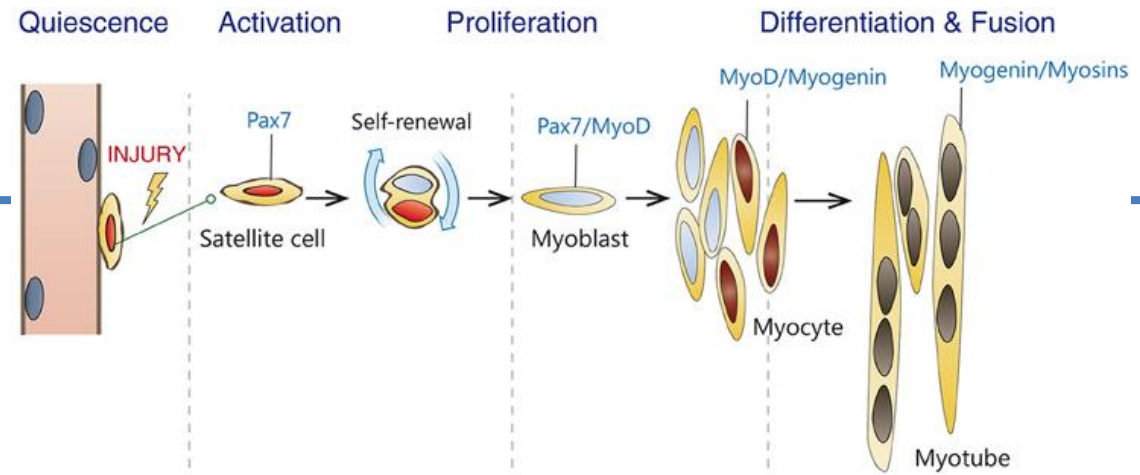
Clinically advanced p38 inhibitors suppress DUX4 expression



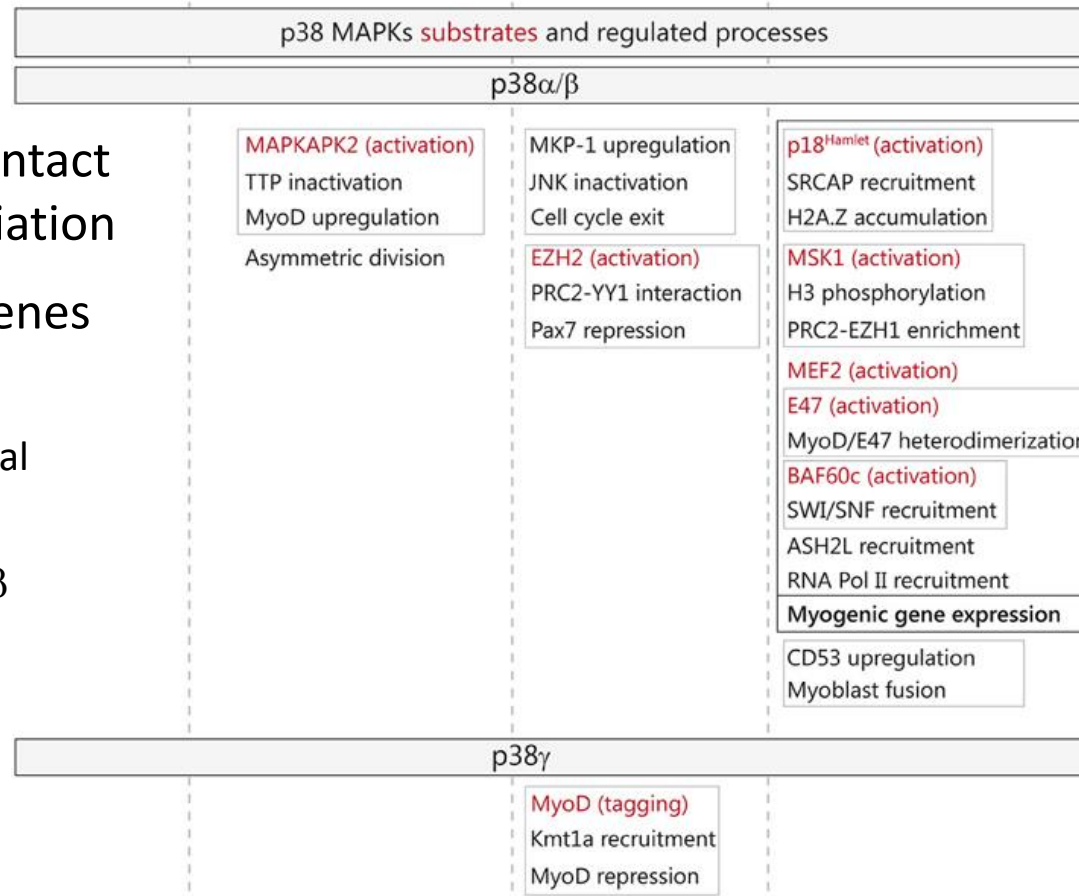
- p38 α/β inhibitors suppress DUX4 at levels that do not inhibit myogenesis

p38 MAP kinases

Role in muscle biology



- p38 α , p38 β and p38 γ isoforms expressed in skeletal muscle
- p38 α / β activated by cell-cell contact during normal muscle differentiation
- p38 α regulates large number genes during muscle differentiation
 - Many activities involved in temporal order of events
 - 5-fold higher expression than p38 β



p38

Therapeutic potential

- Knock out mice:
 - p38 α : (muscle-specific)
 - NOT DIRECTLY PATHOGENIC: Delayed myofiber growth and maturation, hyperproliferation of progenitors
 - reduced pathology in Mdx- and Sgcd-null dystrophic mice
 - p38 β : no muscle phenotype
- In vivo p38 inhibition (inhibitors targeting p38 α/β)
 - Improved self-renewal of satellite cells in aged muscles
 - Reduced pathology in Sgcd-null dystrophic mice
- p38 inhibition is valid therapeutic strategy
 - Suppress DUX4
 - Maintain muscle health
 - Potentially enhance muscle progenitor pools (satellite cells/myoblasts)

Clinically Advanced p38 Inhibitors

| Inhibitor | Mechanism/ Selectivity | Indications, Phs |
|----------------------------|---|--|
| PH-797804 | p38 α , 4X > p38 β | RA, COPD, Pain Phs II |
| Losmapimod (GW856553) | p38 α/β | Cardiovascular (ACS, MI) Phs III, MDD, COPD, Phs II |
| Dilmapimod (SB- 681323) | p38 | COPD Phs I, RA, ACS, LI Phs II |
| VX-702 | p38 α , 14X > p38 β | RA, Phs II |
| ARRY-371797 | p38 | LMNA-Cardiomyopathy, Phs III |
| Pamapimod (RO4402257) | p38 α , 34X > p38 β | RA Phs II |
| Acumapimod (BCT197) | p38 α/β | COPD Phs II |
| Pexmetinib (ARRY-614) | p38/Tie2 | Myelodysplastic syndrome, Phs I |
| Ralimetinib (LY2228820) | p38 α/β JNK2, JNK3 > JNK1 | Advanced cancer, Phs II |
| Talmapimod (SCIO 469) | p38 α , 10X > p38 β | RA, Myelodyplastic syndrome, Phs II |
| BMS-582949 | P38 α , 5X > p38 β | Atherosclerosis, RA, Phs II |
| TAK-715 | p38 α , 28X > p38 β | |
| Neflamapimod (VX-745) | p38 α , 22X > p38 β | Alzheimer's |
| Doramapimod (BIRB 796) | p38 $\alpha/\beta/\gamma/\delta$ | Phs II |

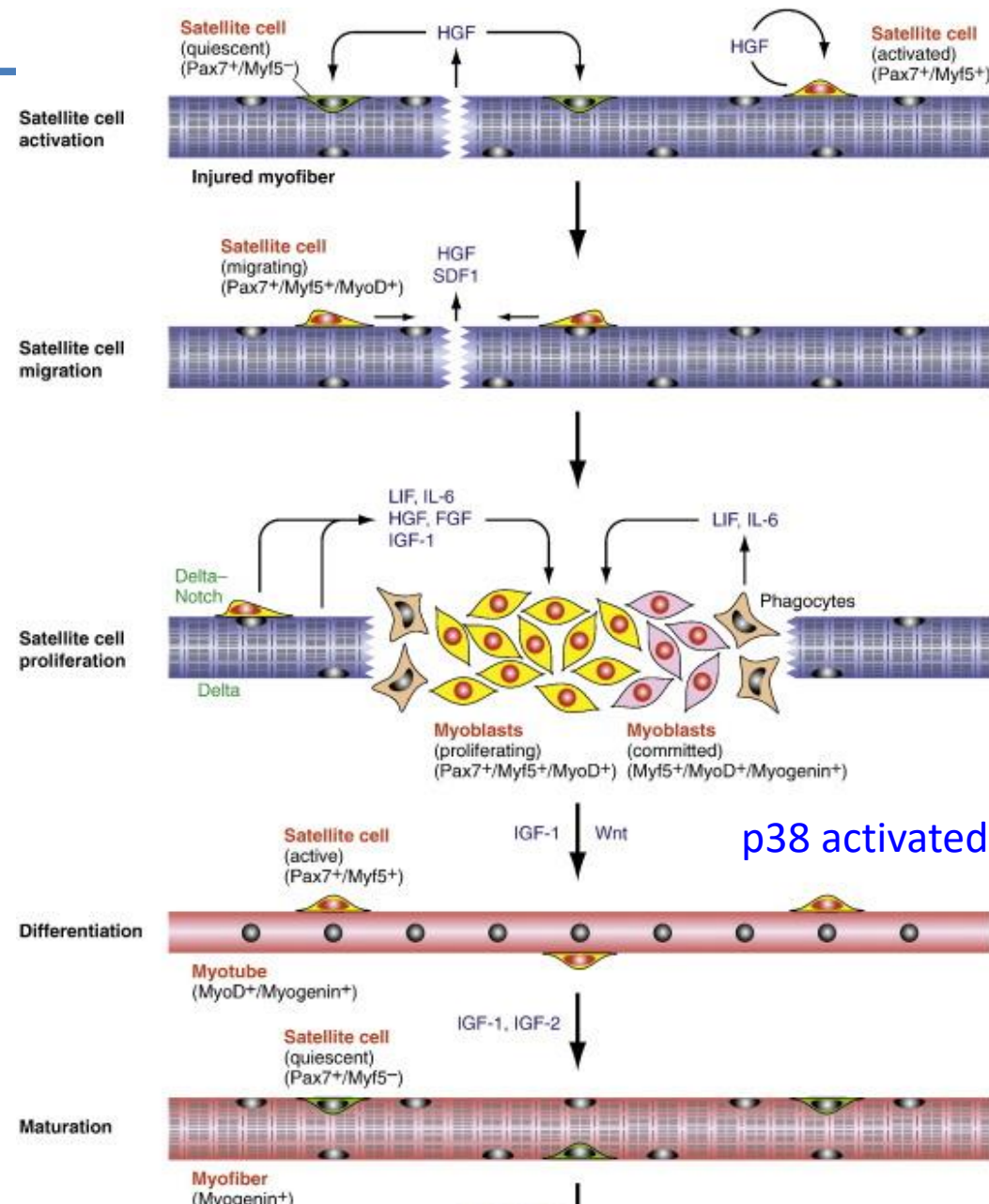
Failure in the clinic for efficacy in intended indications leaves many potential drug candidates to repurpose for FSHD!

RANK by:

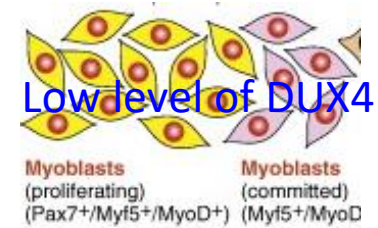
- Efficacy in mouse model
- Safety profile/experience

Muscle Regeneration in vivo

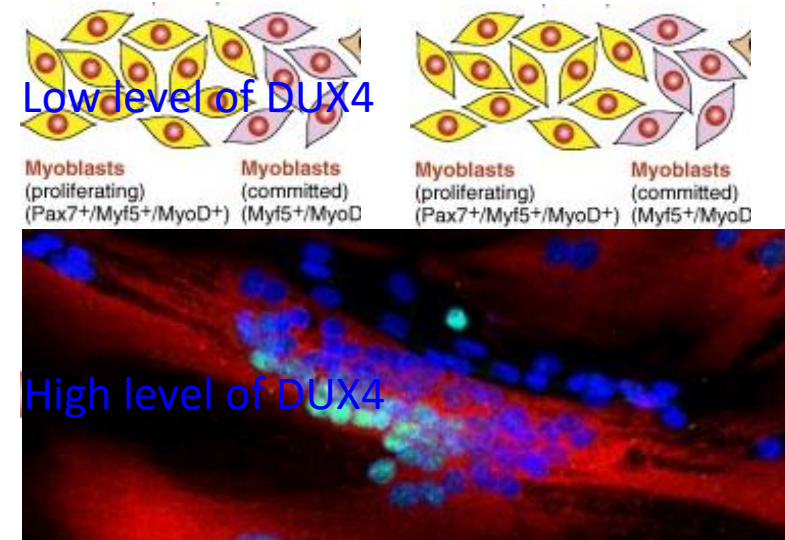
borrowed from: Endo, T. Bone. 2015. 80:2-15.



Cell culture model

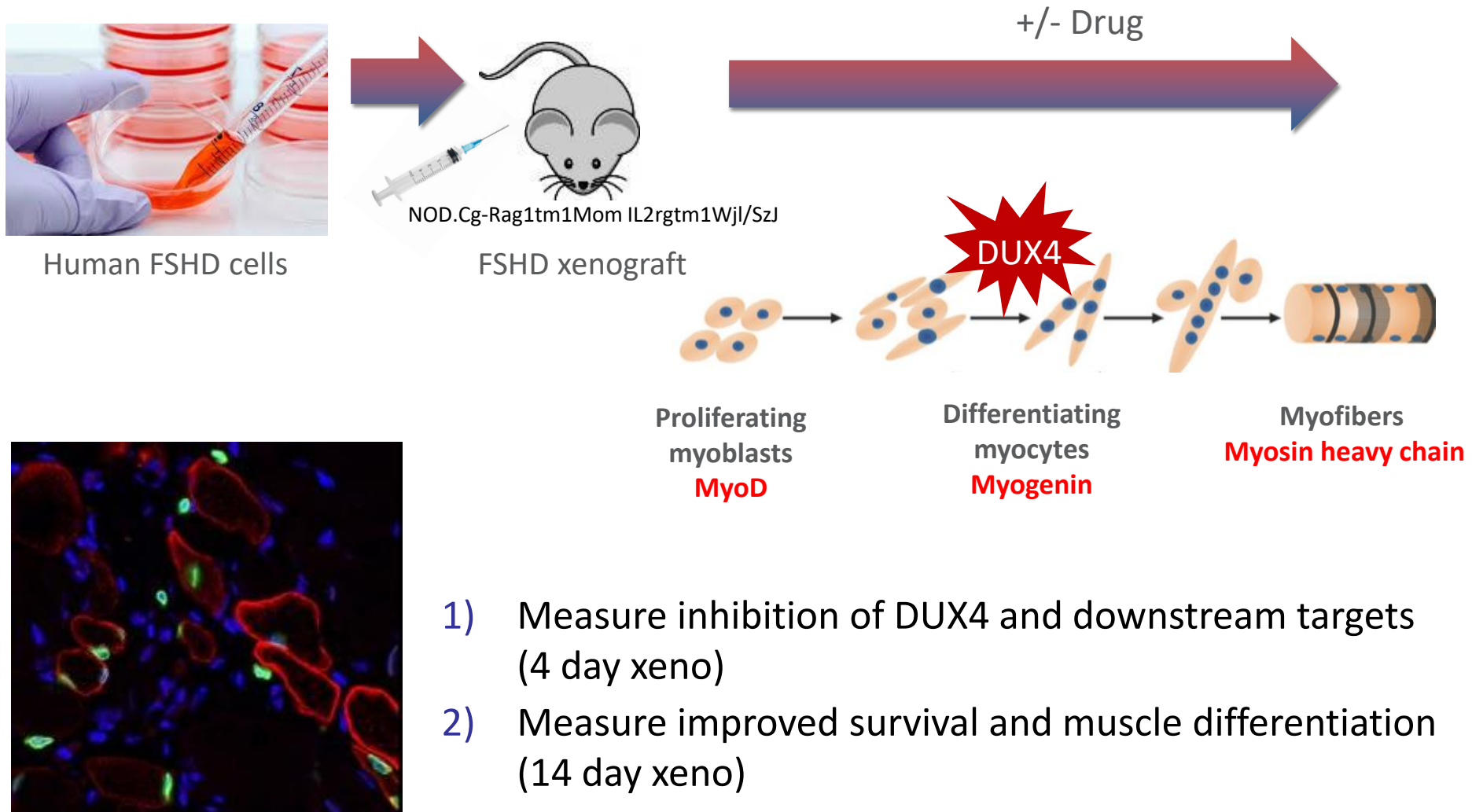


Mouse xenograft model



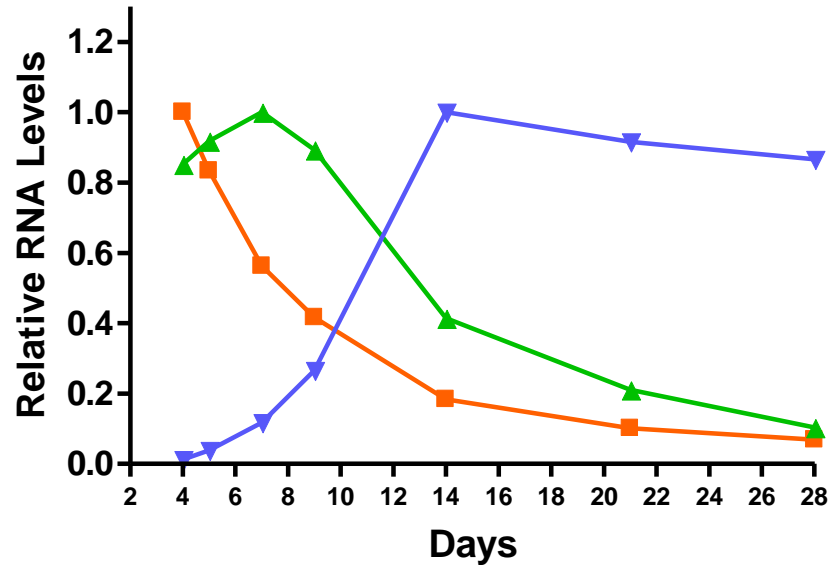
Xenograft model of FSHD

Human Epigenetic Regulation of DUX4



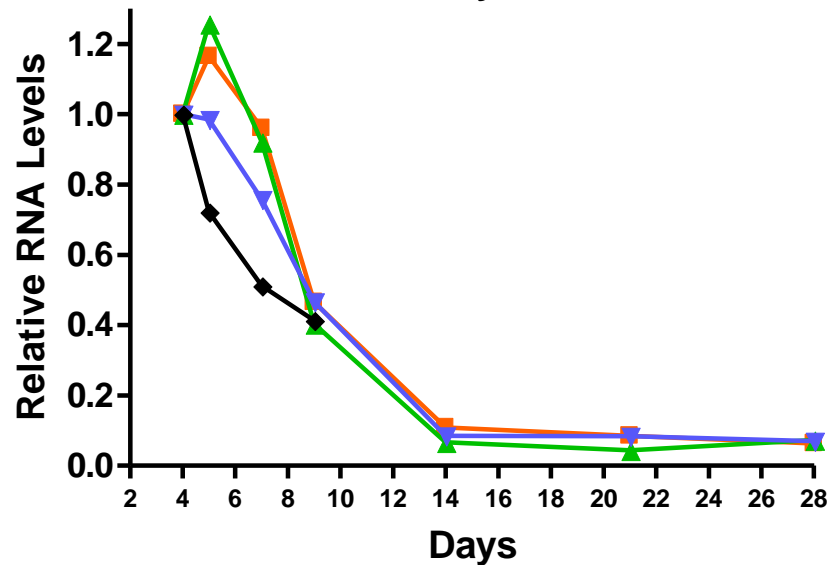
Xenograft model of FSHD

4 week profiling of gene expression



n = 6 animals per time point

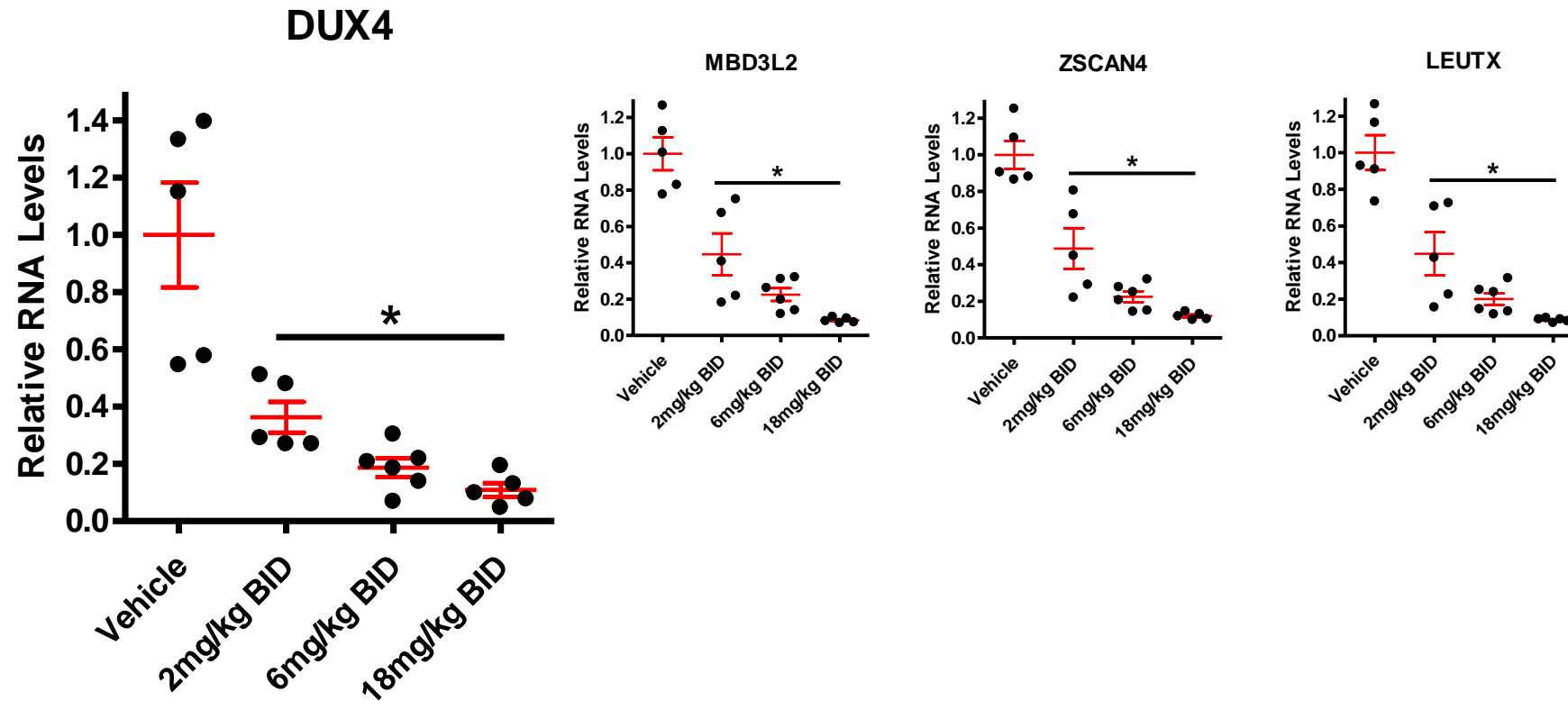
- MYOG (early differentiation) peaks on or before day 4
- MYH3 (regeneration) peaks ~day 7
- MYH2 (late differentiation) peaks ~day 14



- DUX4 peaks around day 4
- DUX4 targets peak ~day 5

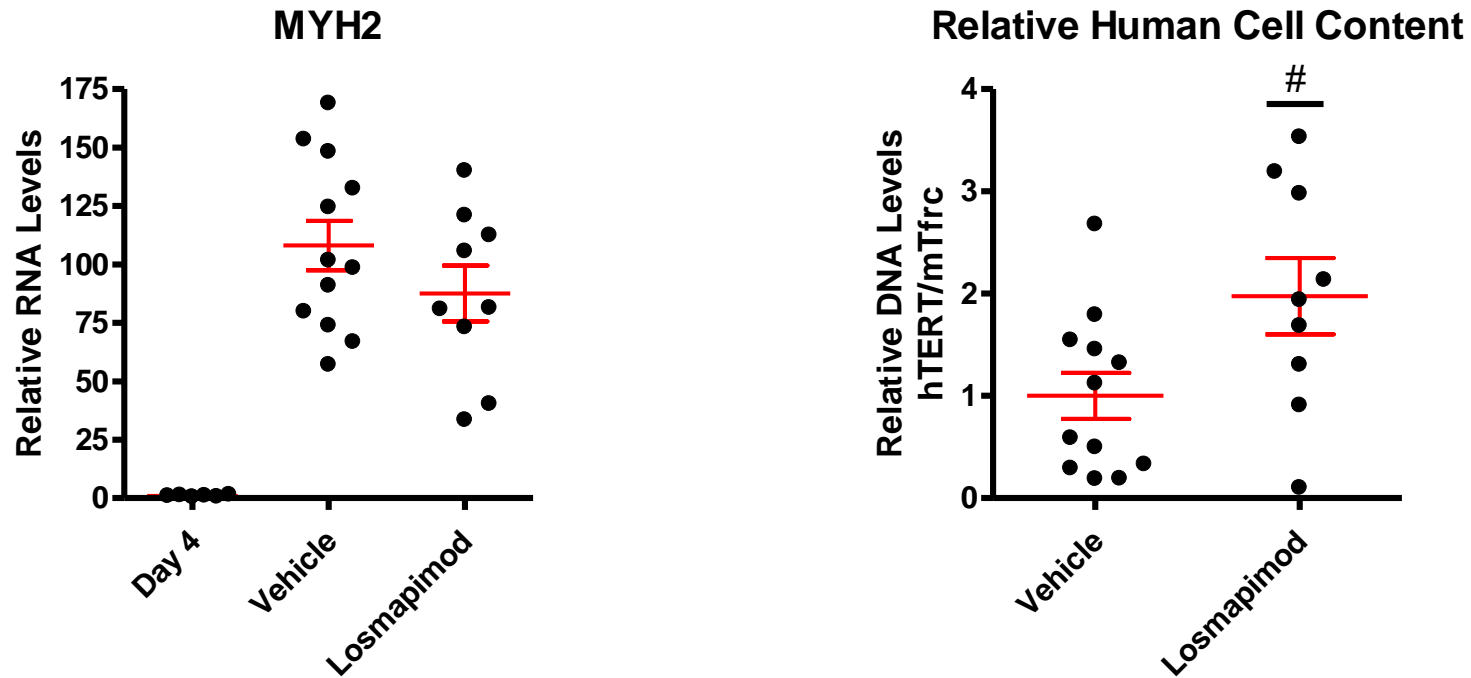
Xenograft model of FSHD

Losmapimod suppress DUX4 in human FSHD cells transplanted to mice



- Losmapimod reduces DUX4 expression by 80-90%
- These doses produce drug levels in mice that are similar to drug levels achieved in humans

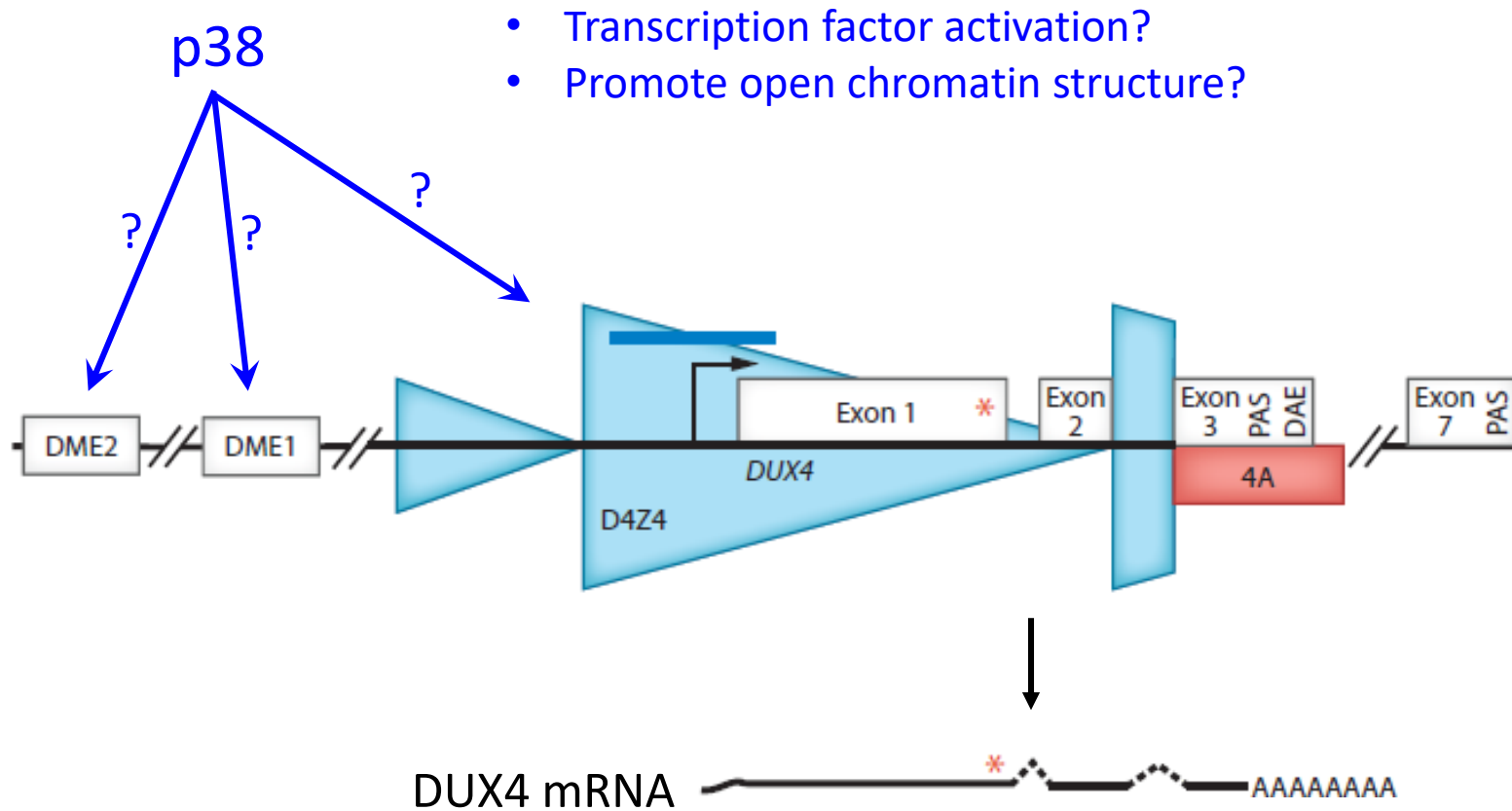
Losmapimod 14-day dosing (6 mpk BID)



- Human cells differentiate to mature myofibers with treatment (MYH2)
- Increase in human cell content suggests treatment may protect human cells from DUX4 toxicity

Mechanism

How does p38 turn on DUX4?



Summary

p38 inhibition turns off DUX4

- Clinically advanced p38i suppress DUX4 (and downstream target genes) without inhibiting muscle differentiation
 - Pharmacological and genetic depletion suggest viable therapeutic approach
 - Phs II/III p38 inhibitors are attractive drug candidates for FSHD
- Losmapimod suppresses DUX4 in mouse xenograft model at dosing levels that are relevant to human
- Losmapimod stands out as candidate p38 inhibitor for FSHD
 - Published human PK, muscle exposure in mice (internal), xenograft efficacy
 - Safety profile in large number of patients
- How p38 inhibitors suppress DUX4 currently under investigation
 - Potentially 2 or more mechanism

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Caroline Murphy

Katie Phelps

Ultragenyx Pharmaceutical

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Scott Galasinski

Marcus Andrews

Yael Weiss

Stephanie Watters

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Ultragenyx Pharmaceutical, Inc.

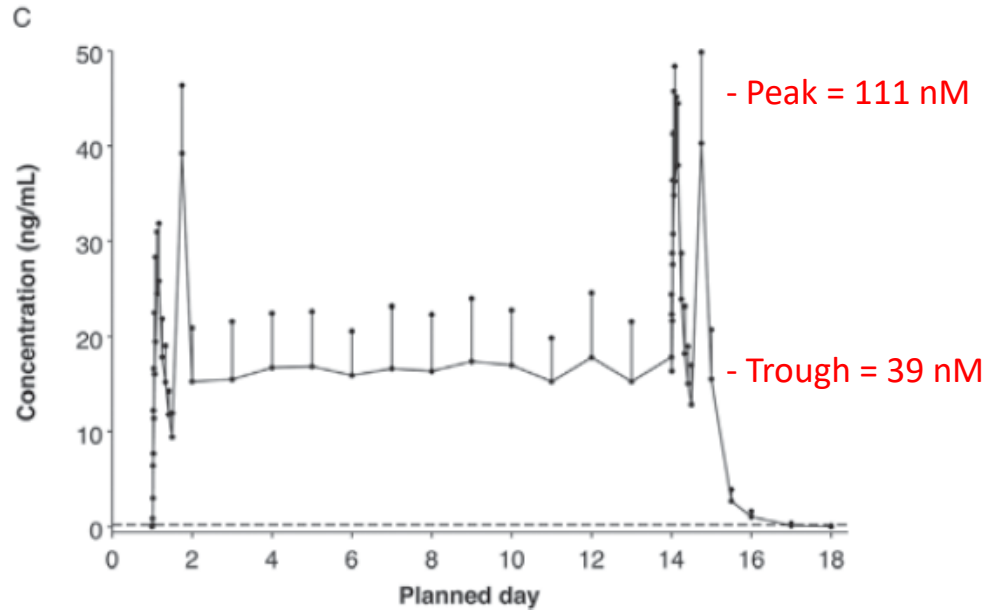
Muscular Dystrophy Association

Chris Carrino Foundation

Losmapimod

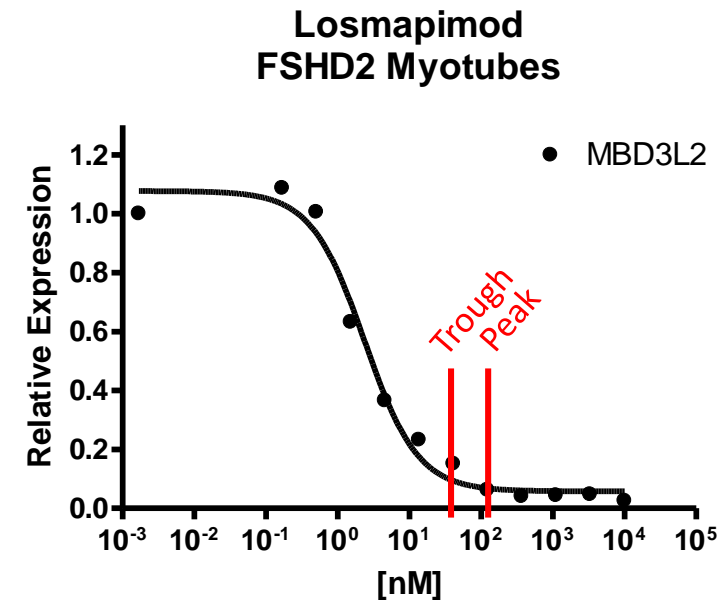
Clinical Pharmacokinetics

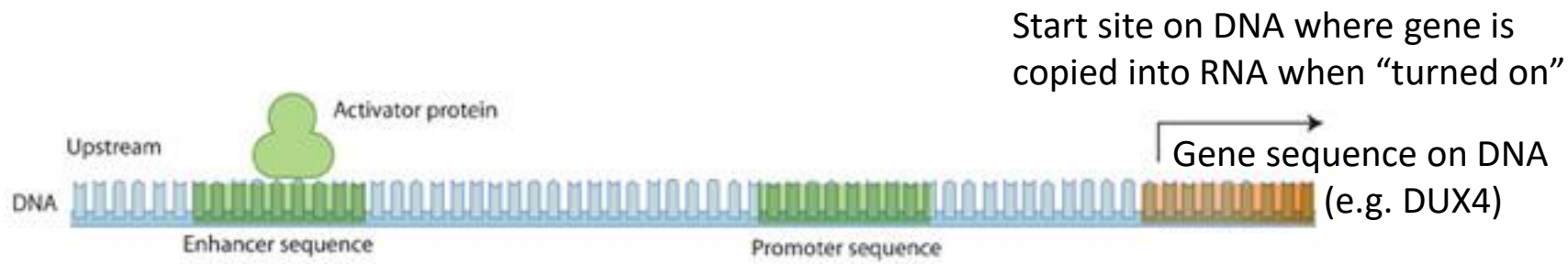
Clin Pharmacol Drug Dev. 2015; 4(4):262-9.



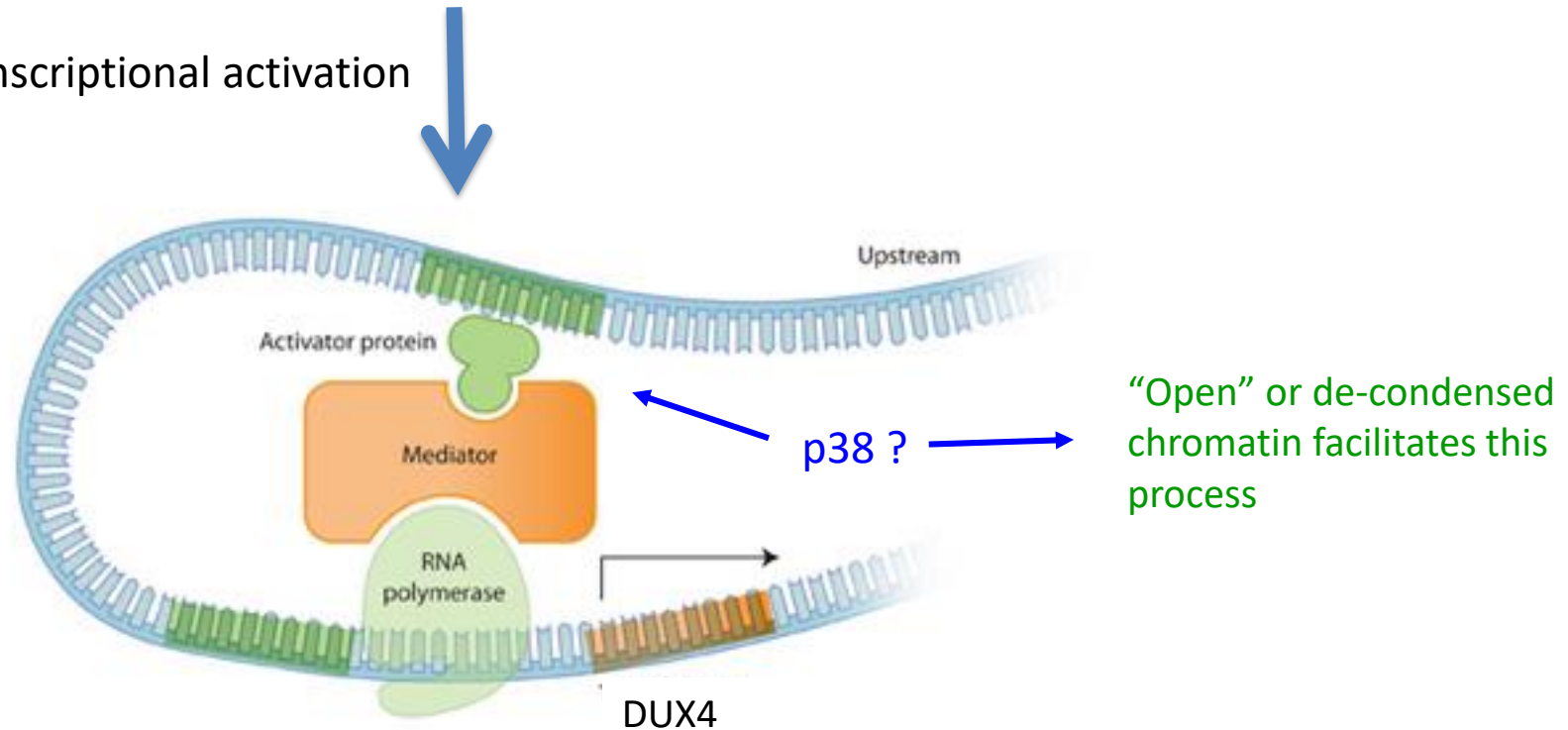
- Multi dose PK in Japan, PK consistent with single dose studies in US
- 7.5 mg BID

- Clinical dosing maintains narrow range of plasma levels
- Trough at FSHD myotube IC_{90}
- Trough above 14-day xenograph (23 nM)
- Muscle tissue levels approximately equal to plasma levels in mice

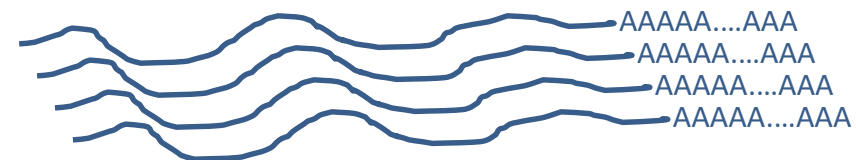




Transcriptional activation

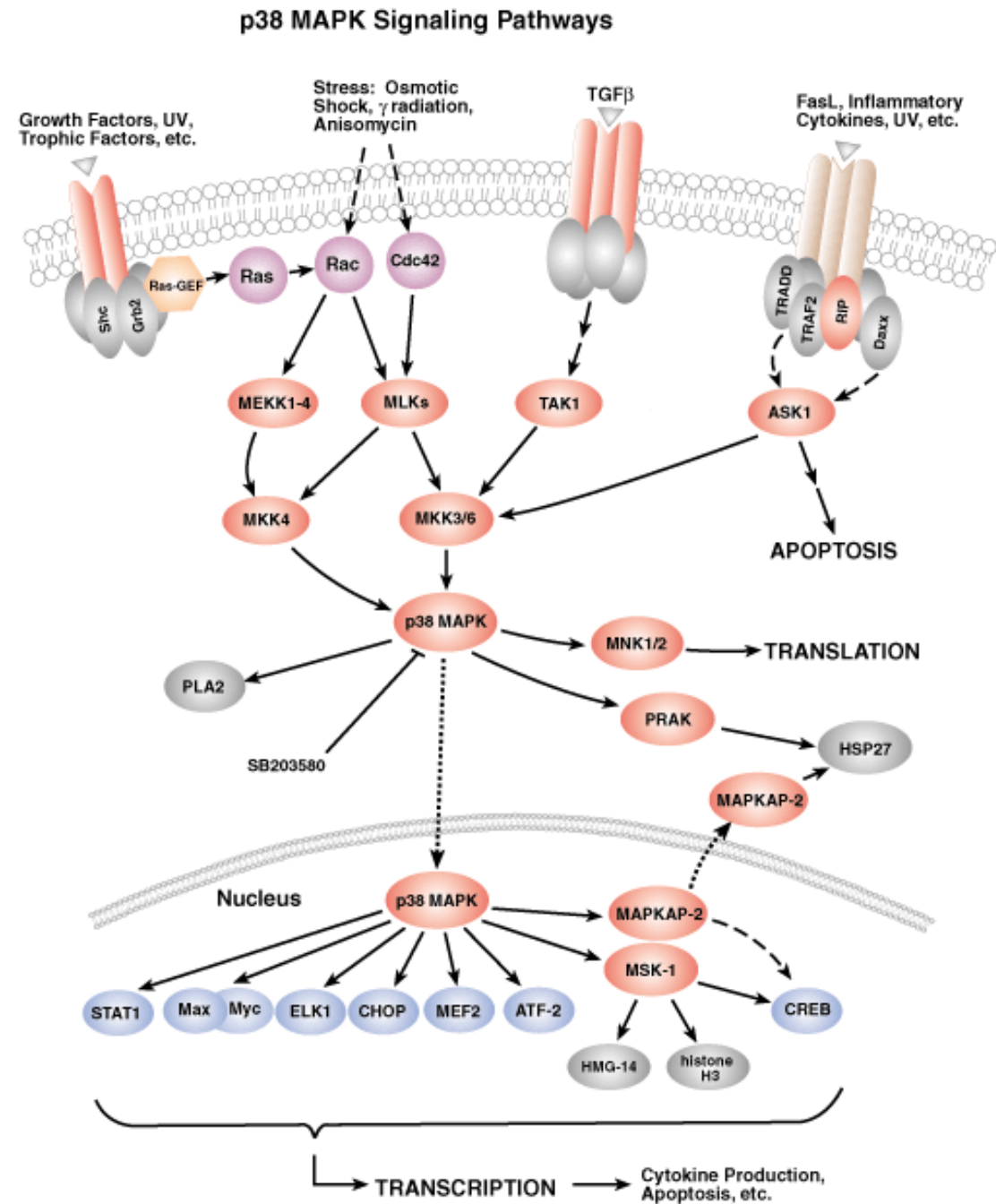


RNA copies of DUX4 gene sequence (DUX4 mRNA)

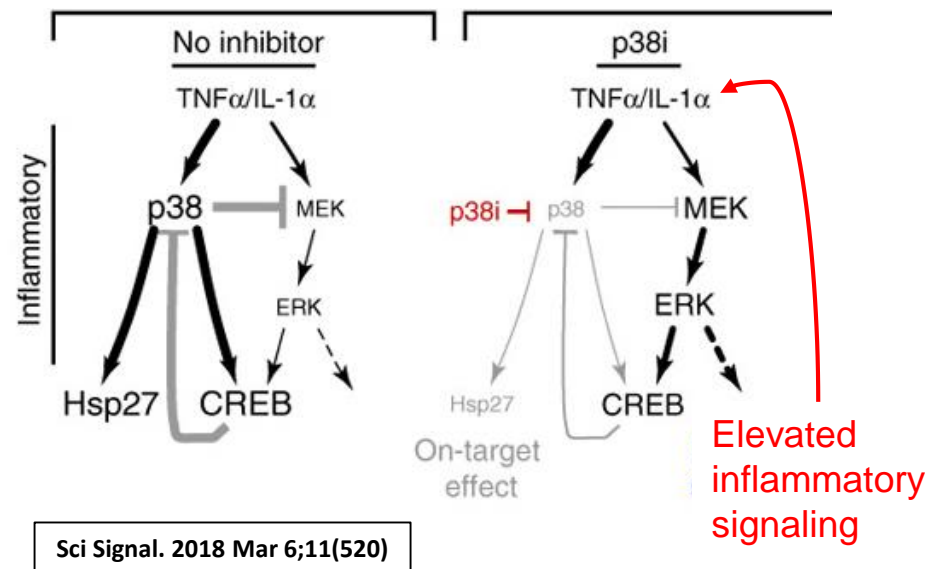
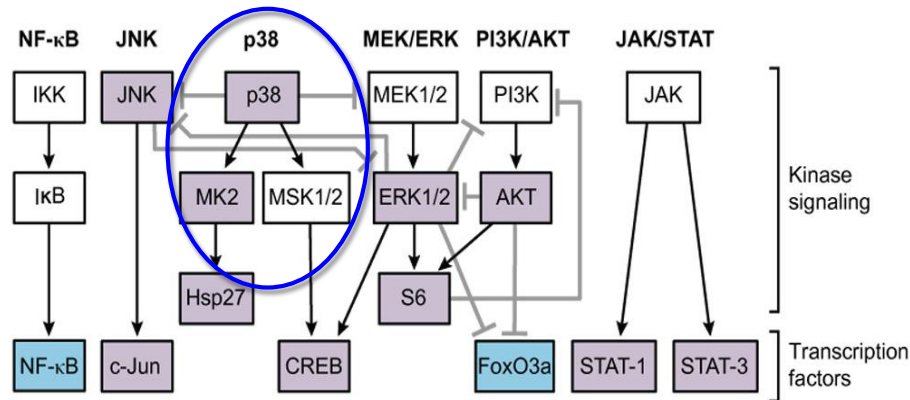


p38 MAP kinases

Inflammatory signaling



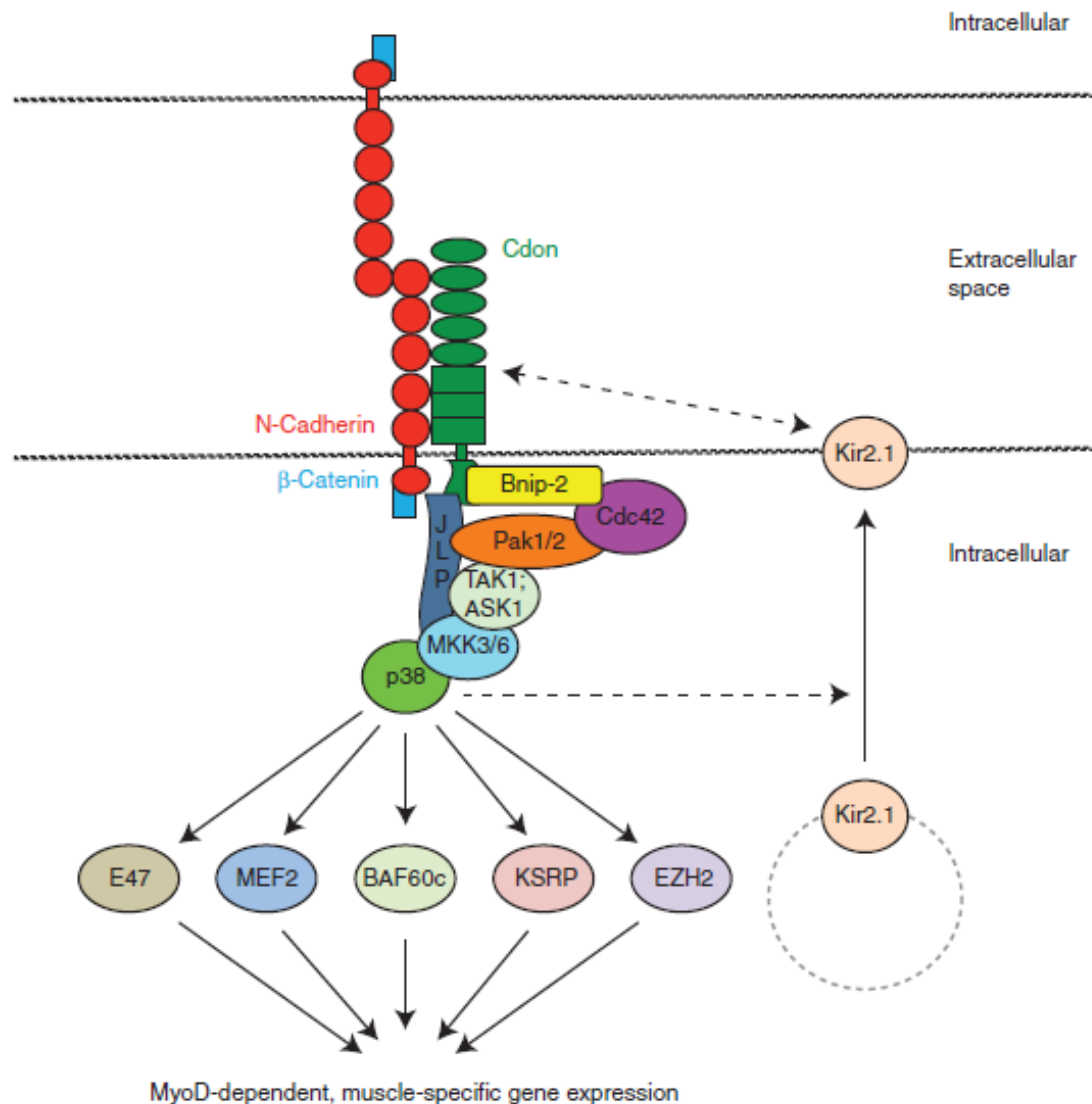
p38 pathway for inflammation is different than in muscle



- In rheumatoid arthritis synovial fibroblasts (SF), p38 activated by inflammatory stimuli
 - In muscle, p38 activated by normal differentiation program
- In SF, MK2 and MSK1/2 are key p38 targets that promotes inflammation
 - DUX4 expression not mediated by MK2 or MSK1/2
- In SF, inflammatory environment promotes alternate MEK/ERK signaling
 - Inflammatory cytokines do not stimulate DUX4 expression
 - DUX4 expression not mediated by MEK/ERK

p38 kinase

Activated by muscle differentiation



- Cell-cell contact activates p38 during the normal differentiation process
- NOT inflammatory signaling

Imaging and Biopsy: Clinical Trial Design Implications

Kathryn Wagner, MD, PhD
Center for Genetic Muscle Disorders
Kennedy Krieger Institute
Johns Hopkins School of Medicine

FSHD Challenges and Opportunities

- Challenges

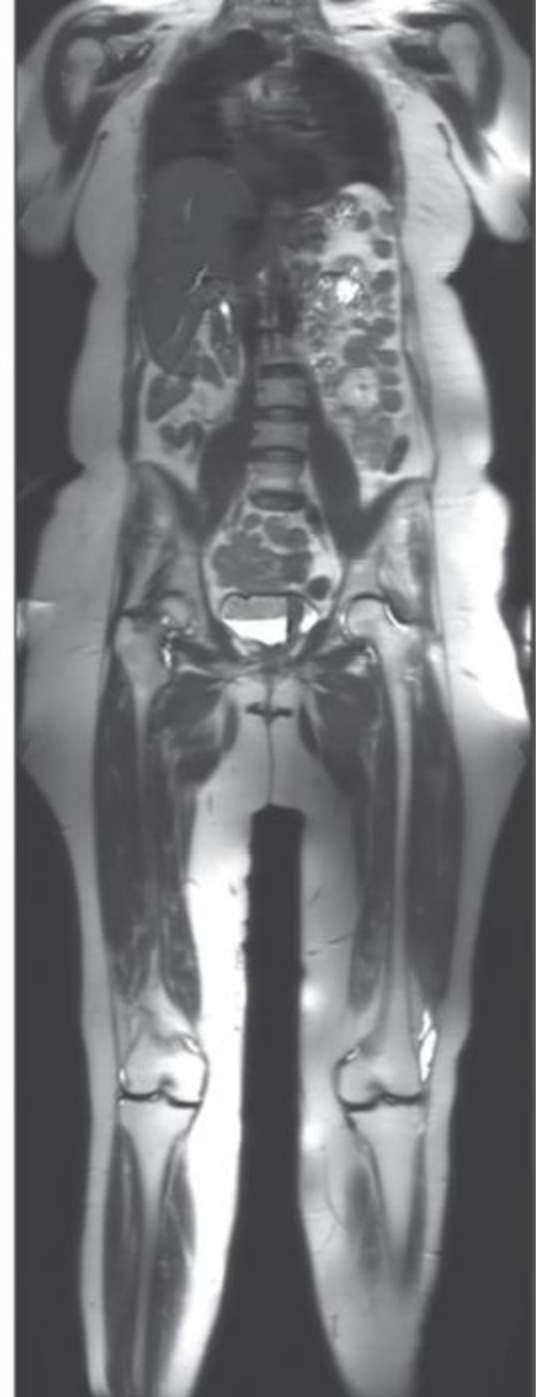
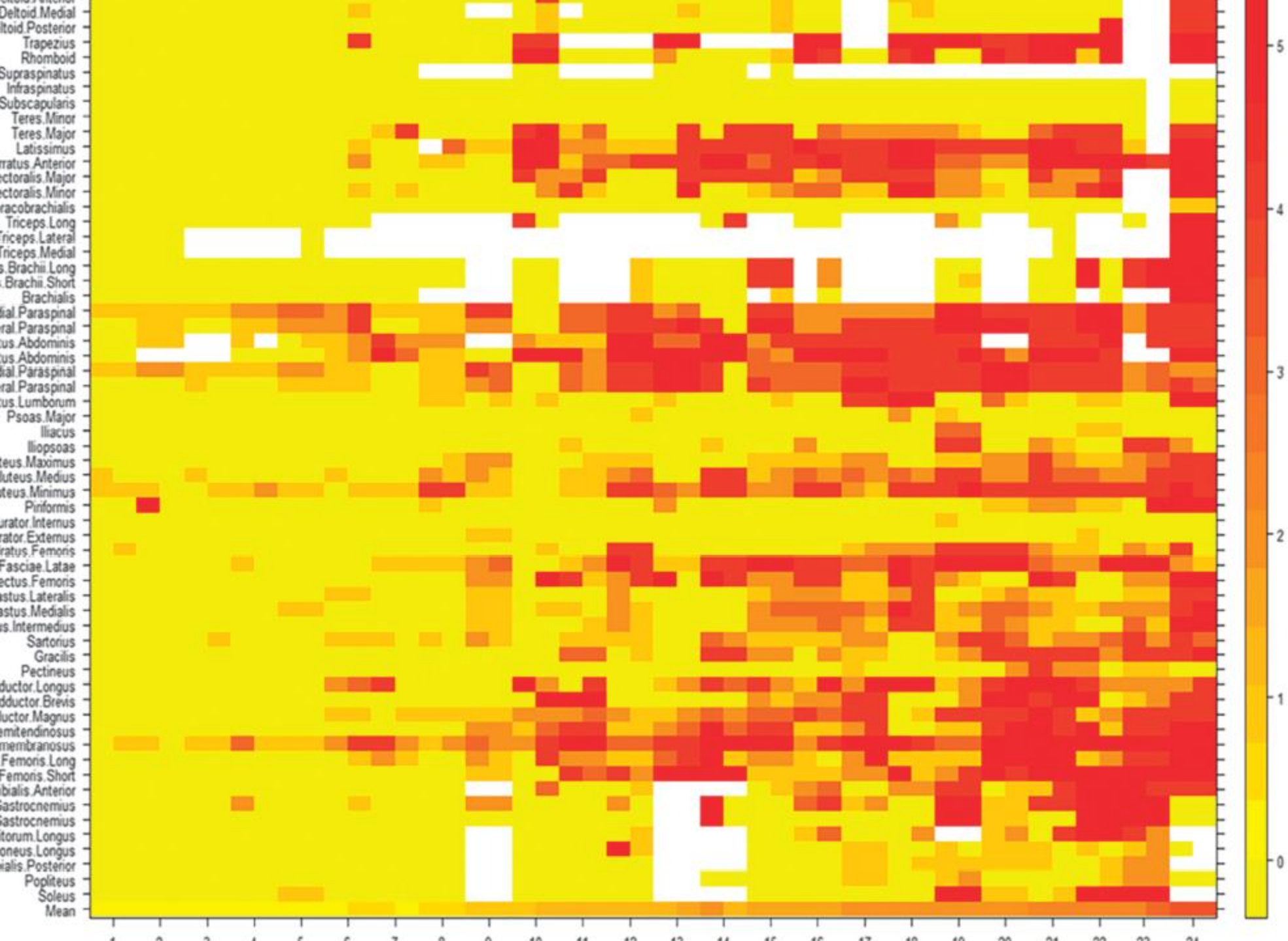
- Slowly progressing disorder
- Disease heterogeneity
- No established regulatory pathway

- Opportunities

- Common rare disease
- Highly motivated patient population
- Engaged, experienced investigator community
- Little competition (unlike DMD)

MRI

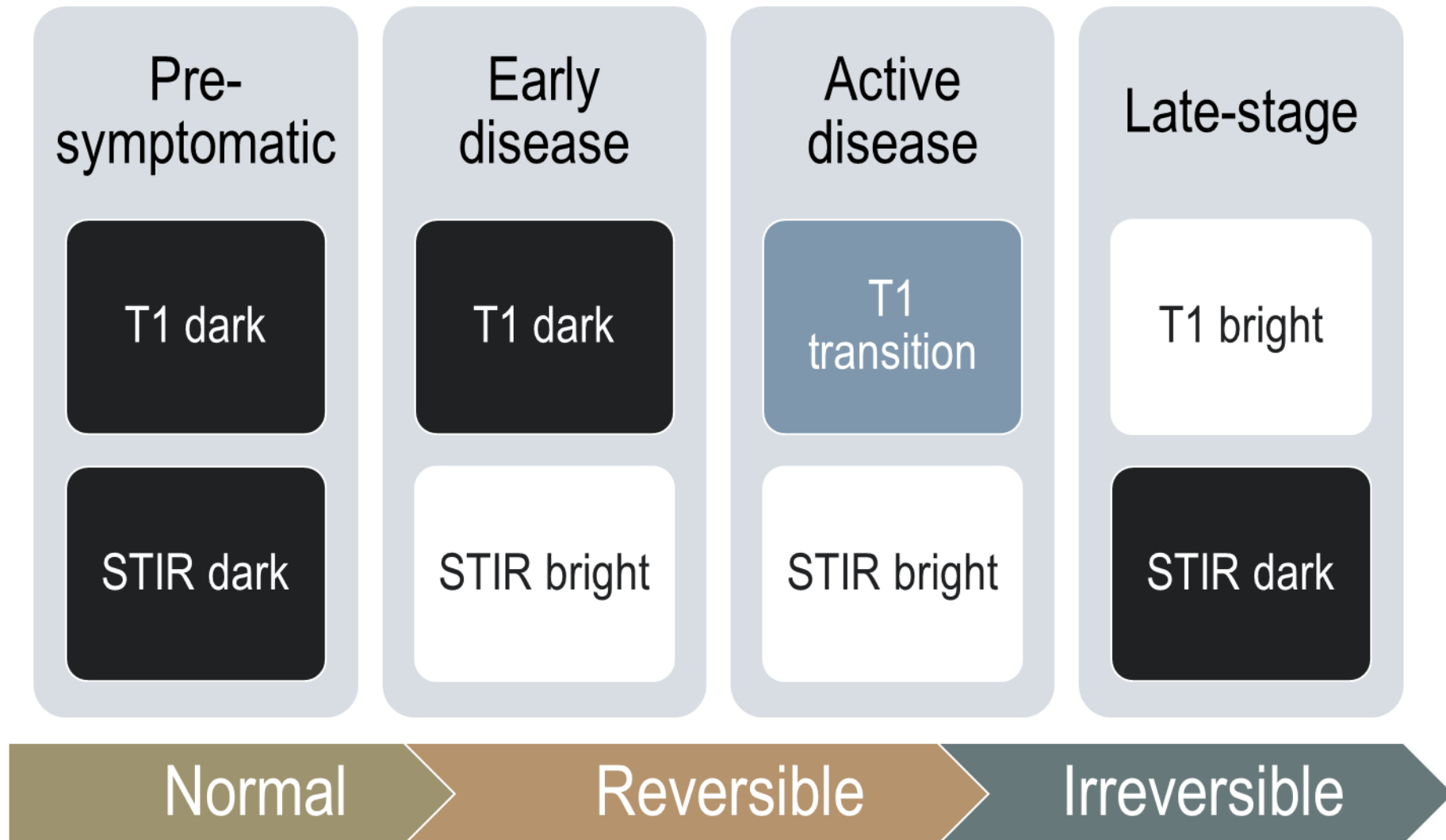
- Standard MRI sequence can identify healthy muscle, acute intramuscular inflammation and infiltration of fat and fibrosis
- Patterns of muscle involvement differentiate various genetic myopathies
- Distinctive pattern of muscle involvement in FSHD but not used as diagnostic due to superior specificity of genetic diagnosis
- Powerful clinical outcome measure
 - Noninvasive
 - Nonirradiating
 - Independent of patient effort, daily clinical variability and learning effects
 - Can be performed on most patients irrespective of disease severity
 - Sensitive to small increments of change
 - Repeatable measurements



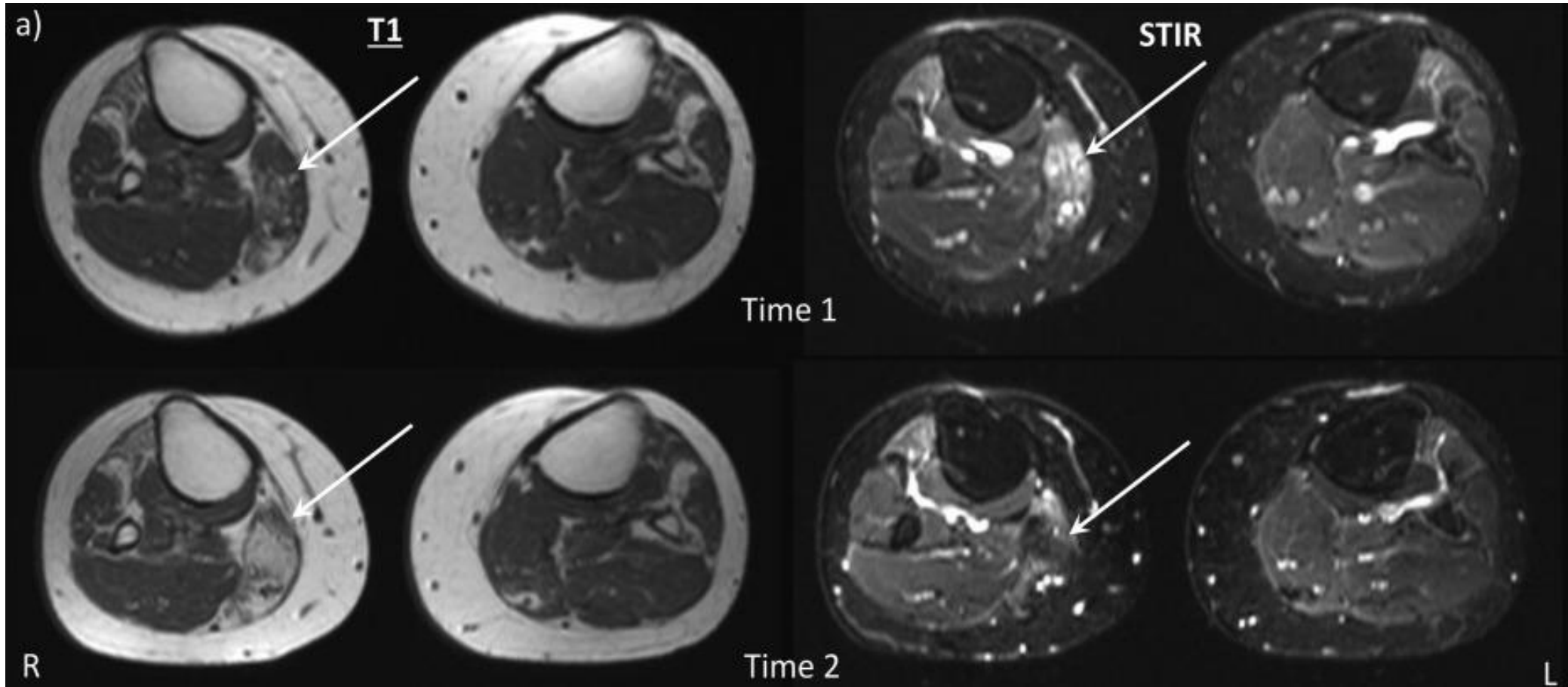
MRI

- Longitudinal studies confirm slowly progressive disorder
- Fatty infiltration does not progress over 6.9 to 13.8 months of follow up
- Fatty infiltration seen as T1-weighted hyperintensity
- Hyperintensity on STIR (Short-T1 Inversion Recovery) sequences correlate with edema and cellular inflammation
- STIR positivity may precede fatty replacement of T1-weight hyperintensity

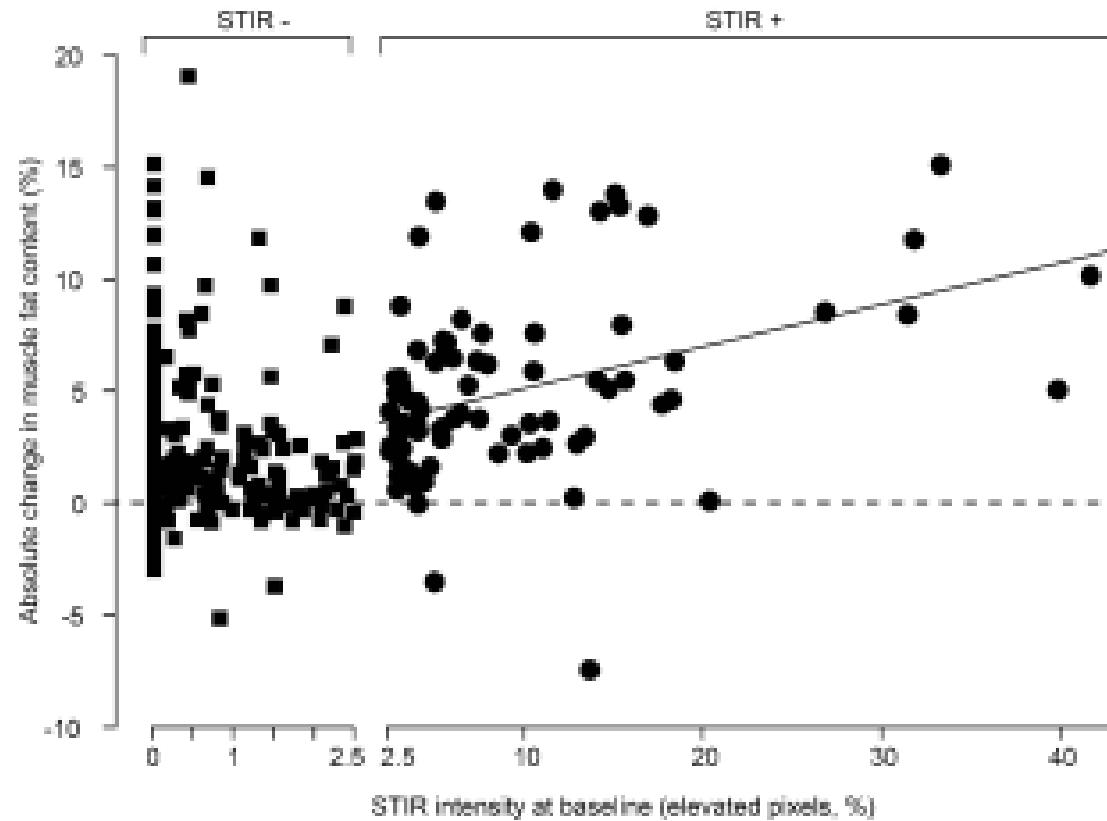
Theory of disease progression in FSHD



STIR Hyperintensity and Fat infiltration

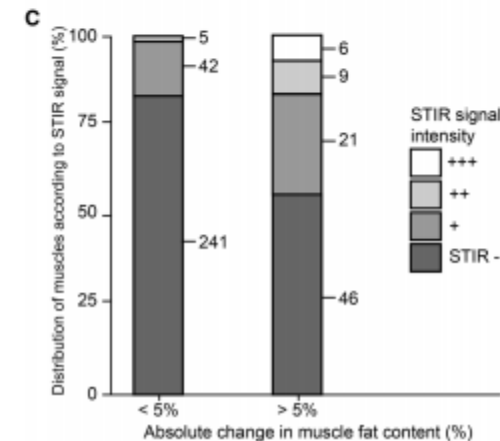
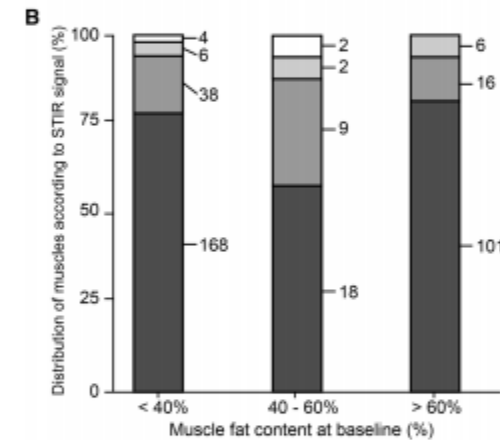
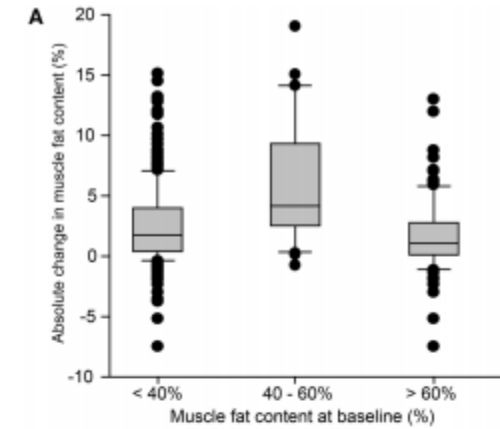


Relationship between STIR+ and fat

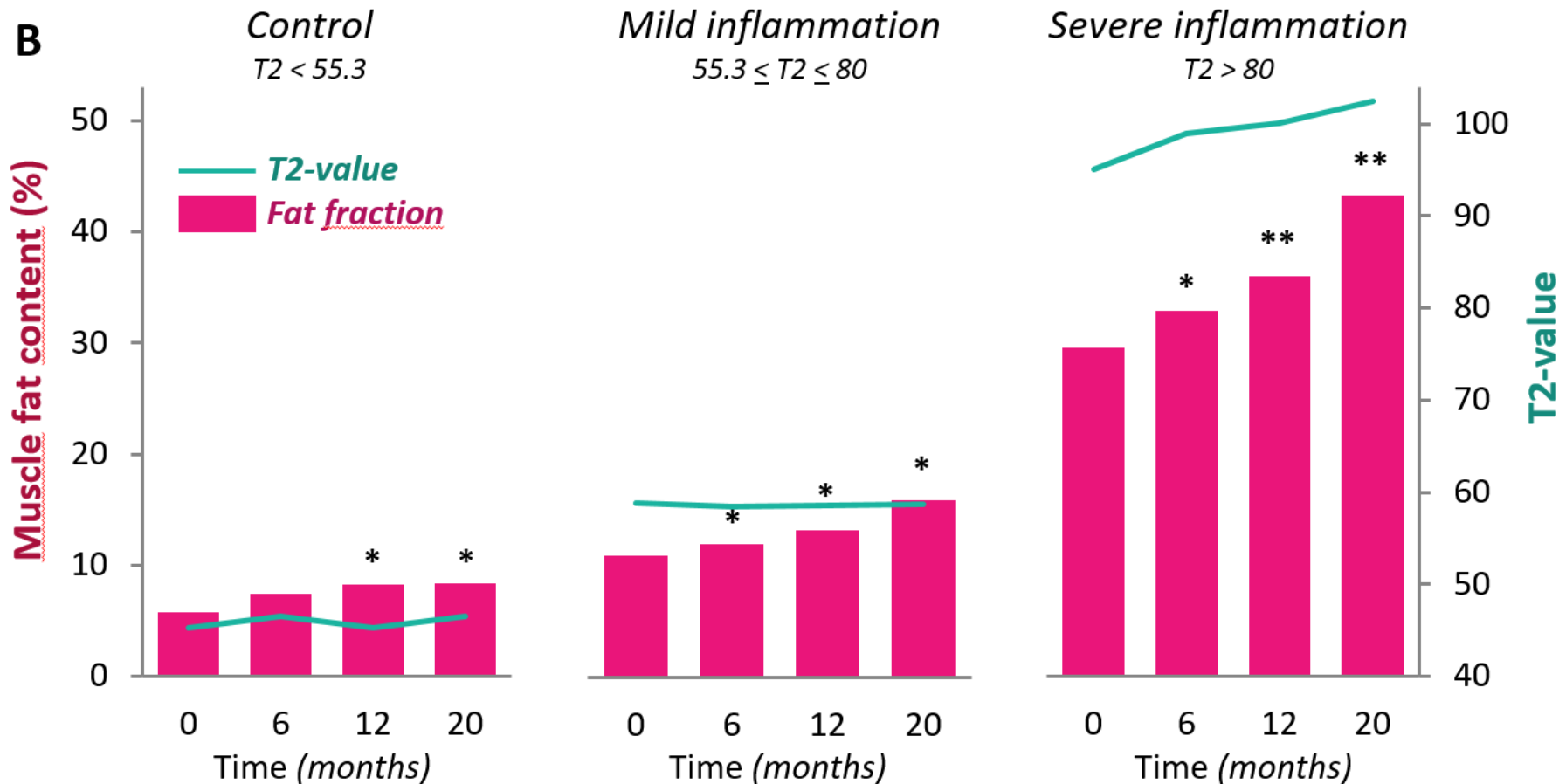


Fat replacement progression

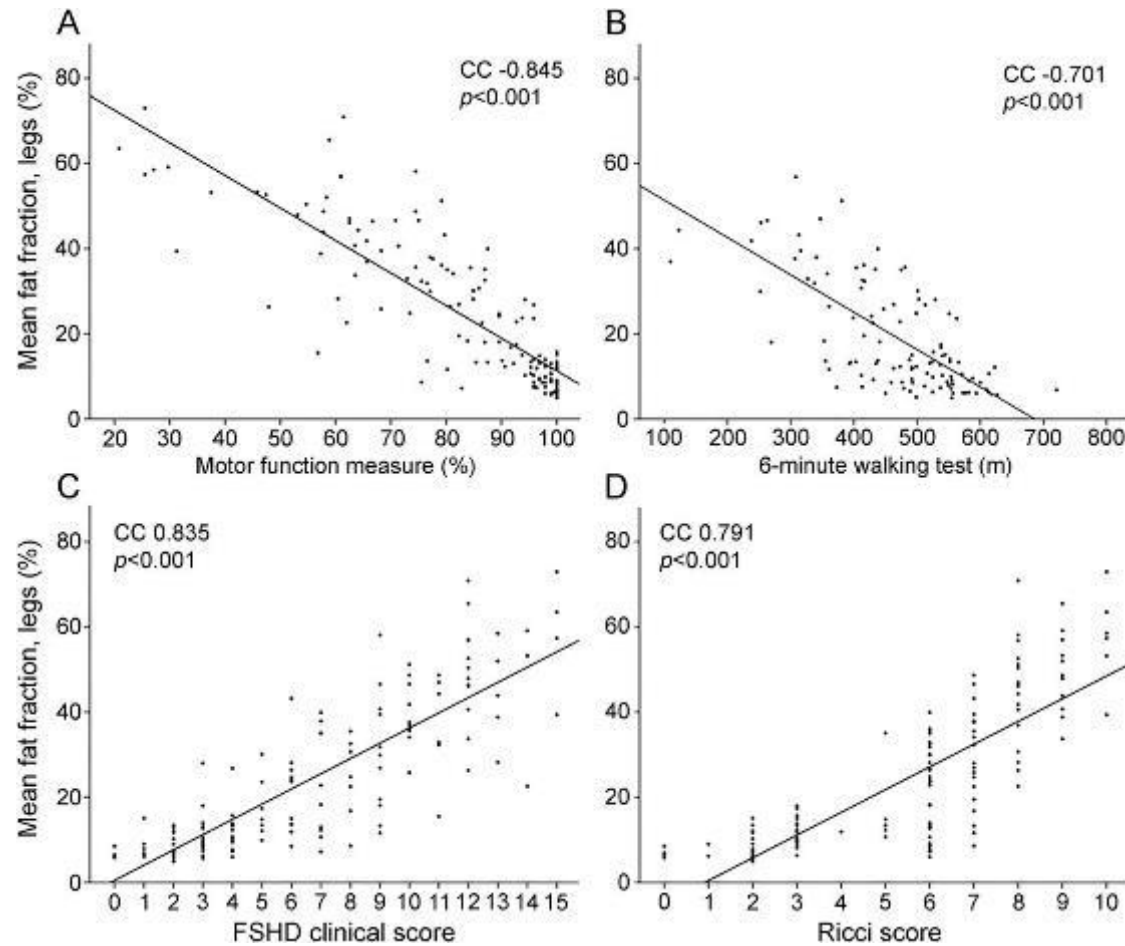
- Intermediate fat replaced muscles had greatest change over time
- Highest percentage of STIR+ muscles were among intermediate fat replaced muscles
- Stir+ muscle had a faster progression of fat replacement



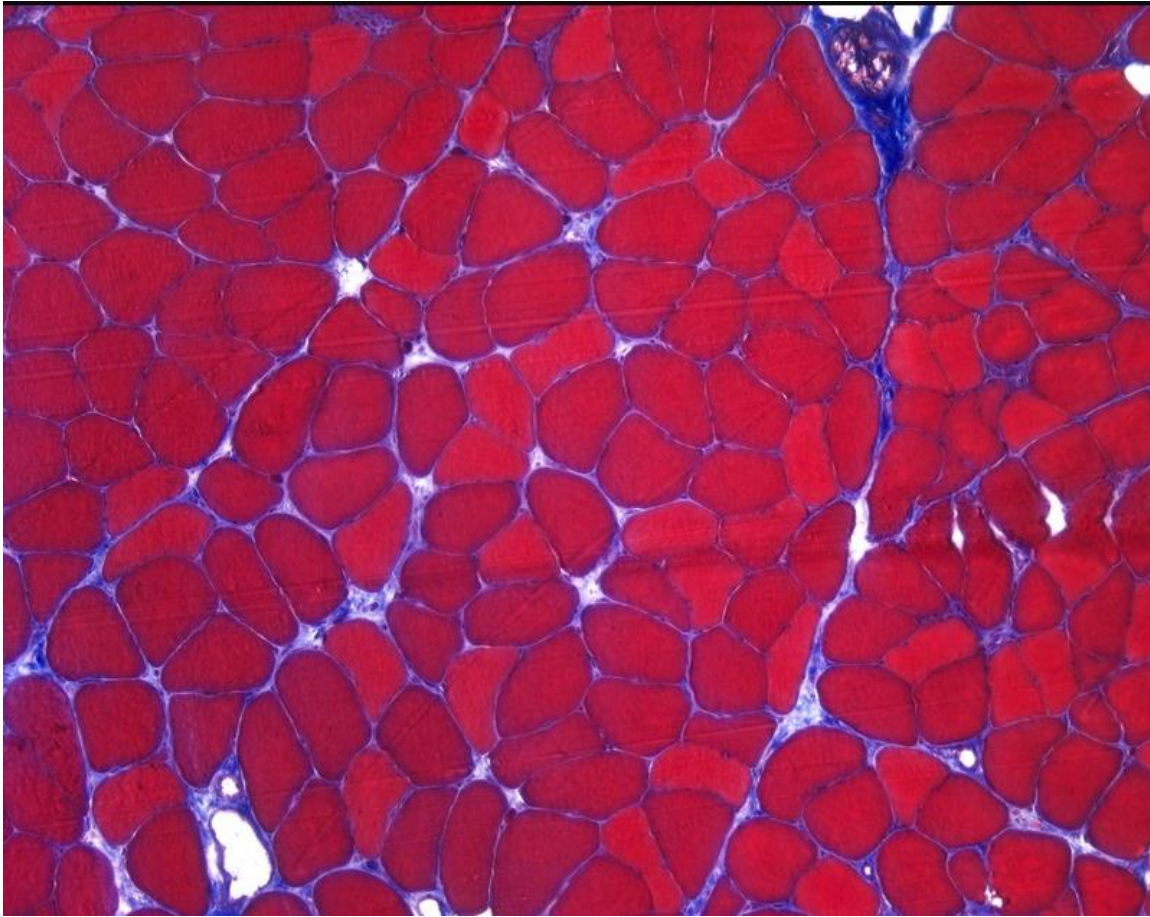
More severe progression with greater T2 signal abnormalities



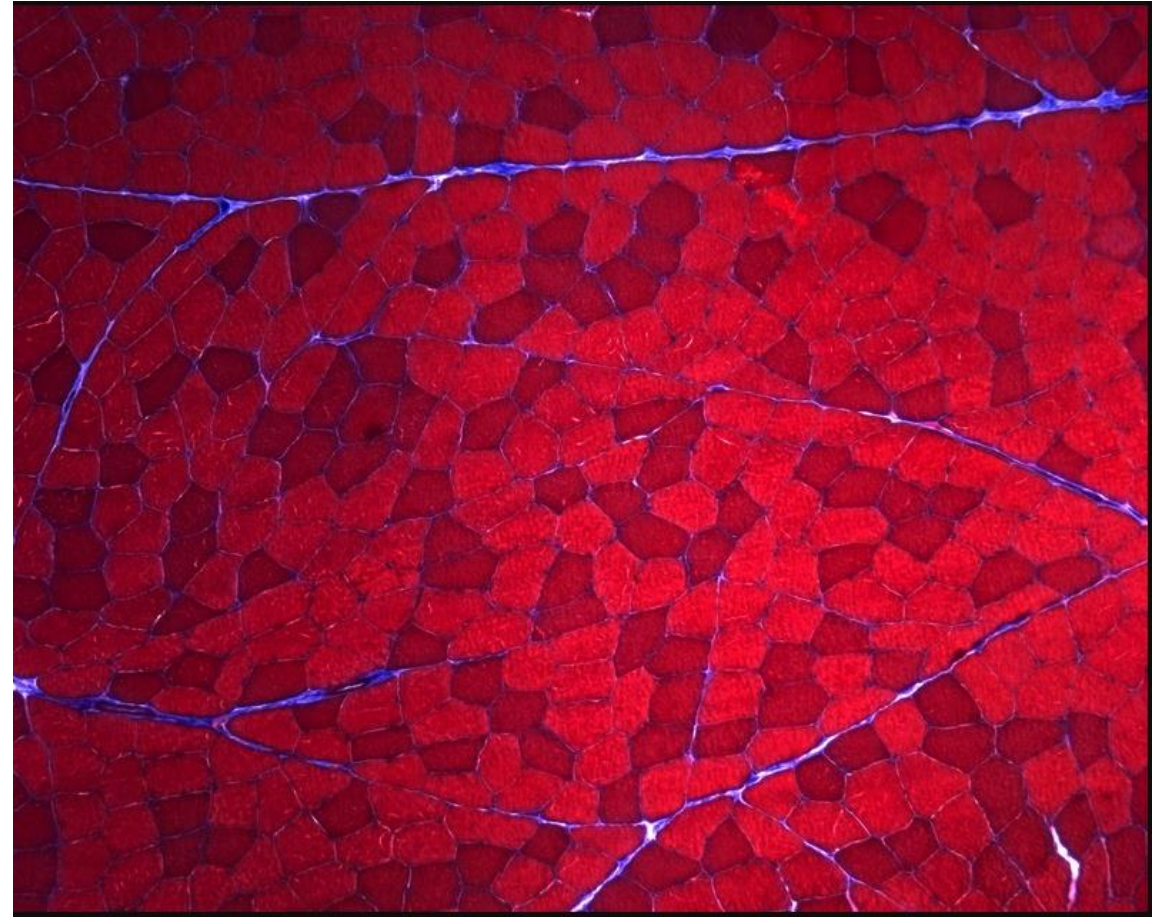
Good correlation of fat fraction to function



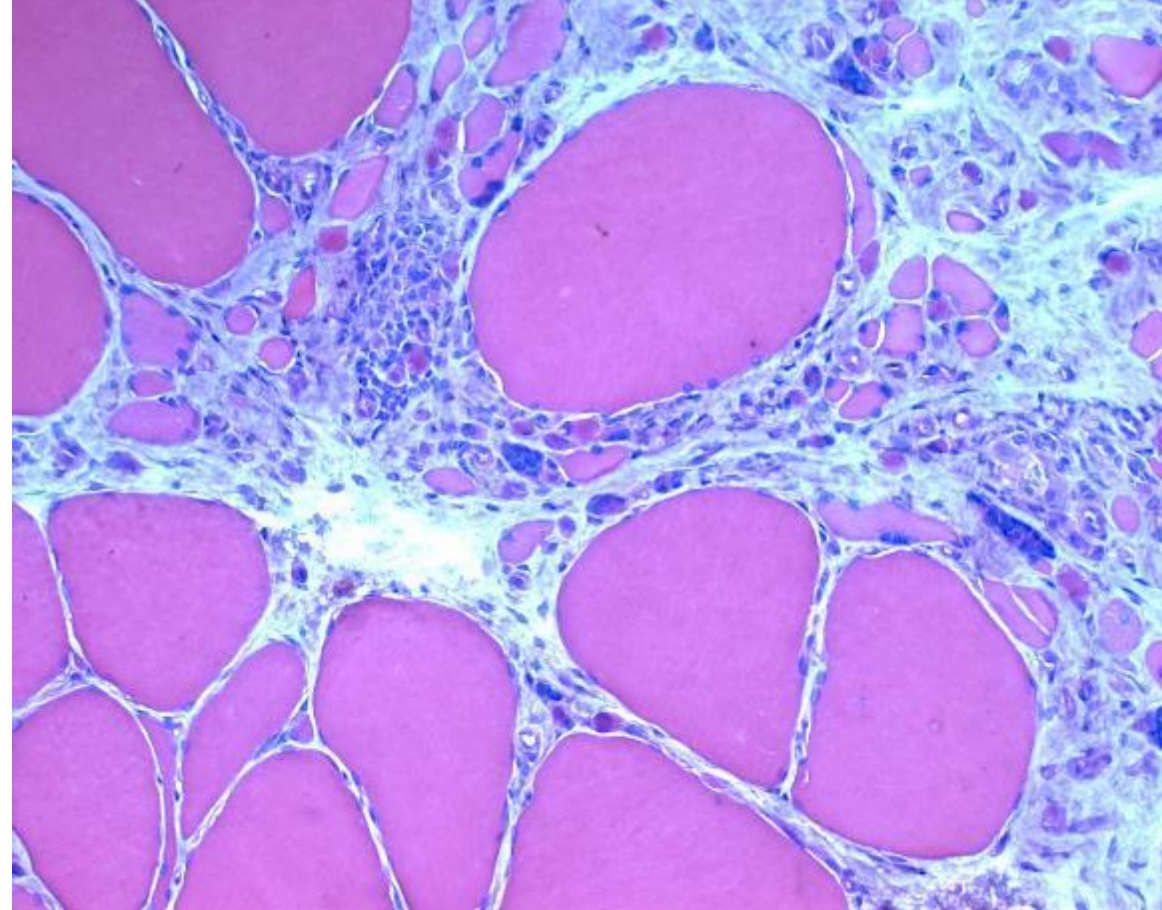
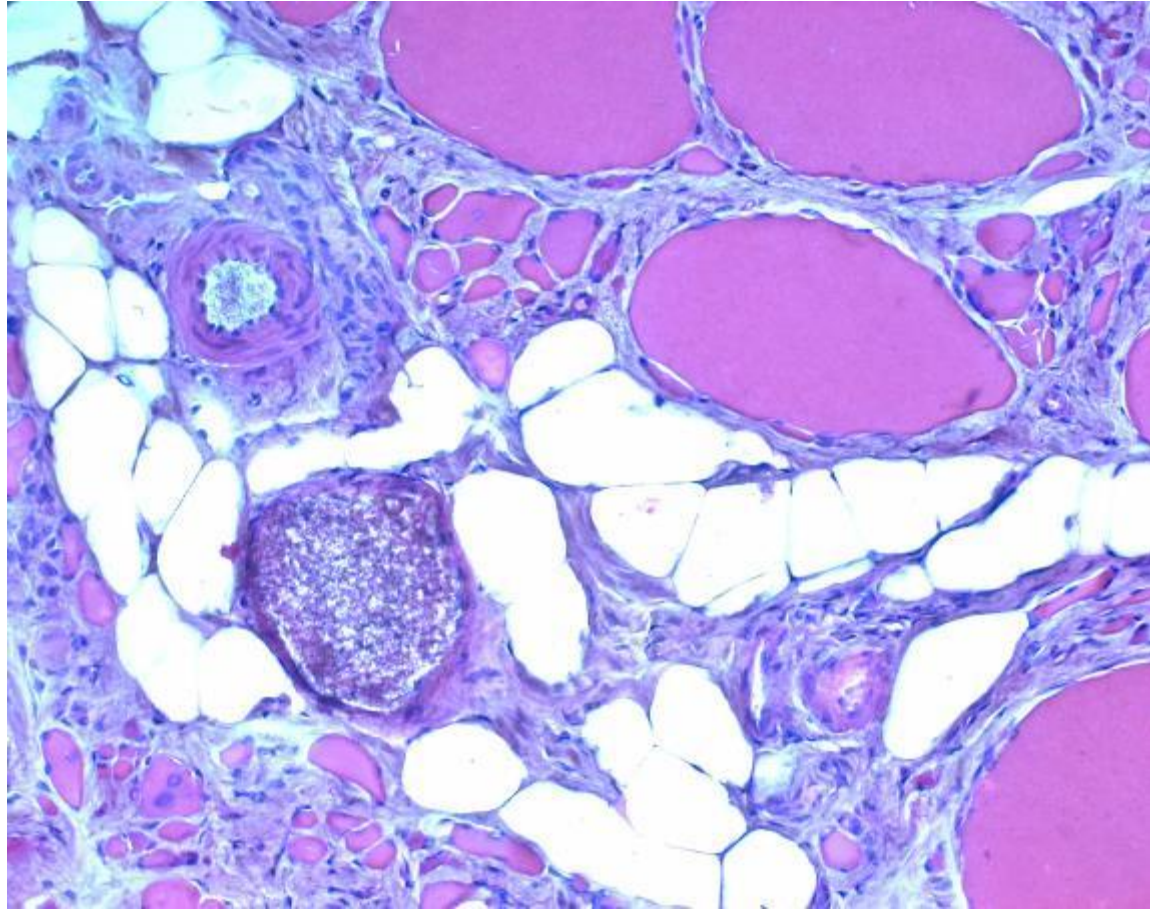
Muscle biopsy



FSHD



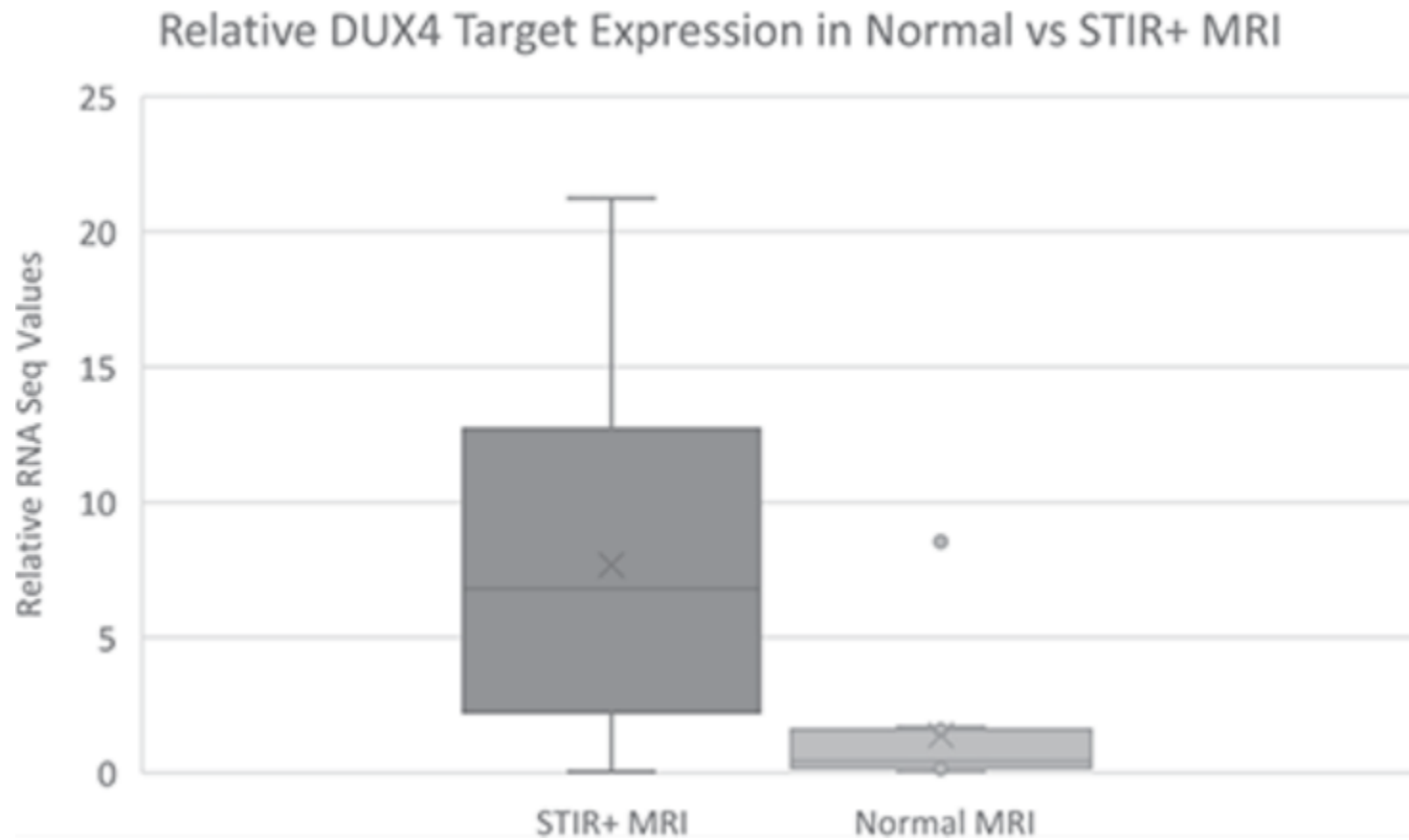
Control



Pathological features

| | FSHD | | Control | | p-value |
|-------------------------|-------------|------------------|-------------|------------------|-----------|
| | <u>Mean</u> | <u>Std. Dev.</u> | <u>Mean</u> | <u>Std. Dev.</u> | |
| Int. Nuclei (%) | 0.028339 | 0.035156 | 0.015608 | 0.016162 | 0.03514 |
| Split Fibers (%) | 0.001185 | 0.002627 | 0.000385 | 0.00118 | 0.073456 |
| Necrotic Fibers (%) | 0.001356 | 0.002417 | 3.35E-05 | 0.000167 | 0.000334 |
| Regenerating Fibers (%) | 0.004628 | 0.010109 | 0.00016742 | 0.00059641 | 0.0125897 |
| Atrophic Fibers (%) | 0.023216 | 0.061632 | 0.002 | 0.002832 | 0.018824 |
| Area of Fibrosis (%) | 0.1 | 0.0396 | 0.07 | 0.0229 | 0.0415 |

Image guided biopsy in FSHD



Wang, *Human Molecular Genetics*, 2018

Open muscle biopsy



<http://vetneuromuscular.ucsd.edu/>

Needle muscle biopsy



Needle muscle biopsy



Open vs Fine Needle Biopsy

| OPEN | FINE NEEDLE |
|---|--|
| Requires OR | Can be done in clinic |
| Leaves incision scar | Puncture scar or none |
| Nerves, vessels, muscle integrity well visualized | Blind |
| Well tolerated in most | Well tolerated in all |
| Hundreds of micrograms | Tens of micrograms |
| Hundreds of fibers | Few fibers |
| Well-oriented fibers for histology | Misaligned fibers not suitable for histology |
| \$\$\$\$\$ | \$ |
| Best for when histology needed | Best for when only RNA/protein analysis needed or when serial evaluations needed |

Other clinical outcome measures

- ReSOLVE natural history study: 160 FSHD subjects across multiple US and EU sites
- Goal to identify clinical outcomes that are more responsive to change over shorter periods of time for drug development trials
- Reachable workspace (Hatch et al., Neuromuscular Disord 2019: ~8% decline per year in upper quadrants)
- EIM
- FSHD-COM
- FSHD-HI

Conclusions

- Muscle progresses from healthy muscle to fatty infiltration
- Muscle inflammation may act as a trigger for this process
- Muscle inflammation can be visualized by STIR positivity
- DUX4 expression has been linked to STIR+ muscles
- DUX4 and DUX4 biomarkers can be assessed by needle muscle biopsy
- For a very slowly progressive disease MRI and biopsy are good outcome measures